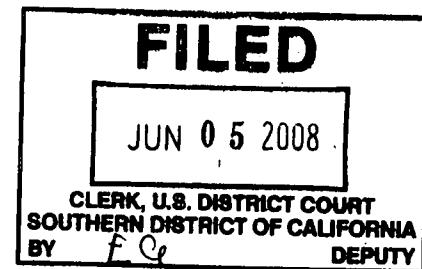


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26 UNITED STATES DISTRICT COURT

27 FOR THE SOUTHERN DISTRICT OF CALIFORNIA

28 BECKMAN COULTER INC., a Delaware  
 29 corporation, and ORCHID CELLMARK INC.,  
 30 a Delaware corporation,

31 Plaintiffs,

32 v.

33 SEQUENOM, INC., a Delaware corporation,

34 Defendant.

Case No.

35 '08 CV 1013 W POR

36 COMPLAINT FOR PATENT  
 37 INFRINGEMENT

38 ORIGINAL

Beckman Coulter Inc. and Orchid Cellmark Inc. file this Complaint and demand for jury trial seeking relief for patent infringement by the Defendant, Sequenom, Inc., stating and alleging the following:

## THE PARTIES

1. Beckman Coulter Inc. ("BCI") is a corporation organized and existing under the laws of the State of Delaware with its principal place of business at 4300 North Harbor Boulevard, Fullerton, California 92834-3100.

2. Orchid Cellmark Inc. ("OCI") is a corporation organized and existing under the laws of the State of Delaware with its principal place of business at 4390 U.S. Route One, Princeton, New Jersey 08540.

3. On information and belief, Sequenom, Inc. ("Sequenom") is a corporation organized and existing under the laws of the State of Delaware with its principal place of business at 3595 John Hopkins Court, San Diego, California 92121.

## **JURISDICTION AND VENUE**

4. This is a civil action for patent infringement arising under the patent laws of the United States, 35 U.S.C. § 101, *et seq.* This Court has subject matter jurisdiction pursuant to 28 U.S.C. §§ 1331 and 1338. Venue is proper in this Federal District pursuant to 28 U.S.C. §§ 1391 and 1400(b) as Defendant Sequenom has its principal place of business in this District, has committed acts of infringement in this District, and continues to commit acts of infringement in this District.

## BACKGROUND

5. BCI is a leading manufacturer of biomedical testing instrument systems, tests and supplies that simplify and automate laboratory processes. Spanning the biomedical testing continuum — from pioneering medical research and clinical trials to laboratory diagnostics and point-of-care testing — BCI's systems provide essential biomedical information to enhance healthcare around the world. One of BCI's advanced systems is its GenomeLab™ SNPstream® Genotyping System. The GenomeLab SNPstream Genotyping System is an integrated system of advanced instrumentation, software and reagents and provides an automated scalable system capable of performing from 4,600 to

1 over 3,000,000 single nucleotide polymorphism ("SNP") genotypes per day. BCI has an exclusive  
 2 licensee to certain patents owned by OCI, including patents that cover BCI's GenomeLab SNPstream  
 3 Genotyping System.

4       6. OCI owns many U.S. patents, including United States Patent No. 5,888,819 ("the '819  
 5 patent"), United States Patent No. 6,004,744 ("the '744 patent"), and United States Patent  
 6 No. 6,537,748 ("the '748 patent"). The '819 patent is entitled "Method for Determining Nucleotide  
 7 Identity Through Primer Extension" and was duly and legally issued by the United States Patent and  
 8 Trademark Office on March 30, 1999. A true and correct copy of the '819 patent is attached hereto as  
 9 Exhibit A. The '744 patent is entitled "Method for Determining Nucleotide Identity Through  
 10 Extension of Immobilized Primer" and was duly and legally issued by the United States Patent and  
 11 Trademark Office on December 21, 1999. A true and correct copy of the '744 patent is attached as  
 12 Exhibit B. The '748 patent is entitled "Reagent for Nucleic Acid Typing by Primer Extension" and  
 13 was duly and legally issued by the United States Patent and Trademark Office on March 25, 2003. A  
 14 true and correct copy of the '748 patent is attached as Exhibit C. The '819 patent covers methods of  
 15 determining the identity of a single nucleotide base at a specific position using at least one terminator  
 16 labeled with a detectable marker and the '744 and '748 patents cover reagents capable of terminating  
 17 an extension reaction wherein at least one terminator is labeled with a detectable marker.

18       7. BCI and OCI (collectively "Beckman") entered into a Patent License Agreement  
 19 effective December 19, 2002 (the "Beckman Agreement"). The Beckman Agreement grants to BCI  
 20 an exclusive license to certain patents within particular fields. Among the patents licensed to BCI  
 21 under the Beckman Agreement are the '819, '774 and '748 patents. BCI marks its GenomeLab  
 22 SNPstream Genotyping System with the '819 patent and '744 patent numbers. BCI's rights under the  
 23 Beckman Agreement include the right to bring suit against entities infringing the OCI patents within  
 24 the fields exclusively licensed to BCI.

25       8. On information and belief, Sequenom is a competitor of BCI in the biomedical testing  
 26 field. Sequenom makes, uses, sells, offers for sale and/or imports into the U.S. and teaches the use of  
 27 its MassARRAY™ iPLEX™ products for SNP genotyping applications. Sequenom's iPLEX

1 products use single nucleotide base extension technology using detectable markers to identify SNPs.  
 2 The methods taught and the products sold by Sequenom to identify SNPs with detectable markers  
 3 infringe, induce infringement of, and contributorily infringe the '819 patent, the '774 patent and the  
 4 '748 patent. Sequenom has knowledge of its continued infringement of the '819 patent and the  
 5 '744 patent, at least through letters addressed to Dereck Tatman and Steven Weiss of Sequenom on or  
 6 about January 2, 2008, and April 15, 2008.

7 **COUNT I**

8 **(Patent Infringement of U.S. Patent No. 5,888,819)**

9 9. Beckman realleges paragraphs 1-8 of its complaint.

10 10. In violation of 35 U.S.C. § 271(a), Sequenom has infringed and continues to infringe  
 11 the '819 patent by using the methods and by making, using, selling, offering for sale and/or importing  
 12 products for use in the methods described and claimed in the '819 patent within the United States and  
 13 this District.

14 11. In violation of 35 U.S.C. § 271(b), Sequenom has actively induced infringement and  
 15 continues to actively induce infringement of the '819 patent by selling, offering for sale and/or  
 16 importing products for use in the methods and by teaching the use of the products in the methods  
 17 described and claimed in the '819 patent within the United States and this District.

18 12. In violation of 35 U.S.C. § 271(c), Sequenom has contributorily infringed and  
 19 continues to contributorily infringe the '819 patent by selling, offering for sale and/or importing  
 20 material for use in the methods described and claimed in the '819 patent, knowing such material to be  
 21 especially made or adapted for use in an infringement of the '819 patent, within the United States and  
 22 this District.

23 13. Despite Sequenom's knowledge of its infringement of the '819 patent, Sequenom  
 24 continues to infringe. Sequenom's infringement of the '819 patent is willful and deliberate, making  
 25 this case exceptional and entitling Beckman to increased damages under 35 U.S.C. § 284 and to  
 26 attorneys' fees and costs incurred in prosecuting this action under 35 U.S.C. § 285.

14. Sequenom has caused, and will continue to cause Beckman damages, including lost profits, by infringing the '819 patent in an amount to be proven at trial.

## COUNT II

**(Patent Infringement of U.S. Patent No. 6,004,744)**

15. Beckman realleges paragraphs 1-8 and 10-14 of its Complaint.

16. In violation of 35 U.S.C. § 271(a), Sequenom has infringed and continues to infringe the '744 patent by making, using, selling, offering for sale and/or importing products described and claimed in the '744 patent within the United States and this District.

17. In violation of 35 U.S.C. § 271(b), Sequenom has actively induced infringement and continues to actively induce infringement of the '744 patent by selling, offering for sale and/or importing products and by teaching the use of products described and claimed in the '744 patent within the United States and this District.

18. In violation of 35 U.S.C. § 271(c), Sequenom has contributorily infringed and continues to contributorily infringe the '744 patent by selling, offering for sale and/or importing component products of the composition described and claimed in the '744 patent, knowing such component products to be especially made or adapted for use in an infringement of the '744 patent, within the United States and this District.

19. Despite Sequenom's knowledge of its infringement of the '744 patent, Sequenom continues to infringe. Sequenom's infringement of the '744 patent is willful and deliberate, making this case exceptional and entitling Beckman to increased damages under 35 U.S.C. § 284 and to attorneys' fees and costs incurred in prosecuting this action under 35 U.S.C. § 285.

20. Sequenom has caused, and will continue to cause Beckman damages, including lost profits, by infringing the '744 patent in an amount to be proven at trial.

**(Patent Infringement of U.S. Patent No. 6,537,748)**

21. Beckman realleges paragraphs 1-8, 10-14, and 16-20 of its Complaint.

22. In violation of 35 U.S.C. § 271(a), Sequenom has infringed and continues to infringe the '748 patent by making, using, selling, offering for sale and/or importing products described and claimed in the '748 patent within the United States and this District.

23. In violation of 35 U.S.C. § 271(b), Sequenom has actively induced infringement and continues to actively induce infringement of the '748 patent by selling, offering for sale and/or importing products and by teaching the use of products described and claimed in the '748 patent within the United States and this District.

24. In violation of 35 U.S.C. § 271(c), Sequenom has contributorily infringed and continues to contributorily infringe the '748 patent by selling, offering for sale and/or importing component products of the composition described and claimed in the '748 patent, knowing such component products to be especially made or adapted for use in an infringement of the '748 patent, within the United States and this District.

25. Despite Sequenom's knowledge of its infringement of the '748 patent, Sequenom continues to infringe. Sequenom's infringement of the '748 patent is willful and deliberate, making this case exceptional and entitling Beckman to increased damages under 35 U.S.C. § 284 and to attorneys' fees and costs incurred in prosecuting this action under 35 U.S.C. § 285.

26. Sequenom has caused, and will continue to cause Beckman damages, including lost profits, by infringing the '748 patent in an amount to be proven at trial.

## PRAYER FOR RELIEF

WHEREFORE, Plaintiffs Beckman Coulter Inc. (“BCI”) and Orchid Cellmark Inc. (“OCI”) request entry of judgment in their favor and against Defendant Sequenom, Inc. (“Sequenom”) as to each count as follows:

- (a) Judgment that Sequenom has infringed and continues to infringe claims of U.S. Patent Numbers 5,888,819, 6,004,744, and 6,537,748;
- (b) An order permanently enjoining Sequenom and its respective officers, agents, employees, and those acting in privity with them, from further infringement, inducing

1 infringement and/or contributory infringement of U.S. Patent Numbers 5,888,819,  
 2 6,004,744, and 6,537,748;

3 (c) An award of damages to BCI and OCI arising out of Sequenom's infringement of U.S.  
 4 Patent Numbers 5,888,819, 6,004,744, and 6,537,748, including lost profits and  
 5 enhanced damages pursuant to 35 U.S.C. § 284, in an amount according to proof and  
 6 no less than a reasonable royalty;

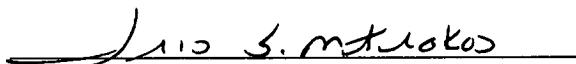
7 (d) An order, pursuant to 35 U.S.C. § 285, declaring this case to be "exceptional" and  
 8 awarding BCI and OCI their reasonable attorneys' fees in connection with this action;

9 (e) An order awarding costs of this action to BCI and OCI;

10 (f) An award to BCI and OCI of such other and further relief as this Court deems just and  
 11 proper.

12 DATED: June 5, 2008

TOWNSEND AND TOWNSEND AND CREW LLP

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 25 ORCHID CELLMARK INC.

26 61391863 v1





US005888819A

**United States Patent** [19]  
**Goelet et al.**

[11] Patent Number: **5,888,819**  
[45] Date of Patent: **Mar. 30, 1999**

[54] **METHOD FOR DETERMINING NUCLEOTIDE IDENTITY THROUGH PRIMER EXTENSION**

[75] Inventors: **Phillip Goelet**, Cockeysville; **Michael R. Knapp**, Baltimore, both of Md.; **Stephen Anderson**, Princeton, N.J.

[73] Assignee: **Molecular Tool, Inc.**, Baltimore, Md.

[21] Appl. No.: **664,837**

[22] Filed: **Mar. 5, 1991**

[51] Int. Cl.<sup>6</sup> ..... **C12Q 1/70**; **C12Q 1/68**;  
**C12P 19/34**

[52] U.S. Cl. ..... **435/5**; **435/6**; **435/91.2**

[58] Field of Search ..... **435/5, 91, 6, 91.2**;  
**935/76, 77, 78**

[56] **References Cited**

U.S. PATENT DOCUMENTS

4,656,127 4/1987 Mundy ..... 435/6  
4,962,020 10/1990 Tabor et al. ..... 435/6

OTHER PUBLICATIONS

K. B. Mullis, *Scientific American*, (Apr. 1990), "The Unusual Origin of the Polymerase Chain Reaction", pp. 56-65.  
Prober et al. (1987) *Science*, vol. 238, pp. 336-341.

*Primary Examiner*—Lisa B. Arthur  
*Attorney, Agent, or Firm*—Howrey & Simon; Jeffrey I. Auerbach

[57] **ABSTRACT**

The invention concerns a reagent composition that employs at least two different terminators of a nucleic acid template-dependent primer extension reaction to determine the identity of a nucleotide base at a specific position in a nucleic acid of interest. The invention also concerns the method for determining such identification. The invention may be used to determine the presence or absence of a specific nucleotide sequence in a sample. It may also be employed in determination of genotype and in the identification of different alleles.

**48 Claims, 8 Drawing Sheets**

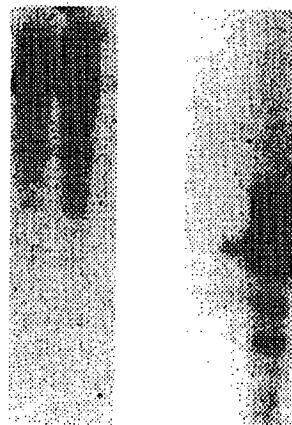
**U.S. Patent**

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**5,888,819**

180 181 180 181 180 181



**FIG. 1A FIG. 1B FIG. 1C**

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## I. Amplification primers

TGL 105: 5'-TTCTTCTTGCATCTATGTTG-3'

TGL 106: 5'-TTAAGCACCACCAACAGTCCT-3'

## II. Polymorphism detection primers

TGL 182: 5'-GCCTTGGCGTTGAGAA-3'

TGL 166: 5'-AGAGAAACAATTCAAG-3'

## III. Target sequence

5' ... TTTCTTCTTG CATCTATGTT CGTTTTTCT ATTGCTACAA 40  
 TGL 105 -----  
 ATGCCTATGC ACGGCCTGAC TTCTGCCTAG AGCCTCCATA 80  
 TACGGGTCCC TGCAAGGCCA GAATTATCAG ATA <sup>C</sup> / TTTCTAC 120  
AACGCCAAGG CTGGGCTCTG CCAGACCTTT GTATATGGTG 160  
 ----- TGL 182  
 GCTGCAGAGC TAAGAGAAC AATTCAAG <sup>A</sup> / G GCGCAGAGGA 200  
 TGL 166 -----  
 CTGCATGAGG ACCTGTGGTG GTGCTTAAGG GCCCCCGGGAA.. 3' 240  
 ----- TGL 106

## IV. Polymorphisms

| <u>Plasmid</u> | <u>Nucleotide 114</u> | <u>Nucleotide 190</u> |
|----------------|-----------------------|-----------------------|
| p183           | C                     | A                     |
| p624           | T                     | A                     |
| p814           | C                     | G                     |

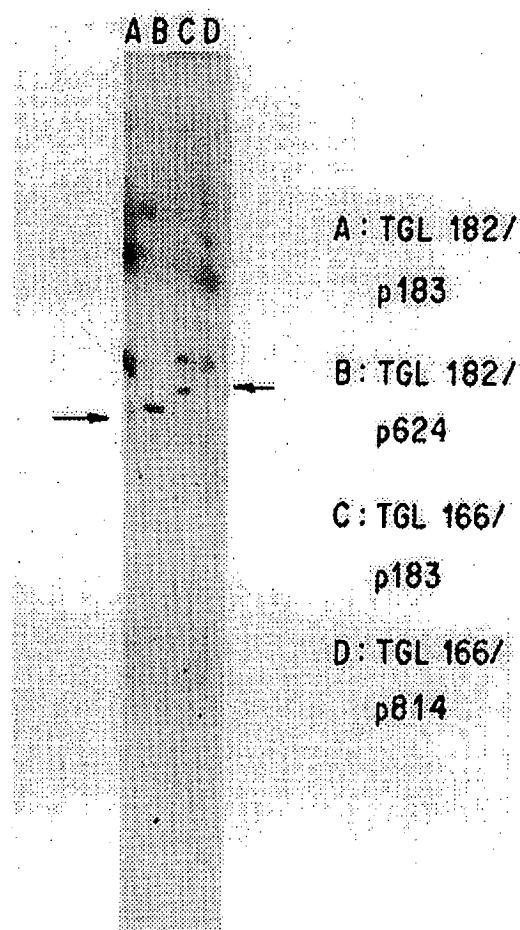
FIG.2

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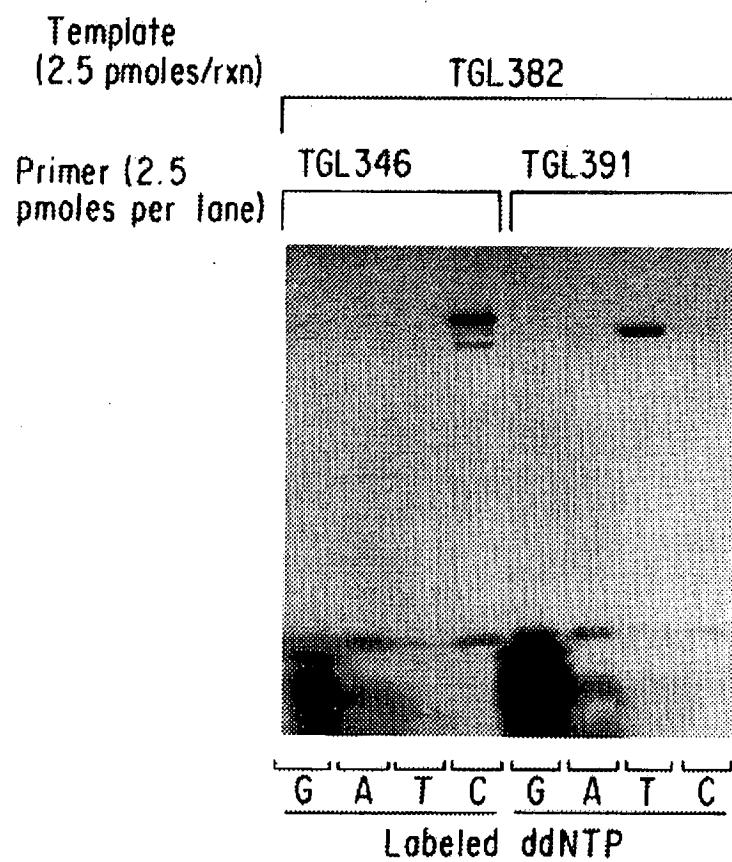
**FIG.3**

**U.S. Patent**

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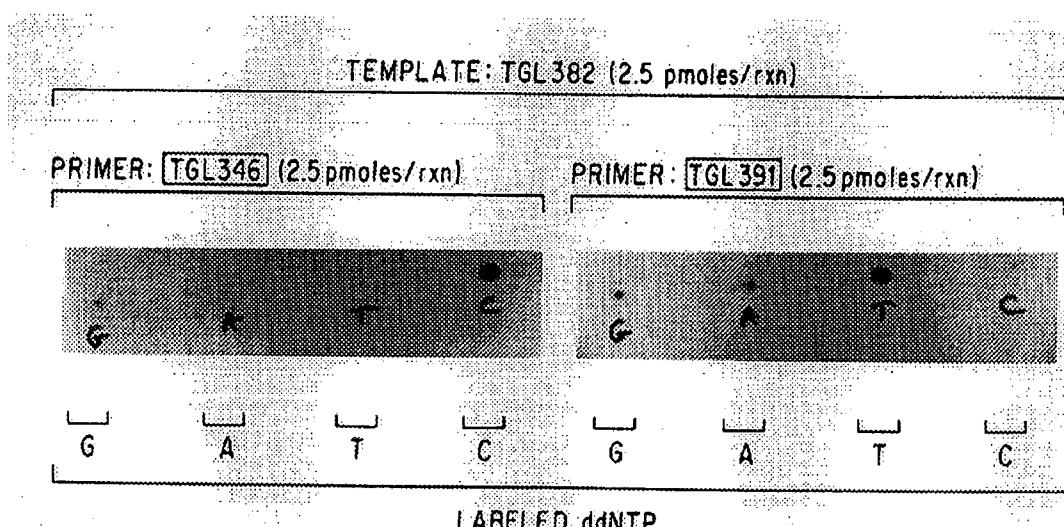
**FIG.4**

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**FIG.5**

U.S. Patent

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Attached to bead

5'  
AGATGATGCT TTTGIGCAA ACACCTTTA ACACCTCTT TAAAATTCTT TTCAAATTCTT ACGGCATTTT  
TCL240 (PCR Primer with Biotin)

TTTCCCTGAAA ATGCCCTTCGGT TTTAGGTCAA AGCTTTATTC TCCTAAGAAC CTAACCTCCA CTGGTCTCAG

GGGCCCTCTC GGAGCCCTCG GGGAGCTCTT GCCCCCAAAT CTTGGCAATT CTCCTGACA CTCGCCCAAG

TCL308  
5'  
TGGTCCGT GGTCGCCAG ACTCCGA  
GCCCTAACCC TGCACCCGGG CACCAAGCA CCACGGCA TGAGGCTICA GCAGGGAAAG CCTGCTCTCC  
GA T C B allele  
differences

TCL239 (Non-biotinylated PCR Primer)  
3'  
GTCCCCAC AGCCCTGACT CCATAACT 5'  
GTTCACACTG CTTTCAGGCC CGGCAGGTG TCGGGACTCA GGTTATTGA 3'

FIG. 6

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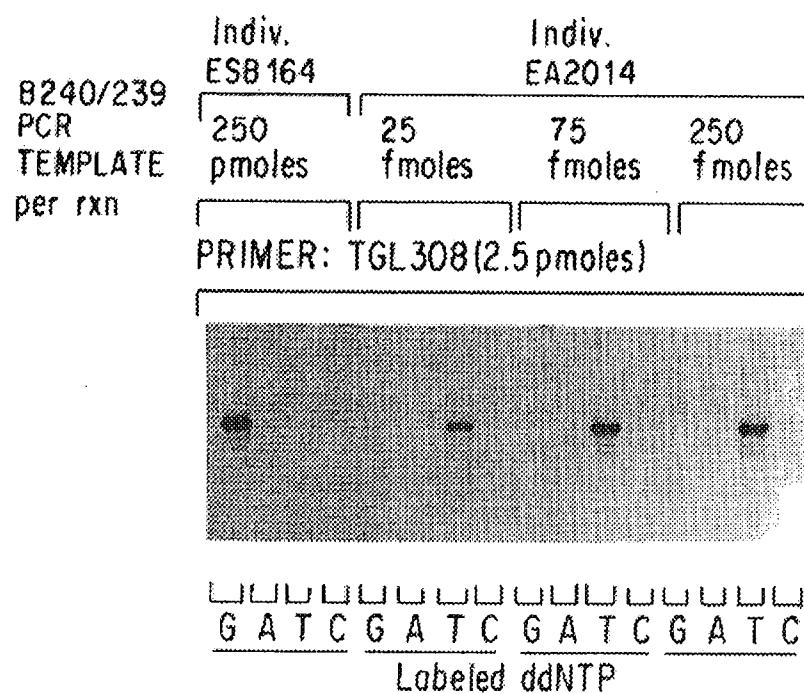


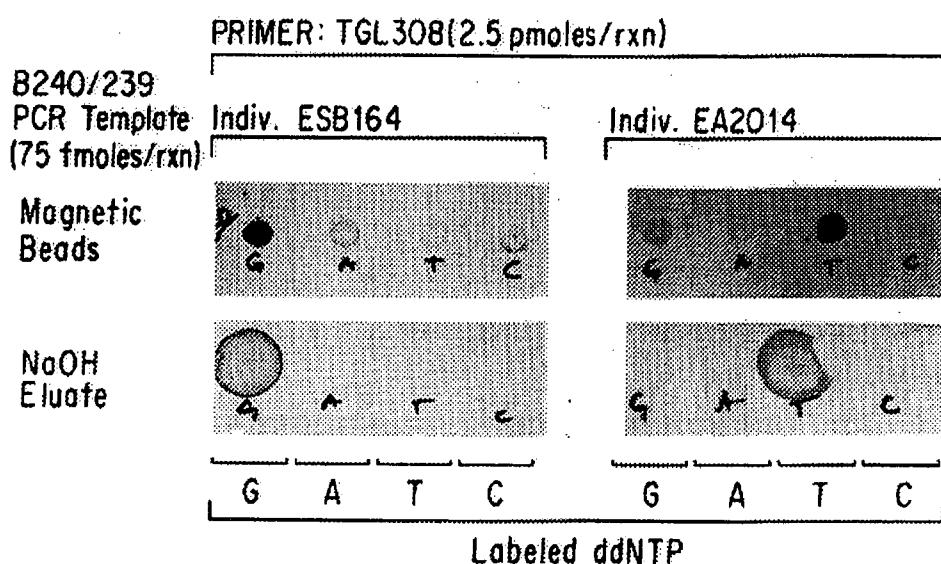
FIG. 7

U.S. Patent

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5,888,819



**METHOD FOR DETERMINING  
NUCLEOTIDE IDENTITY THROUGH  
PRIMER EXTENSION**

**BACKGROUND OF THE INVENTION**

This invention relates to the field of nucleic acid sequence detection. The detection of nucleic acid sequences can be used in two general contexts. First, the detection of nucleic acid sequences can be used to determine the presence or absence of a particular genetic element. Second, the detection of nucleic acid sequences can be used to determine the specific type of a particular genetic element that is present. Variant genetic elements usually exist. Many techniques have been developed (1) to determine the presence of specific nucleic acid sequences, and (2) to compare homologous segments of nucleic acid sequence to determine if the segments are identical or if they differ at one or more nucleotides. Practical applications of these techniques include genetic disease diagnoses, infectious disease diagnoses, forensic techniques, paternity determinations, and genome mapping.

In general, the detection of nucleic acids in a sample and the subtypes thereof depends on the technique of specific nucleic acid hybridization in which the oligonucleotide probe is annealed under conditions of high stringency to nucleic acids in the sample, and the successfully annealed probes are subsequently detected (see Spiegelman, S., *Scientific American*, Vol. 210, p. 48 (1964)).

The most definitive method for comparing DNA segments is to determine the complete nucleotide sequence of each segment. Examples of how sequencing has been used to study mutations in human genes are included in the publications of Engelke, et al., *Proc. Natl. Acad. Sci. U.S.A.*, 85:544-548 (1988) and Wong, et al., *Nature*, 330:384-386 (1987). At the present time, it is not practical to use extensive sequencing to compare more than just a few DNA segments because the effort required to determine, interpret, and compare sequence information is time-consuming.

A commonly used screen for DNA polymorphisms arising from DNA sequence variation consists of digesting DNA with restriction endonucleases and analyzing the resulting fragments by means of Southern blots, as described by Botstein, et al., *Am. J. Hum. Genet.*, 32:314-331 (1980) and White, et al., *Sci. Am.*, 258:40-48 (1988). Mutations that affect the recognition sequence of the endonuclease will preclude enzymatic cleavage at that site, thereby altering the cleavage pattern of that DNA. DNAs are compared by looking for differences in restriction fragment lengths. A major problem with this method (known as restriction fragment length polymorphism mapping or RFLP mapping) is its inability to detect mutations that do not affect cleavage with a restriction endonuclease. Thus, many mutations are missed with this method. One study, by Jeffreys, *Cell*, 18:1-18 (1979), was able to detect only 0.7% of the mutational variants estimated to be present in a 40,000 base pair region of human DNA. Another problem is that the methods used to detect restriction fragment length polymorphisms are very labor intensive, in particular, the techniques involved with Southern blot analysis.

A technique for detecting specific mutations in any segment of DNA is described in Wallace, et al., *Nucl. Acids Res.*, 9:879-894 (1981). It involves hybridizing the DNA to be analyzed (target DNA) with a complementary, labeled oligonucleotide probe. Due to the thermal instability of DNA duplexes containing even a single base pair mismatch, differential melting temperature can be used to distinguish

target DNAs that are perfectly complementary to the probe from target DNAs that differ by as little as a single nucleotide. In a related technique, described in Landegren, et al., *Science*, 41:1077-1080 (1988), oligonucleotide probes are constructed in pairs such that their junction corresponds to the site on the DNA being analyzed for mutation. These oligonucleotides are then hybridized to the DNA being analyzed. Base pair mismatch between either oligonucleotide and the target DNA at the junction location prevents the efficient joining of the two oligonucleotide probes by DNA ligase.

**A. Nucleic acid hybridization**

The base pairing of nucleic acids in a hybridization reaction forms the basis of most nucleic acid analytical and diagnostic techniques. In practice, tests based only on parameters of nucleic acid hybridization function poorly in cases where the sequence complexity of the test sample is high. This is partly due to the small thermodynamic differences in hybrid stability, generated by single nucleotide changes, and the fact that increasing specificity by lengthening the probe has the effect of further diminishing this differential stability. Nucleic acid hybridization is, therefore, generally combined with some other selection or enrichment procedure for analytical and diagnostic purposes.

Combining hybridization with size fractionation of hybridized molecules as a selection technique has been one general diagnostic approach. Size selection can be carried out prior to hybridization. The best known prior size selection technique is Southern Blotting (see Southern, E., *Methods in Enzymology*, 69:152 (1980)). In this technique, a DNA sample is subjected to digestion with restriction enzymes which introduce double stranded breaks in the phosphodiester backbone at or near the site of a short sequence of nucleotides which is characteristic for each enzyme. The resulting heterogeneous mixture of DNA fragments is then separated by gel electrophoresis, denatured, and transferred to a solid phase where it is subjected to hybridization analysis *in situ* using a labeled nucleic acid probe. Fragments which contain sequences complementary to the labeled probe are revealed visually or densitometrically as bands of hybridized label. A variation of this method is Northern Blotting for RNA molecules. Size selection has also been used after hybridization in a number of techniques, in particular by hybrid protection techniques, by subjecting probe/nucleic acid hybrids to enzymatic digestion before size analysis.

**B. Polymerase extension of duplex primer:template complexes**

Hybrids between primers and DNA targets can be analyzed by polymerase extension of the hybrids. A modification of this methodology is the polymerase chain reaction in which the purification is produced by sequential hybridization reactions of anti-parallel primers, followed by enzymatic amplification with DNA polymerase (see Saiki, et al., *Science* 239:487-491 (1988)). By selecting for two hybridization reactions, this methodology provides the specificity lacking in techniques that depend only upon a single hybridization reaction.

It has long been known that primer-dependent DNA polymerases have, in general, a low error rate for the addition of nucleotides complementary to a template. This feature is essential in biology for the prevention of genetic mistakes which would have detrimental effects on progeny. The specificity inherent in this enzymological reaction has been widely exploited as the basis of the "Sanger" or dideoxy chain termination sequencing methodology which

5,888,819

3

is the ultimate nucleic acid typing experiment. One type of Sanger DNA sequencing method makes use of mixtures of the four deoxynucleoside triphosphates, which are normal DNA precursors, and one of the four possible dideoxynucleoside triphosphates, which have a hydrogen atom instead of a hydroxyl group attached to the 3' carbon atom of the ribose sugar component of the nucleotide. DNA chain elongation in the 5' to 3' direction ("downstream") requires this hydroxyl group. As such, when a dideoxynucleotide is incorporated into the growing DNA chain, no further elongation can occur. With one dideoxynucleotide in the mixture, DNA polymerases can, from a primer/template combination, produce a population of molecules of varying length, all of which terminate after the addition of one out of the four possible nucleotides. The series of four independent reactions, each with a different dideoxynucleotide, generates a nested set of fragments, all starting at the same 5' terminus of the priming DNA molecule and terminating at all possible 3' nucleotide positions.

Another utilization of dideoxynucleoside triphosphates and a polymerase in the analysis of DNA involves labeling the 3' end of a molecule. One prominent manifestation of this technique provides the means for sequencing a DNA molecule from its 3' end using the Maxam-Gilbert method. In this technique, a molecule with a protruding 3' end is treated with terminal transferase in the presence of radioactive dideoxy-ATP. One radioactive nucleotide is added, rendering the molecule suitable for sequencing. Both methods of DNA sequencing using labeled dideoxynucleotides require electrophoretic separation of reaction products in order to derive the typing information. Most methods require four separate gel tracks for each typing determination.

The following two patents describe other methods of typing nucleic acids which employ primer extension and labeled nucleotides. Mundy (U.S. Pat. No. 4,656,127) describes a method whereby a primer is constructed complementary to a region of a target nucleic acid of interest such that its 3' end is close to a nucleotide in which variation can occur. This hybrid is subject to primer extension in the presence of a DNA polymerase and four deoxynucleoside triphosphates, one of which is an  $\alpha$ -thionucleotide. The hybrid is then digested using an exonuclease enzyme which cannot use thio-derivatized DNA as a substrate for its nucleolytic action (for example Exonuclease III of *E. coli*). If the variant nucleotide in the template is complementary to one of the thionucleotides in the reaction mixture, the resulting extended primer molecule will be of a characteristic size and resistant to the exonuclease; hybrids without thio-derivatized DNA will be digested. After an appropriate enzyme digest to remove underderivatized molecules, the thio-derivatized molecule can be detected by gel electrophoresis or other separation technology.

Vary and Diamond (U.S. Pat. No. 4,851,331) describes a method similar to that of Mundy wherein the last nucleotide of the primer corresponds to the variant nucleotide of interest. Since mismatching of the primer and the template at the 3' terminal nucleotide of the primer is counterproductive to elongation, significant differences in the amount of incorporation of a tracer nucleotide will result under normal primer extension conditions. This method depends on the use of a DNA polymerase, e.g., AMV reverse transcriptase, that does not have an associated 3' to 5' exonuclease activity. The methods of Mundy and of Vary and Diamond have drawbacks. The method of Mundy is useful but cumbersome due to the requirements of the second, different enzymological system where the non-derivatized hybrids are

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digested. The method of Vary is complicated by the fact that it does not generate discrete reaction products. Any "false" priming will generate significant noise in such a system which would be difficult to distinguish from a genuine signal.

The present invention circumvents the problems associated with the methods of Mundy and of Vary and Diamond for typing nucleic acid with respect to particular nucleotides. With methods employing primer extension and a DNA polymerase, the current invention will generate a discrete molecular species one base longer than the primer itself. In many methods, particularly those employing the polymerase chain reaction, the type of reaction used to purify the nucleic acid of interest in the first step can also be used in the subsequent detection step. Finally, with terminators which are labeled with different detector moieties (for example different fluorophors having different spectral properties), it will be possible to use only one reagent for all sequence detection experiments. Furthermore, if techniques are used to separate the terminated primers post-reaction, sequence detection experiments at more than one locus can be carried out in the same tube.

A recent article by Mullis (*Scientific American*, April 1990, pp. 56-65) suggests an experiment, which apparently was not performed, to determine the identity of a targeted base pair in a piece of double-stranded DNA. Mullis suggests using four types of dideoxynucleosides triphosphate, with one type of dideoxynucleoside triphosphate being radioactively labeled.

The present invention permits analyses of nucleic acid sequences that can be useful in the diagnosis of infectious diseases, the diagnosis of genetic disorders, and in the identification of individuals and their parentage.

A number of methods have been developed for these purposes. Although powerful, such methodologies have been cumbersome and expensive; generally involving a combination of techniques such as gel electrophoresis, blotting, hybridization, and autoradiography or non-isotopic revelation. Simpler technologies are needed to allow the more widespread use of nucleic acid analysis. In addition, tests based on nucleic acids are currently among the most expensive of laboratory procedures and for this reason cannot be used on a routine basis. Finally, current techniques are not adapted to automated procedures which would be necessary to allow the analysis of large numbers of samples and would further reduce the cost.

The current invention provides a method that can be used to diagnose or characterize nucleic acids in biological samples without recourse to gel electrophoretic size separation of the nucleic acid species. This feature renders this process easily adaptable to automation and thus will permit the analysis of large numbers of samples at relatively low cost. Because nucleic acids are the essential blueprint of life, each organism or individual can be uniquely characterized by identifiable sequences of nucleic acids. It is, therefore, possible to identify the presence of particular organisms or demonstrate the biological origin of certain samples by detecting these specific nucleic acid sequences.

#### SUMMARY OF THE INVENTION

The subject invention provides a reagent composition comprising an aqueous carrier and an admixture of at least two different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the

unpaired nucleotide base in the template immediately adjacent to, and downstream of, the 3' end of the primer. In addition, at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and one, two, three, or four of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent as described above wherein the terminators comprise nucleotides, nucleotide analogs, dideoxynucleotides, or arabinoside triphosphates. The subject invention also provides a reagent wherein the terminators comprise one or more of dideoxyadenosine triphosphate (ddATP), dideoxycytosine triphosphate (ddCTP), dideoxyguanosine triphosphate (ddGTP), dideoxythymidine triphosphate (ddTTP), or dideoxyuridine triphosphate (ddUTP).

The subject invention also provides a method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with a stretch of nucleotide bases present in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest. Enzymatic extension of the oligonucleotide primer in the resultant duplex by one nucleotide, catalyzed, for example, by a DNA polymerase, thus depends on correct base pairing of the added nucleotide to the nucleotide base to be identified.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. The net result is that the oligonucleotide primer has been extended by one terminator. Next, the identity of the detectable marker present at the 3' end of the extended primer is determined. The identity of the detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

The subject invention also provides another method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. This additional method uses a reagent containing four terminators, only one of the terminators having a detectable marker.

The subject invention also provides a method of typing a sample of nucleic acids which comprises identifying the

base or bases present at each of one or more specific positions, each such nucleotide base being identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. Each specific position in the nucleic acid of interest is determined using a different primer. The identity of each nucleotide base or bases at each position can be determined individually or the identities of the nucleotide bases at different positions can be determined simultaneously.

The subject invention further provides a method for identifying different alleles in a sample containing nucleic acids which comprises identifying the base or bases present at each of one or more specific positions. The identity of each nucleotide base is determined by the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention also provides a method for determining the genotype of an organism at one or more particular genetic loci which comprises obtaining from the organism a sample containing genomic DNA and identifying the nucleotide base or bases present at each of one or more specific positions in nucleic acids of interest. The identity of each such base is determined by using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. The identities of the nucleotide bases determine the different alleles and, thereby, determine the genotype of the organism at one or more particular genetic loci.

#### BRIEF DESCRIPTION OF THE FIGURES

FIGS. 1A-1C. Autoradiography of labeled DNA products after fractionation on a polyacrylamide/urea gel. Panel A shows products of the "A" extension reaction on oligonucleotide primer 182 (SEQ ID NO:1) directed by template oligonucleotides 180 (SEQ ID NO:2) or 181. (SEQ ID NO:3) Panel B shows products of the "B" termination reaction on oligonucleotide primer 182 (SEQ ID NO:1) annealed to template oligonucleotides 180 or 181. Panel C shows the same products as in panel B after purification on magnetic beads. Note: oligodeoxynucleotide 182 was used as supplied by Midland Certified Reagents with no further purification. The minor bands above and below the main band are presumably contaminants due to incomplete reactions or side reactions that occurred during the step-wise synthesis of the oligonucleotide. For a definition of the "A" extension reaction and the "B" termination reaction, see "A. GENERAL METHODS" in the Detailed Description of the Invention.

FIG. 2. Detection of Sequence Polymorphisms in PCR Products. Target polymorphic DNA sequence (SEQ ID NO:13) showing amplification primers, detection primers, and molecular clone (plasmid) designations. For each primer, sites of binding to one or the other strand of the target DNA sequence are indicated by underlining, and the direction of DNA synthesis is indicated by an arrow. Numbering for the target sequence is shown in the righthand margin. Polymorphic sites at positions 114 and 190 are indicated by bold lettering and a slash between the two polymorphic possibilities.

FIG. 3. Autoradiogram of gel-analyzed polymorphism test on PCR products. Templates from PCR products of p183, p624, or p814 were analyzed with the detection primers, TGL182 (SQ ID NO:1) and TGL166, (SEQ ID NO:12) in a template-directed chain extension experiment, as described in the specification. Reaction products were

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fractionated by size on a polyacrylamide/urea DNA sequencing gel, and incorporation of [ $^{35}\text{S}$ ]- $\alpha$ -thio-dideoxy adenosine monophosphate was assayed by autoradiography.

FIG. 4. Gel electrophoretic analysis of the labelled extension products of primers TGL346 (SEQ ID NO:5) and TGL391 (SEQ ID NO:4). Productive primer-template complexes of TGL346 or TGL391 with the bead-bound oligonucleotide template, TGL382, (SEQ ID NO:6) were subjected to primer extension labelling reactions with the four different [ $\alpha$ -thio- $^{35}\text{S}$ ]dideoxynucleoside triphosphate mixes. Labelled primer DNA was released from the washed beads and electrophoresed on an 8% polyacrylamide/8M urea DNA sequencing gel (2.5 pmoles of primer/lane), then analyzed by autoradiography. The four lanes shown for the primer TGL346 indicate that labelling occurred predominantly with the ddC mix, indicating that the next unpaired base in the TGL382 template adjacent to the 3' end of TGL346 was a G (see sequence given in Example 4). The four lanes shown for the primer TGL391 indicate that the labelling occurred predominantly with the ddT mix, indicating that the next unpaired base in the TGL382 template adjacent to the 3' end of TGL391 was an A.

FIG. 5. Autoradiographic analyses of total radioactivity bound to beads. The bead suspensions, containing the products of the extension reactions described in FIG. 5, were spotted onto filter paper (1 pmole of primer per spot) and exposed to X-ray film to assay total bead-bound radioactivity. As shown, TGL346 (SEQ ID NO:5) predominantly incorporated label from the ddC mix and TGL391 (SEQ ID NO:4) predominantly from the ddT mix.

FIG. 6. PCR-amplified polymorphic locus of mammalian DNA (SEQ ID NO:14 and SEQ ID NO:15). Shown is a 327 basepair segment of mammalian DNA that was amplified from samples of genomic DNA using the PCR primers TGL240 (SEQ ID NO:7) (biotinylated) and TGL239 (SEQ ID NO:8) (unbiotinylated). Samples of DNA from two homozygous individuals, ESB164 (genotype AA) and EA2014 (genotype BB), were subjected to the analyses described in Example 5. The complete DNA sequence of the A allele at this locus is shown, as SEQ ID NO:14 with the polymorphic sites where the B allele sequence (SEQ ID NO:15) differs from the A allele sequence indicated by the bases underneath the A sequence. The detection primer, TGL308, (SEQ ID NO:9) is shown base-paired with the template strand extending from the biotinylated primer. For the A allele, the first unpaired template base immediately downstream of the 3' end of TGL308 (SEQ ID NO:9) is a C, and for the B allele this base is an A. Thus, the A allele should result in labelling of TGL308 by the ddG mix only, and the B allele should result in labelling by the ddT mix only.

FIG. 7. Gel electrophoretic analysis of PCR products from two different homozygous individuals. Primers TGL240 (SEQ ID NO:7) and TGL239 (SEQ ID NO:8) were used to amplify genomic DNA (obtained from blood) from two individuals, ESB164 and EA2014. The products of the extension reactions for primer TGL308, (SEQ ID NO:19) annealed to the bead-bound, PCR-generated template as outlined in FIG. 7, were analyzed by electrophoresis on an 8% polyacrylamide/8 M urea DNA sequencing gel as outlined in FIG. 5. Shown for individual ESB164 (genotype AA: labelling expected from the ddG mix) are 250 fmoles of extended primer from the four different ddNTP labelling reactions. Shown for individual EA2014 (genotype BB: labelling expected from the ddT mix) are loadings of 25, 75, and 250 fmoles of extended primer from the four different ddNTP labelling reactions.

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FIG. 8. Autoradiographic analyses of total and NaOH-eluted radioactivity from TGL308 (SEQ ID NO:9) primer extension reactions. Primer TGL308 was used to analyze the genotypes of individuals ESB164 and EA2014 as outlined in Example 5 and FIGS. 7 and 8. Total bead-associated radioactivity was determined by directly spotting a suspension of beads containing 75 fmoles of primer onto filter paper followed by autoradiographic detection of the label in the spot. Radioactivity specifically associated with the TGL308 primer (SEQ ID NO:9) was determined by magnetically immobilizing the beads, eluting the primer with NaOH as described in Examples 4 and 5, and spotting on filter paper an amount corresponding to 75 fmoles. Label in these spots was also detected by autoradiography.

#### DETAILED DESCRIPTION OF THE INVENTION

The subject invention provides a reagent composition comprising an aqueous carrier and an admixture of at least two different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the unpaired nucleotide base in the template immediately adjacent to, and downstream of, the 3' end of the primer. In addition, at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and two, three, or four of the terminators are labeled with a different detectable marker.

The subject invention further provides a reagent as described above wherein the terminators comprise nucleotides, nucleotide analogs, dideoxynucleotides, or arabinoside triphosphates. The subject invention also provides a reagent wherein the terminators comprise one or more of dideoxyadenosine triphosphate (ddATP), dideoxycytosine triphosphate (ddCTP), dideoxyguanosine triphosphate (ddGTP), dideoxythymidine triphosphate (ddTTP), or dideoxyuridine triphosphate (ddUTP).

The subject invention further provides a reagent as described above wherein each of the detectable markers attached to the terminators is an isotopically labeled moiety, a chromophore, a fluorophore, a protein moiety, or a moiety to which an isotopically labeled moiety, a chromophore, a fluorophore, or a protein moiety can be attached. The subject invention also provides a reagent wherein each of the different detectable markers is a different fluorophore.

The subject invention also provides a reagent as described above wherein the reagent further comprises pyrophosphatase.

The invented reagent consists of two or more chain terminators with one or more of the chain terminators being identifiably tagged. This reagent can be used in a DNA polymerase primer extension reaction to type nucleic acid sequences of interest that are complementary to one or more

oligonucleotide primers by chemically or physically separating the polymerase extended primers from the chain terminator reagent and analyzing the terminal additions. Any kind of terminator that inhibits further elongation can be used, for example, a dideoxynucleoside triphosphate. Several approaches can be used for the labeling and detection of terminators: (1) radioactivity and its detection by either autoradiography or scintillation counting, (2) fluorescence or absorption spectroscopy, (3) mass spectrometry, or (4) enzyme activity, using a protein moiety. The identity of each terminator can be determined individually, i.e., one at a time. In addition, methods which permit independent analyses of each of the terminators permit analysis of incorporation of up to four terminators simultaneously.

The subject invention also provides a method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with a stretch of nucleotide bases present in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest. Enzymatic extension of the oligonucleotide primer in the resultant duplex by one nucleotide, catalyzed, for example, by a DNA polymerase, thus depends on correct base pairing of the added nucleotide to the nucleotide base to be identified.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer.

The net result is that the oligonucleotide primer has been extended by one terminator. Next, the identity of the detectable marker present at the 3' end of the extended primer is determined. The identity of the detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

The subject invention also provides another method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with nucleotide bases in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic

acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest.

5 The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four terminators, only one of the terminators having a detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. The net result is that the oligonucleotide primer has been extended by one terminator.

10 The original duplex of primer and the nucleic acid of interest is then contacted with three different reagents, with a different one of each of the four terminators being labeled in each of the four parallel reaction steps. Next, the products of the four parallel template-dependent, primer extension reactions are examined to determine which of the products has a detectable marker. The product with a detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

15 Both of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest label the primer after hybridization between the primer and the template. If the template-dependent enzyme has no exonuclease function, the 3' end of the primer must be base paired for the labeling by a terminator to occur.

20 The subject invention also provides a method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids. First, the sample of nucleic acids is treated, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids. If the nucleic acids in the sample are single-stranded, this step is not necessary. 25 Second, the sample of nucleic acids is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with the particular nucleotide sequence, if the particular nucleotide sequence is present, so as to form a duplex between the primer and the particular nucleotide sequence.

30 The duplex of primer and the particular nucleotide sequence, if any, is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the particular nucleotide sequence, if any, is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the unpaired template nucleotide base downstream of the 3' end of the primer, the primer being hybridized with the particular nucleotide sequence in the template, and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. Next, the absence or presence and identity of a detectable marker at the 3' end of the primer are determined. The presence or absence of the detectable marker indicates whether the primer has hybridized to the template. If a detectable marker is absent, the primer did not hybridize to the template, and, therefore, the particular nucleotide sequence is not present in the sample of nucleic acids. If a detectable marker is present, the primer did hybridize to the template, and, therefore, the particular nucleotide sequence is present in the sample of nucleic acids.

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The subject invention also provides another method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids. First, the sample of nucleic acids is treated, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids. Second, the sample of nucleic acids is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with the particular nucleotide sequence, if the particular nucleotide sequence is present, so as to form a duplex between the primer and the particular nucleotide sequence.

The duplex of primer and the particular nucleotide sequence, if any, is then contacted with a reagent containing four terminators, only one of the terminators having a detectable marker. The duplex of primer and the particular nucleotide sequence, if any, is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the unpaired template nucleotide base downstream of the 3' end of the primer, the primer being hybridized with the particular nucleotide sequence in the template, and the occurrence of a template-dependent, primer extension reaction. The net result is the incorporation of the terminator at the 3' end of the primer.

The original duplex of primer and the particular nucleotide sequence, if any, is then contacted with three different reagents, with a different one of each of the four terminators being labeled in each of the four parallel reaction steps. Next, the products of the four parallel, template-dependent, primer extension reactions are examined to determine which, if any, of the products have detectable markers. The absence or presence and identity of the detectable marker indicates whether the primer has hybridized to the template. If no detectable marker is present in any of the products, the primer did not hybridize to the template, and, therefore, the particular nucleotide sequence was not present in the sample of nucleic acids. If a detectable marker is present in any of the products, the primer did hybridize to the template, and, therefore, the particular nucleotide sequence was present in the sample of nucleic acids.

Different versions of the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest and the method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids are possible. In the first version, the template is a deoxyribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a DNA polymerase. This version gives a DNA product. In a second version, the template is a ribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a reverse transcriptase. This version gives a DNA product. In a third version, the template is a deoxyribonucleic acid, the primer is an oligoribonucleotide, and the enzyme is an RNA polymerase. This version gives an RNA product. In a fourth version, the template is a ribonucleic acid, the primer is an oligoribonucleotide, and the template-dependent enzyme is an RNA replicase. This version gives an RNA product.

Preferably, before the primer extension reaction is performed, the template is capped by the addition of a terminator to the 3' end of the template. The terminator is capable of terminating a template-dependent, primer extension reaction. The template is capped so that no additional

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labeled terminator will attach at the 3' end of the template. The extension reaction should occur on the primer, not on the template. A dideoxynucleotide can be used as a terminator for capping the template.

Another modification of the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest is to separate the primer from the nucleic acid of interest after the extension reaction by using appropriate denaturing conditions. The denaturing conditions can comprise heat, alkali, formamide, urea, glyoxal, enzymes, and combinations thereof. The denaturing conditions can also comprise treatment with 2.0N NaOH.

The nucleic acid of interest can comprise non-natural nucleotide analogs such as deoxyinosine or 7-deaza-2-deoxyguanosine. These analogues destabilize DNA duplexes and could allow a primer annealing and extension reaction to occur in a double-stranded sample without completely separating the strands.

The sample of nucleic acids can be from any source. The sample of nucleic acids can be natural or synthetic (i.e., synthesized enzymatically in vitro). The sample of nucleic acids can comprise deoxyribonucleic acids, ribonucleic acids, or copolymers of deoxyribonucleic acid and ribonucleic acid. The nucleic acid of interest can be a deoxyribonucleic acid, a ribonucleic acid, or a copolymer of deoxyribonucleic acid and ribonucleic acid. The nucleic acid of interest can be synthesized enzymatically in vivo, synthesized enzymatically in vitro, or synthesized non-enzymatically. The sample containing the nucleic acid or acids of interest can comprise genomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof. The sample containing the nucleic acid or acids of interest can also comprise extragenomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof. Also, the nucleic acid or acids of interest can be synthesized by the polymerase chain reaction.

The sample can be taken from any organism. Some examples of organisms to which the method of the subject invention is applicable include plants, microorganisms, viruses, birds, vertebrates, invertebrates, mammals, human beings, horses, dogs, cows, cats, pigs, or sheep.

The nucleic acid of interest can comprise one or more moieties that permit affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer. The nucleic acid of interest can comprise biotin which permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via binding of the biotin to streptavidin which is attached to a solid support. The sequence of the nucleic acid of interest can comprise a DNA sequence that permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via base pairing to a complementary sequence present in a nucleic acid attached to a solid support. The nucleic acid of interest can be labeled with a detectable marker; this detectable marker can be different from any detectable marker present in the reagent or attached to the primer.

The oligonucleotide primer can be an oligodeoxyribonucleotide, an oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides. The oligonucleotide primer can be either natural or synthetic. The oligonucleotide primer can be synthesized either enzymatically in vivo, enzymatically in vitro, or non-enzymatically in vitro. The oligonucleotide primer can be labeled with a detectable marker; this detectable marker can

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be different from any detectable marker present in the reagent or attached to the nucleic acid of interest. In addition, the oligonucleotide primer must be capable of hybridizing or annealing with nucleotides present in the nucleic acid of interest, immediately adjacent to, and upstream of, the nucleotide base to be identified. One way to accomplish the desired hybridization is to have the template-dependent primer be substantially complementary or fully complementary to the known base sequence immediately adjacent to the base to be identified.

The oligonucleotide primer can comprise one or more moieties that permit affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest. The oligonucleotide primer can comprise biotin which permits affinity separation of the primer from the unincorporated reagent and/or nucleic acid of interest via binding of the biotin to streptavidin which is attached to a solid support. The sequence of the oligonucleotide primer can comprise a DNA sequence that permits affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest via base pairing to a complementary sequence present in a nucleic acid attached to a solid support.

The subject invention also provides a method of typing a sample of nucleic acids which comprises identifying the base or bases present at each of one or more specific positions, each such nucleotide base being identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. Each specific position in the nucleic acid of interest is determined using a different primer. The identity of each nucleotide base or bases at each position can be determined individually or the identities of the nucleotide bases at different positions can be determined simultaneously.

The subject invention also provides another method of typing a sample of nucleic acids which comprises determining the presence or absence of one or more particular nucleotide sequences, the presence or absence of each such nucleotide sequence being determined using one of the methods for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids as outlined above.

The subject invention also provides an additional method of typing a sample containing nucleic acids. First, the presence or absence of one or more particular nucleotide sequences is determined; the presence or absence of each such nucleotide sequence is determined using one of the methods for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids as outlined above. Second, the nucleotide base or bases present at each of one or more specific positions is identified; each such base is identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention further provides a method for identifying different alleles in a sample containing nucleic acids which comprises identifying the base or bases present at each of one or more specific positions. The identity of each nucleotide base is determined by the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention also provides a method for determining the genotype of an organism at one or more particular genetic loci which comprises obtaining from the organism a sample containing genomic DNA and identifying the nucleotide base or bases present at each of one or more

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specific positions in nucleic acids of interest. The identity of each such base is determined by using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. The identity of the nucleotide bases determine the different alleles and, thereby, determine the genotype of the organism at one or more particular genetic loci.

The chain termination reagent in combination with an appropriate oligonucleotide primer, and a DNA polymerase with or without an associated 3' to 5' exonuclease function, and an appropriate salt and cofactor mixture, can be used under appropriate hybridization conditions as a kit for diagnosing or typing nucleic acids, if appropriate primer separation techniques are used. To simplify the primer separation and the terminal nucleotide addition analysis this invention makes use of oligonucleotides that are modified in such ways that permit affinity separation as well as polymerase extension. The 5' termini and internal nucleotides of synthetic oligonucleotides can be modified in a number of different ways to permit different affinity separation approaches, e.g., biotinylation. These affinity reagents can be used with the terminator mixture to facilitate the analysis of extended oligonucleotide(s) in two ways:

(1) If a single affinity group is used on the oligonucleotide(s), the oligonucleotide(s) can be separated from the unincorporated terminator reagent. This eliminates the need of physical or size separation.

(2) More than one oligonucleotide can be separated from the terminator reagent and analyzed simultaneously if more than one affinity group is used. This permits the analysis of several nucleic acid species or more nucleic acid sequence information per extension reaction.

The affinity group(s) need not be on the priming oligonucleotide but could, alternatively, be present on the template. As long as the primer remains hydrogen bonded to the template during the affinity separation step, this will allow efficient separation of the primer from unincorporated terminator reagent. This also has the additional benefit of leaving sites free on the primer for the convenient attachment of additional moieties. For example, the 5'-terminus of the primer could be modified by coupling it to a suitable fluorescent group such as rhodamine, allowing the amount of primer in the primer:template complex to be easily quantified after the affinity separation step. The amounts of 3'-terminating terminators could then be normalized to the total amount of annealed primer.

The oligonucleotide primers and template can be any length or sequence, can be DNA or RNA, or any modification thereof. It is necessary, however, that conditions are chosen to optimize stringent hybridization of the primers to the target sequences of interest.

The conditions for the occurrence of the template-dependent, primer extension reaction can be created, in part, by the presence of a suitable template-dependent enzyme. Some of the suitable template-dependent enzymes are DNA polymerases. The DNA polymerase can be of several types. The DNA polymerase must, however, be primer and template dependent. For example, *E. coli* DNA polymerase I or the "Klenow fragment" thereof, T4 DNA polymerase, T7 DNA polymerase ("Sequenase"), *T. aquaticus* DNA polymerase, or a retroviral reverse transcriptase can be used. RNA polymerases such as T3 or T7 RNA polymerase could also be used in some protocols. Depending upon the polymerase, different conditions must be used, and different temperatures ranges may be required for the hybridization and extension reactions.

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The reagents of the subject invention permit the typing of nucleic acids of interest by facilitating the analysis of the 3' terminal addition of terminators to a specific primer or primers under specific hybridization and polymerase chain extension conditions. Using only the terminator mixture as the nucleoside triphosphate substrate ensures addition of only one nucleotide residue to the 3' terminus of the primer in the polymerase reaction. Using all four terminators simultaneously ensures fidelity, i.e., suppression of misreading.

By specifically labeling one or more of the terminators, the sequence of the extended primer can be deduced. In principle, more than one reaction product can be analyzed per reaction if more than one terminator is specifically labeled.

By specifically tagging the oligonucleotide primer(s), or template(s) with a moiety that does not affect the 3' extension reaction yet permits affinity separation, the extension product(s) can be separated post-reaction from the unincorporated terminators, other components of the reagents, and/or the template strand. Several oligonucleotides can be analyzed per extension reaction if more than one affinity agent is used.

In principle, the combination of four differently labeled terminators and many primers or templates tagged with different groups permits the typing of many different nucleic acid sequences simultaneously.

Specificity in this diagnostic reaction is determined by (1) the stringency of oligonucleotide hybridization and (2) the sequence information gained by the single residue extension.

#### A. General Methods

##### 1. Biotinylation of oligodeoxynucleotides.

Oligodeoxynucleotides, terminated at their 5'-ends with a primary amino group, were ordered from Midland Certified Reagents, Midland, Tex. These were biotinylated using biotin-XX-NHS ester (Clontech Laboratories, Inc., Palo Alto, Calif.), a derivative of biotin-N-hydroxysuccinimide. Reagents used were from the Clontech biotinylation kit. Typically, the oligonucleotide (9 nanomoles) was dissolved in 100  $\mu$ l of 0.1M NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> (pH 9), and 25  $\mu$ l of N,N-dimethylformamide containing 2.5 mg biotin-XX-NHS-ester was added. The mixture was incubated overnight at room temperature. It was then passed over a 6 ml Sephadex G-25 column ("DNA grade"—Pharmacia) equilibrated with H<sub>2</sub>O. Eluate fractions containing DNA were identified by mixing 4  $\mu$ l aliquots with an equal volume of ethidium bromide (2  $\mu$ g/ml) and the DNA-induced fluorescence was monitored with a UV transilluminator. Unreacted ester was detected by UV absorption at 220 nm. The tubes containing DNA were pooled, concentrated in a Centricon-3 microconcentrator (Amicon), and passed over Sephadex again.

Inhibition of the binding of [<sup>3</sup>H]-biotin to magnetic M-280 streptavidin Dynabeads (Dynal) was used to assay quantitatively the extent of biotinylation of the oligonucleotides. Eppendorf tubes and pipet tips were siliconized. A known amount (5–10 pmoles) of biotin-labeled oligonucleotide in 10  $\mu$ l 0.1M NaCl was added to tubes containing 25  $\mu$ l of 1:4 suspension of beads in 0.1M NaCl. The tubes were rotated for one hour on a Labquake shaker (Labindustries, Inc.). Increasing amounts of [<sup>3</sup>H]-biotin (5–35 pmoles) in 20  $\mu$ l of 0.1M NaCl were added to the tubes and these were rotated again for one hour. Tubes were put on a Dynal MPC-E magnet to remove the beads from suspension, 10  $\mu$ l aliquots of the supernatant were withdrawn, and the amount

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of radioactivity in these was measured using a Beckman LS 5000 TD liquid scintillation counter. Counts were compared to those from tubes to which no oligonucleotide had been added. Alternatively, for some primers, biotinylation was monitored by size fractionation of the reaction products using analytical polyacrylamide gel electrophoresis in the presence of 8M urea.

##### 2. Template-dependent primer extension/termination reactions.

Approximately five pmoles of 5'-biotinylated oligodeoxynucleotide template (see above) were mixed with approximately three pmoles of primer in 1X sequencing buffer (from Sequenase Version 2.0 kit, US Biochemical Corp.) (10  $\mu$ l final volume), the mixture was incubated at 65° C. for 2 min, then allowed to cool to room temperature in order to anneal the primer and template. The solution containing the annealed template-primer was separated into two 5  $\mu$ l portions, A and B, to which were added the following: Reactions A (for normalizing template concentrations)—0.5  $\mu$ l of 100 mM dithiothreitol, 1  $\mu$ l each of 10  $\mu$ M DATP, dGTP, ddCTP, 0.5  $\mu$ l of "Mn buffer" (from Sequenase Version 2.0 kit, US Biochemical Corp.), 0.5  $\mu$ l of [<sup>35</sup>S]- $\alpha$ -thio-dTTP (10 mCi/ml, 1180 Ci/mmol) (Dupont-NEN), 1  $\mu$ l of Sequenase (1:8 dilution, US Biochemical Corp.); Reactions B (for template-specific labeling of primer 3'-ends)—same additions as in Reactions A except the nucleotides used were ddCTP, ddGTP, ddTTP, and [<sup>35</sup>S]- $\alpha$ -thio-ddATP. Reactions were for 5 min at 37° C. Control reactions omitting the primer or the Sequenase were also performed. Aliquots were removed and analyzed by electrophoresis on a 15% polyacrylamide, 8 M urea, DNA sequencing gel (see Maniatis, T., et al., *Molecular Cloning: a Laboratory Manual*, Cold Spring Harbor Laboratory (1982)). The gel was fixed in 10% methanol, 10% acetic acid, dried down onto Whatman's 3MM paper, and exposed to Kodak X-Omat AR film. Alternatively, for purposes of analyzing the products by liquid scintillation counting, the biotinylated template or template-primer was bound to an excess of M-280 streptavidin Dynabeads (Dynal) before or after the Sequenase reaction (see above, "1. Biotinylation of oligodeoxynucleotides", for binding conditions). Beads were washed three times with 0.1M NaCl to remove unincorporated label, then scintillation fluid was added and the radioactivity measured by liquid scintillation counting.

##### 3. Generation of templates from polymerase chain reaction products

Polymerase chain reaction (PCR) reactions were carried out where one or the other of the amplification primers flanking the target stretch of DNA were biotinylated as described above. These primers (2  $\mu$ mol final concentration) and the target DNA (up to 1  $\mu$ g) were incubated with 2.5 units of Taq polymerase (Perkin Elmer/Cetus), 200  $\mu$ M each of DATP, dCTP, dGTP, and dTTP, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl<sub>2</sub>, and 0.01% gelatin (Sigma). Reaction mixtures were overlayed with paraffin oil and incubated for 30 cycles in Perkin Elmer/Cetus thermocycler. Each cycle consisted of 1 min at 94° C., 2 min at 60° C., and 3 min at 72° C. Reaction products were purified by phenol/chloroform extraction and ethanol precipitation, then analyzed by ethidium bromide staining after electrophoresis on a polyacrylamide gel. The yield of duplex PCR product was typically about 10  $\mu$ g.

Approximately 5  $\mu$ g of this PCR product was incubated with gentle agitation for 60 min with 50  $\mu$ L of a suspension of prewashed M-280 Dynabeads in 0.1M NaCl. The beads with the bound DNA (approximately 15 pmoles) were then

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incubated for 5 min at 25° C. with 0.15M NaOH. Beads were washed once with 0.15M NaOH to remove the unbiotinylated DNA strand, then washed three times with H<sub>2</sub>O. The beads were resuspended in H<sub>2</sub>O and the strand bound to the beads via the biotin-streptavidin link was used as template for further primer extension reactions.

#### B. Examples

##### Example 1

|                                   |   |
|-----------------------------------|---|
| Primer oligo 182:                 | 5' GCCTTGGCGTGTAGAA'                      |
| Template oligos<br>180(C)/181(T): | 3' TCGGGTCCGAACCGAACATCITC/<br>TATAGACTA' |

Oligonucleotides 180 (SEQ ID NO:2) and 181 (SEQ ID NO:3) were synthesized with primary amino groups attached to their 5' termini. These were coupled with biotin as described above. Oligonucleotide 182 (SEQ ID NO:4) was annealed as a primer and extension reactions "A" and "B" (see above) were carried out. The expected template-dependent 3'-terminal extensions to oligonucleotide 182 (SEQ ID NO:1) were as follows ("\*" preceding a nucleotide signifies a radioactive label):

| Template | Reaction A        | Reaction B |
|----------|-------------------|------------|
| 180      | dG-*dT-dk-*dT-ddC | -ddG       |
| 181      | dA-*dT-dA-*dT-ddC | -*ddA      |

Thus, in the "A" reactions, both template oligonucleotides will direct a radioactively-labelled five nucleotide extension of the primer; the amount of labeling should be proportional to the amount of productively primed template present in the reactions. In the "B" reactions, both templates will direct a one nucleotide extension of the primer, but only for template 181 (SEQ ID NO:3) should this result in labeling of the primer. The "B" reaction, therefore, is an example of template-directed, sequence-specific labeling of an oligonucleotide via DNA polymerase-catalyzed extension of a productive primer-template complex.

The reaction products were fractionated by size on a 15% polyacrylamide/8M urea sequencing gel and visualized by autoradiography. The results (FIG. 1) show that, as expected, the "A" reactions yield labeling and extension of both primers whereas the "B" reaction results in labeling that is strongly biased in favor of template 181 (SEQ ID NO:3). Panel C in FIG. 1 shows a gel analysis of the same reaction products as in Panel B, except the reaction products were first purified as described above using M-280 streptavidin Dynabeads.

##### Example 2

The experiment described in Example 1 shows template-directed labeling of oligonucleotide primer 182 (SEQ ID NO:4) in which the labeling is specific with respect to oligonucleotides or other species that migrate similarly on a polyacrylamide gel. In order to assess more generally the template-directed specific labeling of oligonucleotide 182 (SEQ ID NO:1) with respect to all other labeled species, regardless of gel mobility, a direct measurement of incorporated radioactivity was performed. In this experiment, both reactions "A" and "B" were performed, reaction products were purified using Dynabeads, and total radioactivity in the aliquots was measured by liquid scintillation counting.

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This procedure assesses both misincorporation of label into other species and, in addition, the efficiency of the Dynabead washing procedure with respect to unincorporated nucleotides. As a practical matter, it would be of interest to minimize both sources of non-specific label in order to have a simple, non-gel-based, procedure for assessing specific, template-directed labeling of the primer. The results of directly counting the reaction products after washing on the magnetic beads are as follows (all results expressed as cpm 10 of <sup>35</sup>S):

| Reaction         | Template 180 | Template 181 |
|------------------|--------------|--------------|
| A, complete      | 325,782      | 441,823      |
| A, no polymerase | 5,187        | 5,416        |
| A, no primer     | 4,351        | 12,386       |
| B, complete      | 5,674        | 176,291      |
| B, no polymerase | 2,988        | 1,419        |
| B, no primer     | 1,889        | 1,266        |

As can be seen from these results, specific template-directed labeling of primer 182 (SEQ ID NO:1) can also be determined by measuring the total radioactivity of the reaction products after washing with magnetic beads to remove unreacted nucleotides. The background in this experiment due to nonspecific label from all other sources was approximately 3-4% (compare templates 180 (SEQ ID NO:2) and 181 (SEQ ID NO:30 in the "B, complete" reaction). Control experiments ("no polymerase" and "no primer") showed that the bulk of the background label was probably contributed by unincorporated nucleotides that were not completely removed by the washing step. The "A, complete" reactions showed that, for both templates, productive template:primer complexes were present.

##### Example 3

Two amplification primers, TGL 105 (SEQ ID NO:10) and TGL 106 (SEQ ID NO:11) (FIG. 2), were used to amplify a cloned stretch of bovine DNA (SEQ ID NO:13) containing two DNA sequence polymorphisms: a C or T at position 114 and an A or G at position 190 (FIG. 2). DNAs containing these polymorphisms were molecularly cloned and available on plasmids, as follows: plasmid p183, C114 and A190; plasmid p624, T114 and A190; plasmid p814, C114 and G190. Four PCR reactions with biotinylated primers were performed to amplify and purify specific strands of these plasmids for use as templates:

| Primers                                 | Plasmids      | Detection Primers          |
|---|---------------|----------------------------|
| 105 biotinylated,<br>106 unbiotinylated | p183 and p624 | TGL 182<br>(SEQ ID NO: 1)  |
| 105 unbiotinylated,<br>106 biotinylated | p183 and p814 | TGL 166<br>(SEQ ID NO: 12) |

The duplex PCR products were bound to magnetic microspheres, denatured with NaOH, and the biotinylated strand purified as described above. Templates prepared with biotinylated TGL 105 (SEQ ID NO:10) were subjected to analysis by DNA sequencing with unbiotinylated primer TGL 106 (SEQ ID NO:11) in order to measure the amount of template present. Similarly, template prepared using biotinylated TGL 106 (SEQ ID NO:11) was analyzed by sequencing with unbiotinylated TGL 105 (SEQ ID NO:10).

Approximately equal amounts of template (2 pmoles) were annealed for 5 min at 65° C to the polymorphism detection primers, TGL 182 (SEQ ID NO:1) and TGL 166

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(SEQ ID NO:12) (see above and FIG. 2). These primers hydrogen-bond to the templates in a sequence-specific fashion such that their 3'-termini are adjacent to nucleotide positions 114 and 190, respectively (FIG. 2 and SEQ ID NO:13). Template-directed primer extension reactions (reaction "B" conditions) were carried out on these primer:template complexes in the presence of the four ddNTPs, one of which (ddATP) was labeled. The products of these extension reactions were analyzed by electrophoresis on a 15% polyacrylamide/8M urea gel followed by autoradiography (FIG. 3).

## Example 4

|                        |  |    |
|------------------------|--|----|
| Primer oligo TGL391:   | <sup>5</sup> TGTTTGACACAAAGCA <sup>3'</sup><br>(SEQ ID NO: 4)                                      | 15 |
| Primer oligo TGL346:   | <sup>5</sup> GTTTGACAAAGCAT <sup>3'</sup><br>(SEQ ID NO: 5)  |    |
| Template oligo TGL382: | <sup>3'</sup> CACAAAACGTGTTTCGTAGGA <sup>5'</sup><br>biotin: (streptavidin-bead)<br>(SEQ ID NO: 6) | 20 |

Oligonucleotide TGL382 (SEQ ID NO:6) was purchased from the Midland Certified Reagent Company, Midland, Tex. It was biotinylated using Midland Certified Reagent Company's "Biotin dX" reagent (a biotin derivative phosphoramidite) which is suitable for use in automated DNA synthesis in the 5' terminal nucleotide position. The biotinylated oligonucleotide was then purified by anion exchange HPLC. Streptavidin-conjugated M-280 Dynabeads were washed in TNET buffer (10 mM Tris-HCl, pH 7.5, 100 mM NaCl, 1 mM EDTA, 0.1% Triton X-100) and resuspended in the same buffer at a concentration of  $7 \times 10^8$  beads/ml. 10–100 pmoles of biotinylated oligonucleotide TGL382 (SEQ ID NO:6) was incubated with 100  $\mu$ l of the Dynabead suspension in TNET for 30 minutes at 20° C. in order to allow the biotin moiety to bind to the streptavidin. The beads were then washed (using a magnet to immobilize them) three times with 200  $\mu$ l of TNET and resuspended in 100  $\mu$ l of TNET. For annealing, 25  $\mu$ l of this suspension of the Dynabeads with the attached template oligonucleotide was immobilized with the magnet, the TNET withdrawn, and 25  $\mu$ l of 40 mM Tris-HCl, pH 7.5, 20 mM MgCl<sub>2</sub>, 50 mM NaCl, containing 2  $\mu$ M of oligonucleotide primers 346 or 391, was added. The template and each primer were annealed by incubating them for 5 minutes at 65° C., followed by slow cooling over a period of 20 minutes to room temperature. Beads containing the bound template-primer complexes were washed twice with 200  $\mu$ l TNET, followed by resuspension in 25  $\mu$ l of 40 mM Tris-HCl, pH 7.5, 20 mM MgCl<sub>2</sub>, 50 mM NaCl.

The following ddNTP mixes were used:

| <sup>35</sup> S-labelled dideoxynucleoside triphosphate mixes<br>(labelled nucleotide indicated in the form ddN*TP): |                                      |                  |                  |
|--|--------------------------------------|------------------|------------------|
| ddG Mix:   | 5 $\mu$ M ddG*TP<br>10 $\mu$ M ddCTP | 10 $\mu$ M ddATP | 10 $\mu$ M ddTTP |
| ddA Mix:   | 10 $\mu$ M ddGTP<br>10 $\mu$ M ddCTP | 5 $\mu$ M ddA*TP | 10 $\mu$ M ddTTP |
| ddT Mix:   | 10 $\mu$ M ddGTP<br>10 $\mu$ M ddCTP | 10 $\mu$ M ddATP | 5 $\mu$ M ddT*TP |
| ddC Mix:   | 10 $\mu$ M ddGTP<br>5 $\mu$ M ddC*TP | 10 $\mu$ M ddATP | 10 $\mu$ M ddTTP |

The ddN\*TPs were the four respective [ $\alpha$ -thio-<sup>35</sup>S] dideoxynucleoside triphosphates (purchased from New England Nuclear).

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For each bead-bound, template-primer complex, four extension reactions were carried out, one reaction for each of the four ddNTP mixes. Extension reactions contained the following components: 5.0  $\mu$ l bead suspension containing the annealed template-primer complex, 0.5  $\mu$ l of 100 mM dithiothreitol, 0.5  $\mu$ l of "Mn<sup>++</sup> solution" (100 mM MnCl<sub>2</sub>, 150 mM DL-isocitrate, pH 7.0; purchased from U.S. Biochemicals, Cleveland, Ohio), 1.0  $\mu$ l of ddG, ddA, ddT, or ddC mix, 2.0  $\mu$ l of H<sub>2</sub>O, and 1.0  $\mu$ l of T7 DNA polymerase ("Sequenase", version 2.0, US Biochemicals, 1625 units/ml in 50 mM Tris-HCl, pH 7.5, 10 mM 2-mercaptoethanol, 1 mg/ml bovine serum albumin).

Reactions were allowed to proceed for 15 minutes at 20° C., then stopped by washing the magnetically immobilized beads three times with 500  $\mu$ l TNET. Beads were resuspended in final volume of 25  $\mu$ l TNET prior to the detection assays.

Incorporation of labelled dideoxynucleotides by the primer extension reaction was assayed two different ways: gel electrophoresis followed by autoradiography, and direct autoradiographic analysis of labelled DNA.

1. Gel electrophoresis followed by autoradiography (<sup>35</sup>S-labelled material only). Samples of washed, bead-bound DNA were heated at 94° C. for 5 minutes in 10  $\mu$ l of formamide loading buffer (80% formamide, 10 mM Tris-HCl, pH 8, 1 mM EDTA, 0.02% bromphenol blue) to denature the DNA and release the labelled primer from the primer:template complex. Samples were analyzed by electrophoresis on 8 or 12.5% polyacrylamide/8M urea sequencing gels (19:1 acrylamide:bis-acrylamide ratio; 100 mM Tris-HCl, 100 mM borate, 2 mM EDTA, pH 8.3, running buffer; 60 watts constant power). After electrophoresis, gels were either dried down onto filter paper or frozen at -80° C. to prevent diffusion, covered with plastic wrap, and exposed to X-ray film to visualize the labelled DNA by autoradiography (FIG. 4).

2. Direct autoradiographic analysis of labelled DNA. For the analysis of total radioactivity bound to the beads, 10  $\mu$ l aliquots of the bead suspensions in TNET were spotted directly onto filter paper or nylon membranes. Filters or membranes were dried under an incandescent lamp, covered with plastic wrap, and exposed to X-ray film (FIG. 5).

## Example 5

|         |   |
|---------|---|
| TGL240: | <sup>5</sup> AGATGATGCTTGTGCAAAACAC <sup>3'</sup> (SEQ ID NO: 7)    |
| TGL239: | <sup>5</sup> TCAATACCTGAGTCGGACACCCCTG <sup>3'</sup> (SEQ ID NO: 8) |
| TGL308: | <sup>5</sup> AGCCTCAGACCGCGTGGGCCCTGGT <sup>3'</sup> (SEQ ID NO: 9) |

Oligonucleotide TGL240 (SEQ ID NO:2) was synthesized with a primary amino group attached to its 5' terminus and coupled with biotin as described above. TGL240 (biotinylated) and TGL239 (SEQ ID NO:8) (unbiotinylated) were used to amplify, via the polymerase chain reaction procedure (see "A. General Methods"), a region of DNA comprising a particular genetic locus in samples of mammalian genomic DNA. DNAs from two different individuals, each homozygous for a particular set of linked sequence polymorphisms (the "A" allele and the "B" allele—see FIG. 6 and SEQ ID NO:14 and SEQ ID NO:15), were examined. After the PCR reaction, 2–20 pmoles of duplex PCR DNA was incubated with 100  $\mu$ l of streptavidin-conjugated M-280 Dynabeads ( $7 \times 10^8$  beads/ml) in TNET buffer in order to bind the biotinylated strand to the beads. After binding, the beads were magnetically immobilized and

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washed three times with 200  $\mu$ l of TNET, then resuspended in 100  $\mu$ l of TNET. To remove the non-biotinylated strand, 500  $\mu$ l of 0.15N NaOH was added and the suspension incubated for 30 minutes at 20° C. The beads were then magnetically immobilized and washed once with 250  $\mu$ l of 0.15N NaOH, three times with 500  $\mu$ l TNET, and resuspended in 100  $\mu$ l of TNET.

The detection primer, oligonucleotide TGL308 (SEQ ID NO:9) (FIG. 6), was annealed to the bead-bound PCR-generated template as described above in Example 4. Further washes, extension reactions, and detection assays were also carried out as described in Example 4. A gel autoradiographic analysis of the labelled primer extension products for the two homozygous individuals, ESB164 ("AA" genotype) and EA2014 ("BB" genotype), is shown in FIG. 7. Autoradiographic analyses of total bead-bound radioactivity, or primer-associated radioactivity after NaOH elution, are shown for these same individuals using the filter spotting assay (FIG. 8). For the analysis of primer only, 10  $\mu$ l of 0.4N NaOH was added to 10  $\mu$ l of the bead suspension. After incubation for 10 minutes at room temperature, the beads were immobilized magnetically and the supernatant withdrawn and spotted onto nylon blotting membrane.

#### C. Preferred Embodiment

A particularly advantageous way to practice the present invention involves obtaining from a convenient source, such as blood, epithelium, hair, or other tissue, samples of DNA or RNA, then amplifying *in vitro* specific regions of the nucleic acid using the polymerase chain reaction, transcription-based amplification (see Kwoh, et al., Proc. Natl. Acad. Sci. 80:1173 (1989)), etc. Amplification is accomplished using specific primers flanking the region of interest, with one or more of the primers being modified by having an attached affinity group (although in any given reaction only one such primer is modified at a time). A preferred modification is attachment of biotin moieties to the 5'-termini of the primers. A sample (typically, 0.5-5 pmoles) of the amplified DNA is then bound to streptavidin-conjugated magnetic microspheres (e.g., Dynal M-280 "Dynabeads") via the attached biotin moiety on the amplification primer. The DNA is denatured by adjusting the aqueous suspension containing the microspheres to a sufficiently alkaline pH, and the strand bound to the microspheres via the biotin-streptavidin link is separated from the complementary strand by washing under similar alkaline conditions. To accomplish this, the microspheres are centrifuged or immobilized by the application of a magnetic field. The microsphere-bound strand is then used as a template in the remaining manipulations.

To the template strand, generated as described above, a specific primer oligonucleotide is bound under high stringency annealing conditions, the sequence of the primer being consistent with unique binding to a site on the template strand immediately adjacent to a known DNA sequence polymorphism. A preferred sequence and mode of binding for the primer ensures that the primer forms a duplex with the template such that the 3'-terminal nucleotide of the primer forms a Watson-Crick basepair with the template nucleotide immediately adjacent to the site of the first nucleotide in the sequence polymorphism, without the duplex overlapping any of the polymorphic sequence to be analyzed. This arrangement causes the nucleotides added via

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template-directed, DNA polymerase-catalyzed, extension of the primer to be determined unambiguously by the polymorphic nucleotide sequence in the template.

The above-described primer:template complex is contacted, under conditions of salt, pH, and temperature compatible with template-directed DNA synthesis, with a suitable DNA polymerase and four different chain-terminating nucleotide analogues known to form specific base pairs with the bases in the template. Most likely, but not necessarily, the bases in the template as well as the chain-terminating analogues are based on the common nucleosides: adenosine, cytosine, guanine or inosine, thymidine or uridine. A preferred set of chain-terminating analogues are the four dideoxynucleoside triphosphates, ddATP, ddCTP, ddGTP, and ddTTP, where each of the four ddNTPs has been modified by attachment of a different fluorescent reporter group. These fluorescent tags would have the property of having spectroscopically distinguishable emission spectra, and in no case would the dideoxynucleoside triphosphate modification render the chain-terminating analogue unsuitable for DNA polymerase-catalyzed incorporation onto primer 3'-termini. The result of DNA polymerase-catalyzed chain extension in such a mixture with such a primer:template complex is the quantitative, specific and unambiguous incorporation of a fluorescent chain-terminating analogue onto the 3'-terminus of the primer, the particular fluorescent nucleotide added being solely dictated by the sequence of the polymorphic nucleotides in the template.

The fluorescently-tagged primer:template complex, still attached to the magnetic microspheres, is then separated from the reaction mix containing the unincorporated nucleotides by, for example, washing the magnetically immobilized beads in a suitable buffer. Additionally, it is desirable in some circumstances to then elute the primer from the immobilized template strand with NaOH, transfer the eluted primer to a separate medium or container, and subsequently determine the identity of the incorporated terminator. The identity of the attached fluorescent group is then assessed by illuminating the modified DNA strand with light, preferably provided by a laser, of a suitable wavelength and intensity and spectrophotometrically analyzing the emission spectrum produced. In general, for a two allele (diploid) system at any given site in the DNA sequence, there are ten possible canonical emission spectra produced, corresponding to the sixteen possible homozygotic and heterozygotic pairings. By suitable matching of the measured spectra to this library of canonical spectra it is possible to identify which chain-terminating nucleotide(s) have been added to the 3'-terminus of the primer and thereby identify the nature of the sequence polymorphism in the template. Spectra produced by multiple allele systems or by alleles present in a ratio other than 1:1 can also be deconvolved by suitable mathematical treatments to identify and estimate the relative ratios of each contributing nucleotide.

All of the above steps involve chemistries, manipulations, and protocols that have been, or are amenable to being, automated. Thereby, incorporation of the preferred mode of practice of this invention into the operation of a suitably programmed robotic workstation should result in significant cost savings and increases in productivity for virtually any diagnostic procedure that depends on the detection of specific nucleotide sequences or sequence differences in nucleic acids derived from biological samples.

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## SEQUENCE LISTING

## ( 1 ) GENERAL INFORMATION:

( i i i ) NUMBER OF SEQUENCES: 15

## ( 2 ) INFORMATION FOR SEQ ID NO:1:

## ( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 17 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( 1 i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:1:

G C C T T G G C G T   T G T A G A A

1 7

## ( 2 ) INFORMATION FOR SEQ ID NO:2:

## ( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 32 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

## ( i x ) FEATURE:

- ( A ) NAME/KEY: modified\_base
- ( B ) LOCATION:
- ( D ) OTHER INFORMATION: /note= "5'terminus biotinylated using biotin-XX-NHS ester, a derivative of biotin-N- hydroxysuccinimide, from Clontech

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:2:

A T C A G A T A C T   T C T A C A A C G C   C A A G G C T G G G   C T

3 2

## ( 2 ) INFORMATION FOR SEQ ID NO:3:

## ( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 32 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

## ( i x ) FEATURE:

- ( A ) NAME/KEY: modified\_base
- ( B ) LOCATION:
- ( D ) OTHER INFORMATION: /note= "5'terminus biotinylated using biotin-XX-NHS ester, a derivative of biotin-N- hydroxysuccinimide, from Clontech

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:3:

A T C A G A T A T T   T C T A C A A C G C   C A A G G C T G G G   C T

3 2

## ( 2 ) INFORMATION FOR SEQ ID NO:4:

## ( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 17 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:4:

T G T T T T G C A C   A A A A G C A

1 7

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-continued

( 2 ) INFORMATION FOR SEQ ID NO:5:

( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 17 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:5:

G T T T T G C A C A A A G C A T

1 7

( 2 ) INFORMATION FOR SEQ ID NO:6:

( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 22 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( i x ) FEATURE:

- ( A ) NAME/KEY: modified\_base
- ( B ) LOCATION:
- ( D ) OTHER INFORMATION: /note= "5'terminus biotinylated using Midland Certified Reagent Company's Biotin dX reagent, a biotin derivative phosphoramidite."

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:6:

A G G A T G C T T T T G T G C A A A C A C

2 2

( 2 ) INFORMATION FOR SEQ ID NO:7:

( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 24 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( i x ) FEATURE:

- ( A ) NAME/KEY: modified\_base
- ( B ) LOCATION:
- ( D ) OTHER INFORMATION: /note= "5'terminus biotinylated using biotin-XX-NHS ester, a derivative of biotin-N- hydroxysuccinimide, from Clontech

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:7:

A G A T G A T G C T T T T G T G C A A A A C A C

2 4

( 2 ) INFORMATION FOR SEQ ID NO:8:

( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 25 base pairs
- ( B ) TYPE: nucleic acid
- ( C ) STRANDEDNESS: single
- ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:8:

T C A A T A C C T G A G T C C C G A C A C C C T G

2 5

( 2 ) INFORMATION FOR SEQ ID NO:9:

( i ) SEQUENCE CHARACTERISTICS:

- ( A ) LENGTH: 25 base pairs

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-continued

( B ) TYPE: nucleic acid  
 ( C ) STRANDEDNESS: single  
 ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:9:

AGCCTCAGAC CGCGTGGTGC CTGGT

25

( 2 ) INFORMATION FOR SEQ ID NO:10:

( i ) SEQUENCE CHARACTERISTICS:  
 ( A ) LENGTH: 21 base pairs  
 ( B ) TYPE: nucleic acid  
 ( C ) STRANDEDNESS: single  
 ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TTCTTCTTGC ATCTATGTT G

21

( 2 ) INFORMATION FOR SEQ ID NO:11:

( i ) SEQUENCE CHARACTERISTICS:  
 ( A ) LENGTH: 21 base pairs  
 ( B ) TYPE: nucleic acid  
 ( C ) STRANDEDNESS: single  
 ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TTAAGCACCA CCACAGGTCC T

21

( 2 ) INFORMATION FOR SEQ ID NO:12:

( i ) SEQUENCE CHARACTERISTICS:  
 ( A ) LENGTH: 17 base pairs  
 ( B ) TYPE: nucleic acid  
 ( C ) STRANDEDNESS: single  
 ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: Other nucleic acid

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:12:

AGAGAAACAA TTTCAAG

17

( 2 ) INFORMATION FOR SEQ ID NO:13:

( i ) SEQUENCE CHARACTERISTICS:  
 ( A ) LENGTH: 240 base pairs  
 ( B ) TYPE: nucleic acid  
 ( C ) STRANDEDNESS: double  
 ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: DNA (genomic)

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:13:

|   |     |
|---|-----|
| TTTCTTCTTG CATCTATGTT CGTTTTTCT ATTGCTACAA ATGCCCTATGC ACGGCCTGAC | 60  |
| TTCTGCCTAG AGCCTCCATA TACGGGTCCC TGCAAGGCCA GAATTATCAG ATAYTTCTAC | 120 |
| AACGCCAAGG CTGGGCTCTG CCAGACCTTT GTATATGCTG GCTOCAGAGC TAAGAGAAAC | 180 |
| AATTTCAGG GCGCAGAGGA CTGCATGAGG ACCTGTGGTG GTGCTTAAGG GCCCCGGGAA  | 240 |

( 2 ) INFORMATION FOR SEQ ID NO:14:

( i ) SEQUENCE CHARACTERISTICS:

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29

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-continued

( A ) LENGTH: 327 base pairs  
 ( B ) TYPE: nucleic acid  
 ( C ) STRANDEDNESS: double  
 ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: DNA (genomic)

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:14:

|            |            |             |            |            |            |       |
|------------|------------|-------------|------------|------------|------------|-------|
| AGATGATGCT | TTTGTGAAA  | ACACTTTTA   | ACACCTCTTT | TAAAATTCT  | TTCAAATTCT | 6 0   |
| ACGGCATT   | TTTCTGAAA  | ATGCTTCGGT  | TTTAGGTCAA | AGCTTTATTC | TCCTAAGAAC | 1 2 0 |
| CTAACTCCC  | CTGGTCTCAG | GCGCCCTCTC  | GGAGCCCTCG | GGGAGTCTTT | GCCCCCAAT  | 1 8 0 |
| CTTGGCATT  | TCCCCGTACA | CTCGCCCAAG  | GCCCCCTAAC | TGCACCCGGG | CACCAGGCAC | 2 4 0 |
| CACGCGGTCT | GAGGCTTCAG | CAGGGAAAGGC | CTGCTCTCCG | TTCACACTGC | TTTCAGGCC  | 3 0 0 |
| GGCAGGGTGT | CGGGACTCAG | GTATTGA     |            |            |            | 3 2 7 |

( 2 ) INFORMATION FOR SEQ ID NO:15:

( i ) SEQUENCE CHARACTERISTICS:  
 ( A ) LENGTH: 328 base pairs  
 ( B ) TYPE: nucleic acid  
 ( C ) STRANDEDNESS: double  
 ( D ) TOPOLOGY: linear

( i i ) MOLECULE TYPE: DNA (genomic)

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:15:

|             |             |            |            |            |            |       |
|-------------|-------------|------------|------------|------------|------------|-------|
| AGATGATGCT  | TTTGTGAAA   | ACACTTTTA  | ACACCTCTTT | TAAAATTCT  | TTCAAATTCT | 6 0   |
| ACGGCATT    | TTTCTGAAA   | ATGCTTCGGT | TTTAGGTCAA | AGCTTTATTC | TCCTAAGAAC | 1 2 0 |
| CTAACTCCC   | CTGGTCTCAG  | GCGCCCTCTC | GGAGCCCTCG | GGGAGTCTTT | GCCCCCAAT  | 1 8 0 |
| CTTGGCATT   | TCCCCGTACA  | CTCGCCCAAG | GCCCCCTAAC | TGCACCCGGG | GAACCAGGCA | 2 4 0 |
| CCACGCGGTCT | TGAGGCTTCAG | GTAGGCAAGG | CCTGCTCTCC | TTTCACACTG | CTTCAGGCC  | 3 0 0 |
| GGGCAGGGTGT | TCGGGACTCAG | GGTATTGA   |            |            |            | 3 2 8 |

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What we claim is:

1. A method of determining the identity of a nucleotide base at a specific position in a nucleic acid of interest, which comprises:

(a) treating a sample containing the nucleic acid of interest, if the nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position, or directly employing step (b) if the nucleic acid of interest is single-stranded;

(b) contacting the sample from step (a), with an oligonucleotide primer which is fully complementary to and which hybridizes specifically to a stretch of nucleotide bases present in the nucleic acid of interest immediately adjacent to the nucleotide base to be identified, under high stringency hybridization conditions, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in said duplex; and

(c) contacting the duplex from step (b), in the absence of dATP, dCTP, dGTP, or dTTP, with at least two different terminators of a nucleic acid template-dependent, primer extension reaction capable of specifically terminating the extension reaction in a manner strictly dependent upon the identity of the unpaired nucleotide base in the template immediately downstream of the 3'

end of the primer wherein one of said terminators is complementary to said nucleotide base to be identified and wherein at least one of said terminators is labeled with a detectable marker; wherein said contacting is under conditions sufficient to permit base pairing of said complementary terminator with the nucleotide base to be identified and occurrence of a template-dependent primer extension reaction sufficient to incorporate said complementary terminator onto the 3' end of the primer to thereby extend said 3' end of said primer by one terminator;

(d) determining the presence and identity of the nucleotide base at the specific position in the nucleic acid of interest by detecting the detectable marker of said incorporated terminator while said terminator is incorporated at the 3' end of the extended primer, and wherein said detection is conducted in the absence of non-terminator nucleotides.

2. The method of claim 1, wherein in step (c), the duplex from step (b) is contacted with four labeled terminators, wherein only one of the terminators has a detectable marker, and wherein in said method said step (c) is performed four times, with a different one of the four terminators being labelled.

3. The method of claim 1, wherein in step (c), the duplex from step (b) is contacted with four labeled terminators, each with a different detectable label.

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4. A method of typing a sample containing nucleic acids which comprises identifying the nucleotide base or bases present at each of one or more specific positions, each such nucleotide base being identified using the method of claim 2 or 3, and each such specific position being determined using a different primer.

5. A method of claim 4, wherein the identity of each nucleotide base or bases at each position is determined individually or wherein the identities of the nucleotide bases at different positions are determined simultaneously.

6. A method for identifying different alleles in a sample containing nucleic acids which comprises identifying the nucleotide base or bases present at each of one or more specific positions, each such nucleotide base being identified by the method of claim 2 or 3.

7. A method for determining the genotype of an organism at one or more particular genetic loci which comprises:

- (a) obtaining from the organism a sample containing genomic DNA; and
- (b) identifying the nucleotide base or bases present at each of one or more specific positions in nucleic acids of interest, each such base or bases being identified using the method of claim 2 or 3, and thereby identifying different alleles and thereby, in turn, determining the genotype of the organism at one or more particular genetic loci.

8. A method of claim 2 or 3, wherein the conditions for the occurrence of the template-dependent, primer extension reaction in step (c) are created, in part, by the presence of a suitable template-dependent enzyme.

9. A method of claim 8, wherein the template-dependent enzyme is *E. coli* DNA polymerase I or the "Klenow fragment" thereof, T4 DNA polymerase, T7 DNA polymerase *T. aquaticus* DNA polymerase, a retroviral reverse transcriptase, or combinations thereof.

10. A method of claim 2 or 3, wherein the nucleic acid of interest is a deoxyribonucleic acid, a ribonucleic acid, or a copolymer of deoxyribonucleic acid and ribonucleic acid.

11. A method of claim 2 or 3, wherein the primer is an oligodeoxyribonucleotide, an oligoribonucleotide, or a copolymer of deoxyribonucleic acid and ribonucleic acid.

12. A method of claim 2 or 3, wherein the template is a deoxyribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a DNA polymerase.

13. A method of claim 2 or 3, wherein the template is a ribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a reverse transcriptase.

14. A method of claim 2 or 3, wherein the template is a deoxyribonucleic acid, the primer is an oligoribonucleotide, and the enzyme is an RNA polymerase.

15. A method of claim 2 or 3, wherein the template is a ribonucleic acid, the primer is an oligoribonucleotide, and the template-dependent enzyme is an RNA replicase.

16. A method of claim 2 or 3, wherein, prior to the primer extension reaction in step (c), the template has been capped at its 3' end by the addition of a terminator to the 3' end of the template, said terminator being capable of terminating a template-dependent, primer extension reaction.

17. A method of claim 16, wherein the terminator is a dideoxynucleotide.

18. A method of claim 2 or 3, wherein the nucleic acid of interest has been synthesized enzymatically in vivo, synthesized enzymatically in vitro, or synthesized non-enzymatically.

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19. A method of claim 2 or 3, wherein the oligonucleotide primer has been synthesized enzymatically in vivo, synthesized enzymatically in vitro, or synthesized non-enzymatically.

20. A method of claim 2 or 3, wherein the oligonucleotide primer comprises one or more moieties that permit affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest.

21. A method of claim 20, wherein the oligonucleotide primer comprises biotin which permits affinity separation of the primer from the unincorporated reagent and/or nucleic acid of interest via binding of the biotin to streptavidin which is attached to a solid support.

22. A method of claim 2 or 3, wherein the sequence of the oligonucleotide primer comprises a DNA sequence that permits affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest via base pairing to a complementary sequence present in a nucleic acid attached to a solid support.

23. A method of claim 2 or 3, wherein the nucleic acid of interest comprises one or more moieties that permit affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer.

24. A method of claim 23, wherein the nucleic acid of interest comprises biotin which permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via binding of the biotin to streptavidin which is attached to a solid support.

25. A method of claim 2 or 3, wherein the sequence of the nucleic acid of interest comprises a DNA sequence that permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via base pairing to a complementary sequence present in a nucleic acid attached to a solid support.

26. A method of claim 2 or 3, wherein the oligonucleotide primer is labeled with a detectable marker.

27. A method of claim 26, wherein the oligonucleotide primer is labeled with a detectable marker that is different from any detectable marker present in the reagent or attached to the nucleic acid of interest.

28. A method of claim 2 or 3, wherein the nucleic acid of interest is labeled with a detectable marker.

29. A method of claim 28, wherein the nucleic acid of interest is labeled with a detectable marker that is different from any detectable marker present in the reagent or attached to the primer.

30. A method of claim 2 or 3, wherein the nucleic acid of interest comprises non-natural nucleotide analogs.

31. A method of claim 30, wherein the non-natural nucleotide analogs comprise deoxyinosine or 7-deaza-2'-deoxyguanosine.

32. A method of claim 2 or 3, wherein the nucleic acid of interest has been synthesized by the polymerase chain reaction.

33. A method of claim 2 or 3, wherein the sample comprises genomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof.

34. A method of claim 2 or 3, wherein the sample comprises extragenomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof.

35. A method of claim 2 or 3, wherein the primer is separated from the nucleic acid of interest after the primer extension reaction in step (c) by using appropriate denaturing conditions.

36. A method of claim 35, wherein the denaturing conditions comprise heat, alkali, formamide, urea, glyoxal, enzymes, and combinations thereof.

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37. A method of claim 36, wherein the denaturing conditions comprise treatment with 0.2N NaOH.

38. A method of claim 33, wherein the organism is a plant, microorganism, virus, or bird.

39. A method of claim 33, wherein the organism is a vertebrate or invertebrate.

40. A method of claim 33, wherein the organism is a mammal.

41. A method of claim 40, wherein the mammal is a human being.

42. A method of claim 40, wherein the mammal is a horse, dog, cow, cat, pig, or sheep.

43. A method of determining the presence or absence of a particular target nucleic acid molecule in a sample of nucleic acids, which comprises:

(a) treating the sample of nucleic acids, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids, or directly employing step (b) if the sample of nucleic acids contains only single-stranded nucleic acids;

(b) contacting the sample from step (a), with an oligonucleotide primer which is fully complementary to and which hybridizes specifically to a stretch of nucleotide bases present in said target nucleic acid molecule immediately adjacent to a preselected nucleotide base, under high stringency hybridization conditions so as to form a duplex between the primer and the nucleic acid of interest such that the preselected nucleotide base is the first unpaired base in the template immediately downstream of the 3' end of the primer in said duplex; and

(c) contacting the duplex from step (b), in the absence of dATP, dCTP, dGTP, or dTTP, with at least two different terminators of a nucleic acid template-dependent, primer extension reaction capable of specifically terminating the extension reaction in a manner strictly dependent upon the identity of the unpaired nucleotide base in the template immediately downstream of the 3' end of the primer wherein one of said terminators is complementary to said nucleotide base to be identified and wherein at least one of said terminators is labeled with a detectable marker; wherein said contacting is under conditions sufficient to permit base pairing of said complementary terminator with said unpaired nucleotide base immediately downstream of the 3' end of said hybridized primer and occurrence of a template-dependent primer extension reaction sufficient to incorporate said complementary terminator onto the 3' end of the primer to thereby extend said 3' end of said primer by one terminator;

(d) determining the presence of the target nucleic acid molecule by detecting the detectable marker of said incorporated terminator while said terminator is incorporated at the 3' end of the extended primer, and wherein said detection is conducted in the absence of non-terminator nucleotides.

44. The method of claim 43, wherein in step (c), the duplex from step (b) is contacted with four labeled terminators, wherein only one of the terminators has a detectable marker, and wherein in said method said step (p) is performed four times, with a different one of the four terminators being labelled.

45. The method of claim 43, wherein in step (c), the duplex from step (b) is contacted with four labeled terminators, each with a different detectable label.

46. A method of determining the identity of a nucleotide base at a specific position in a nucleic acid of interest which comprises:

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(a) (1) incubating a sample containing the nucleic acid of interest with at least two oligonucleotide primers, and a polymerase, said primers being sufficient to mediate a polymerase chain reaction amplification of said nucleic acid of interest, wherein said incubation is conducted under conditions sufficient to permit said amplification to occur;

(2) treating a sample containing said amplified nucleic acid of interest, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position, or directly employing step (b) if the nucleic acid of interest is single-stranded;

(b) contacting the sample from step (a2), under hybridizing conditions, with an oligonucleotide primer which is fully complementary to and which hybridizes specifically to a stretch of nucleotide bases present in the nucleic acid of interest immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in said duplex; and

(c) contacting the duplex from step (b), in the absence of dATP, dCTP, dGTP, or dTTP, with at least two different terminators of a nucleic acid template-dependent, primer extension reaction capable of specifically terminating the extension reaction in a manner strictly dependent upon the identity of the unpaired nucleotide base in the template nucleic acid of interest immediately downstream of the 3' end of the primer wherein one of said terminators is complementary to said nucleotide base to be identified and wherein at least one of said terminators is labeled with a detectable marker, wherein said contacting is under conditions sufficient to permit base pairing of said complementary terminator with the nucleotide base to be identified and occurrence of a template-dependent primer extension reaction sufficient to incorporate said complementary terminator onto the 3' end of the primer to thereby extend said 3' end of said primer by one terminator;

(d) determining the presence and identity of the nucleotide base at the specific position in the nucleic acid of interest by detecting the detectable marker of said incorporated terminator while said terminator is incorporated at the 3' end of the extended primer, and wherein said detection is conducted in the absence of non-terminator nucleotides.

47. A method of determining the presence or absence of a particular target nucleic acid molecule in a sample of nucleic acids which comprises:

(a) (1) incubating said sample of nucleic acids with at least two oligonucleotide primers, and a polymerase, said primers being sufficient to mediate a polymerase chain reaction amplification of said desired polynucleotide if said desired polynucleotide is present in said sample, wherein said incubation is conducted under conditions sufficient to permit said amplification to occur;

(2) treating said incubated sample of nucleic acids, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids, or directly employing step (b) if the sample of nucleic acids contains only double-stranded nucleic acids;

(b) contacting the sample from step (a2), under hybridizing conditions, with an oligonucleotide primer which

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is fully complementary to and which hybridizes specifically to a stretch of nucleotide bases present in said target nucleic acid molecule immediately adjacent to a preselected nucleotide base, so as to form a duplex between the primer and the nucleic acid of interest such that the preselected nucleotide base is the first unpaired base in the template immediately downstream of the 3' end of the primer in said duplex; and

(c) contacting the duplex from step (b), in the absence of dATP, dCTP, dGTP, or dTTP, with at least two different terminators of a nucleic acid template-dependent, primer extension reaction capable of specifically terminating the extension reaction in a manner strictly dependent upon the identity of the unpaired nucleotide base in the template immediately downstream of the 3' end of the primer wherein one of said terminators is complementary to said nucleotide base to be identified and wherein at least one of said terminators is labeled with a detectable marker; wherein said contacting is under conditions sufficient to permit base pairing of

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said complementary terminator with said unpaired nucleotide base immediately downstream of the 3' end of said hybridized primer and occurrence of a template-dependent, primer extension reaction sufficient to incorporate said complementary terminator onto the 3' end of the primer to thereby extend said 3' end of said primer by one terminator; and

(d) determining the presence of the target nucleic acid molecule by detecting the detectable marker of said incorporated terminator while said terminator is incorporated at the 3' end of the extended primer, and wherein said detection is conducted in the absence of non-terminator nucleotides.

48. A method of typing a sample containing nucleic acids which comprises determining the presence or absence of one or more particular nucleotide sequences, the presence or absence of each such nucleotide sequence being determined by the method of claim 40 or 47.

\* \* \* \* \*







US006004744A

**United States Patent** [19]  
**Goelet et al.**

[11] Patent Number: **6,004,744**  
[45] Date of Patent: **\*Dec. 21, 1999**

[54] **METHOD FOR DETERMINING  
NUCLEOTIDE IDENTITY THROUGH  
EXTENSION OF IMMOBILIZED PRIMER**

[75] Inventors: **Phillip Goelet**, Cockeysville; **Michael R. Knapp**, Baltimore, both of Md.; **Stephen Anderson**, Princeton, N.J.

[73] Assignee: **Molecular Tool, Inc.**, Baltimore, Md.

[\*] Notice: This patent is subject to a terminal disclaimer.

[21] Appl. No.: **07/775,786**

[22] Filed: **Oct. 11, 1991**

**Related U.S. Application Data**

[63] Continuation-in-part of application No. 07/664,837, Mar. 5, 1991, Pat. No. 5,888,819.

[51] Int. Cl.<sup>6</sup> ..... **C12Q 1/70**; C12Q 1/68; C12P 19/34; C07H 21/04

[52] U.S. Cl. ..... **435/5**; 435/6; 435/91.2; 435/23.1

[58] Field of Search ..... 435/6, 91.2, 5; 536/26.12, 23.1, 24.33, 26.1

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*Primary Examiner*—Lisa B. Arthur  
*Attorney, Agent, or Firm*—Howrey & Simon; Jeffrey I. Auerbach

[57] **ABSTRACT**

The invention concerns a reagent composition that employs at least two different terminators of a nucleic acid template-dependent primer extension reaction to determine the identity of a nucleotide base at a specific position in a nucleic acid of interest. The invention also concerns an immobilized method for determining such identification. The invention may be used to determine the presence or absence of a specific nucleotide sequence in a sample. It may also be employed in determination of genotype and in the identification of different alleles.

**56 Claims, 12 Drawing Sheets**

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## I. Amplification primers

TGL 105: 5'-TTCTTCTTGCATCTATGTTCG-3'

TGL 106: 5'-TTAAGCACCACACAGGTCT-3'

## II. Polymorphism detection primers

TGL 182: 5'-GCCTTGGCGTTGTAGAA-3'

TGL 166: 5'-AGAGAAACAATTCAG-3'

## III. Target sequence

5' ... TTTCTTCTTG CATCTATGTT CGTTTTTCT ATTGCTACAA 40  
TGL 105 ----->  
ATGCCATATGC ACGGCCTGAC TTCTGCCTAG AGCCTCCATA 80  
TACGGGTCCC TGCAAGGCCA GAATTATCAG ATA <sup>C</sup> / TTTCTAC 120  
AACGCCAAGG CTGGGCTCTG CCAGACCTT GTATATGGTG 160  
----- TGL 182  
GCTGCAGAGC TAAGAGAAC AATTCAG <sup>A</sup> / G GGGCAGAGGA 200  
TGL 166 ----->  
CTGCATGAGG ACCTGTGGTG GTGCTTAAGG GCCCCCGGGAA.. 3' 240  
----- TGL 106

## IV. Polymorphisms

| <u>Plasmid</u> | <u>Nucleotide 114</u> | <u>Nucleotide 190</u> |
|----------------|-----------------------|-----------------------|
| p183           | C                     | A                     |
| p624           | T                     | A                     |
| p814           | C                     | G                     |

**FIG.2**

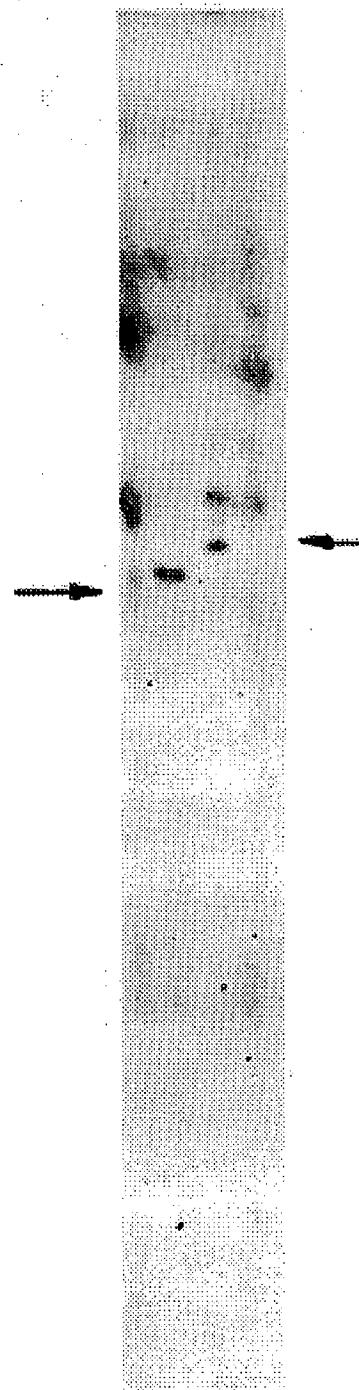
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**A B C D**



**A : TGL 182/**

**p183**

**B : TGL 182/**

**p624**

**C : TGL 166/**

**p183**

**D : TGL 166/**

**p814**

**FIG.3**

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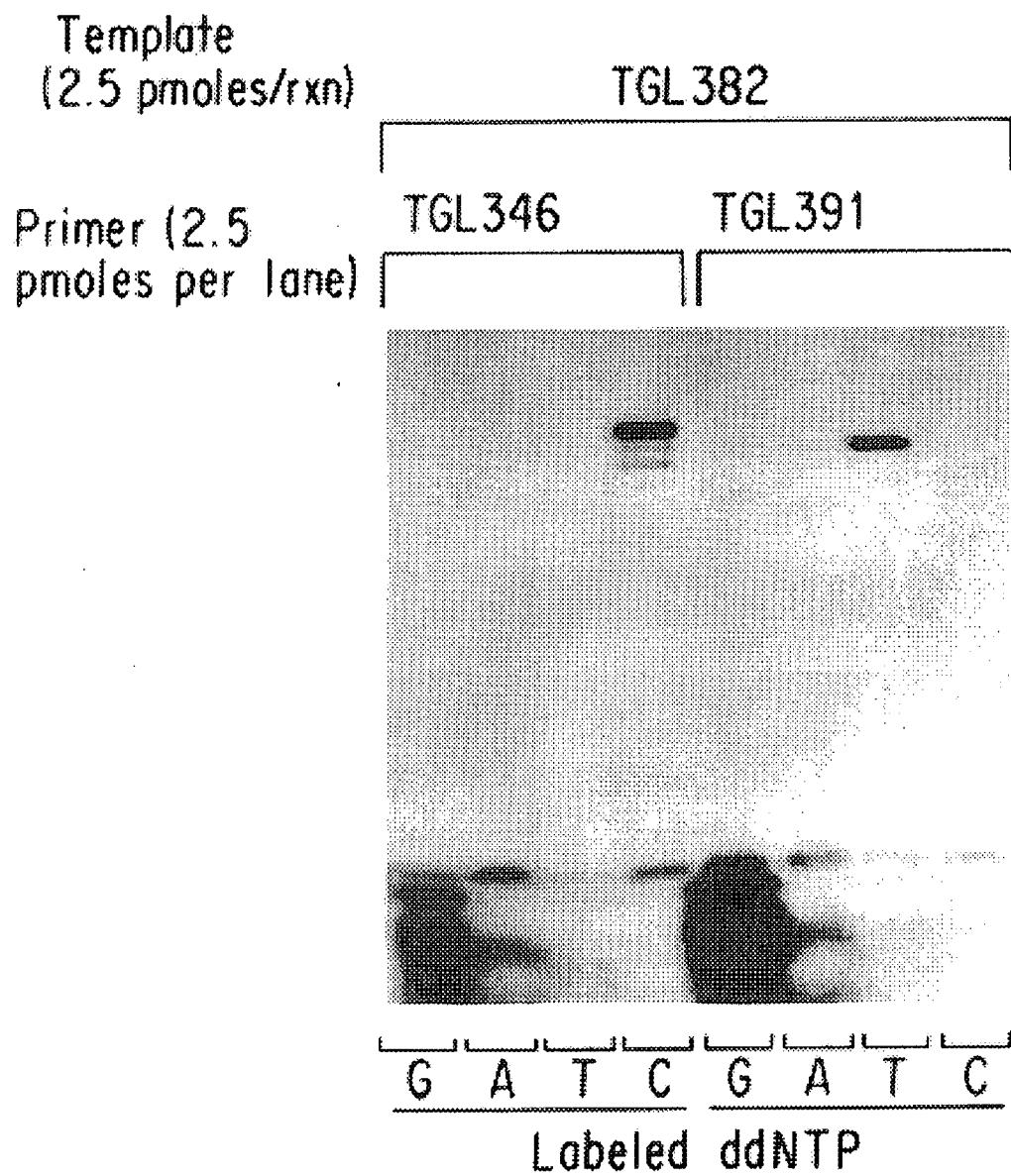


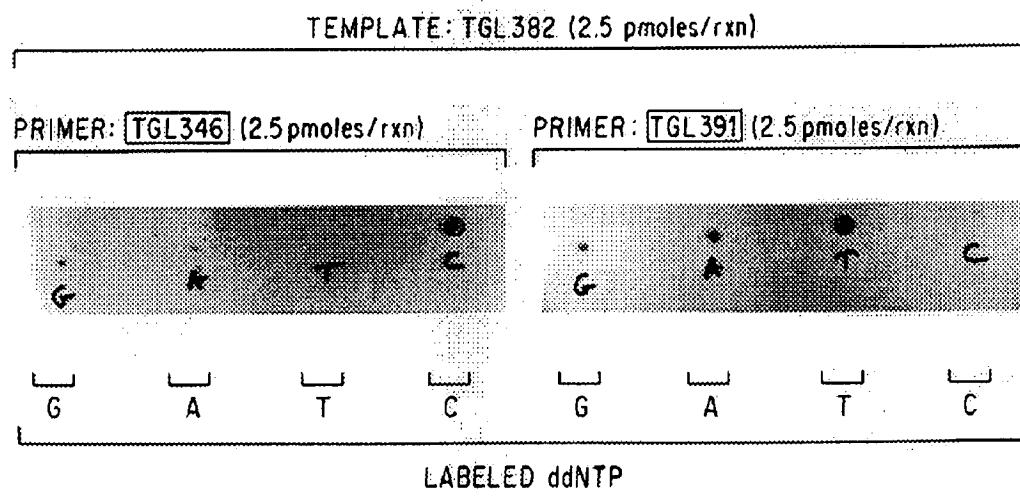
FIG.4

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Attached to bead

5' AGATGATGCT TTTGTCAAA ACACCTTTA ACACCTCTT TAAATTCTT TTCAAATTCTT ACGGCATT  
TGL240 (PCR Primer with Biotin)

TTCCCTGAAA ATGCCCTCGGT TTTAGGTCAA AGCTTTATC TCCIAAGAAC CTAACTCCA CTGGTCTCAG

GGGCCCTCTC GGAGGCCCTCG GGGAGTCTTT GCCCCCAAAT CTTGGCATT CTTGGCTGACA CTCGGCCAAAG

TGL308  
3' TGGTCCGT GCTGGCCAG ACTCCGA  
CCCCCTAACCC TGCACCCCCC 5'  
CACCAGCA CCACGGGTG TGAGGCTTCA GCAGGGAAAGG CCTGGCTCTC  
T C B allele  
GA differences

TGL239 (Non-biotinylated PCR Primer)  
3' GTCCCCAC AGCCCTGAGT CCATAACT 5'  
GTTCACACTG CTTCAGGCC CGGCAGGGTC TGGGACATCA GGTATTGA 3'

**FIG.6**

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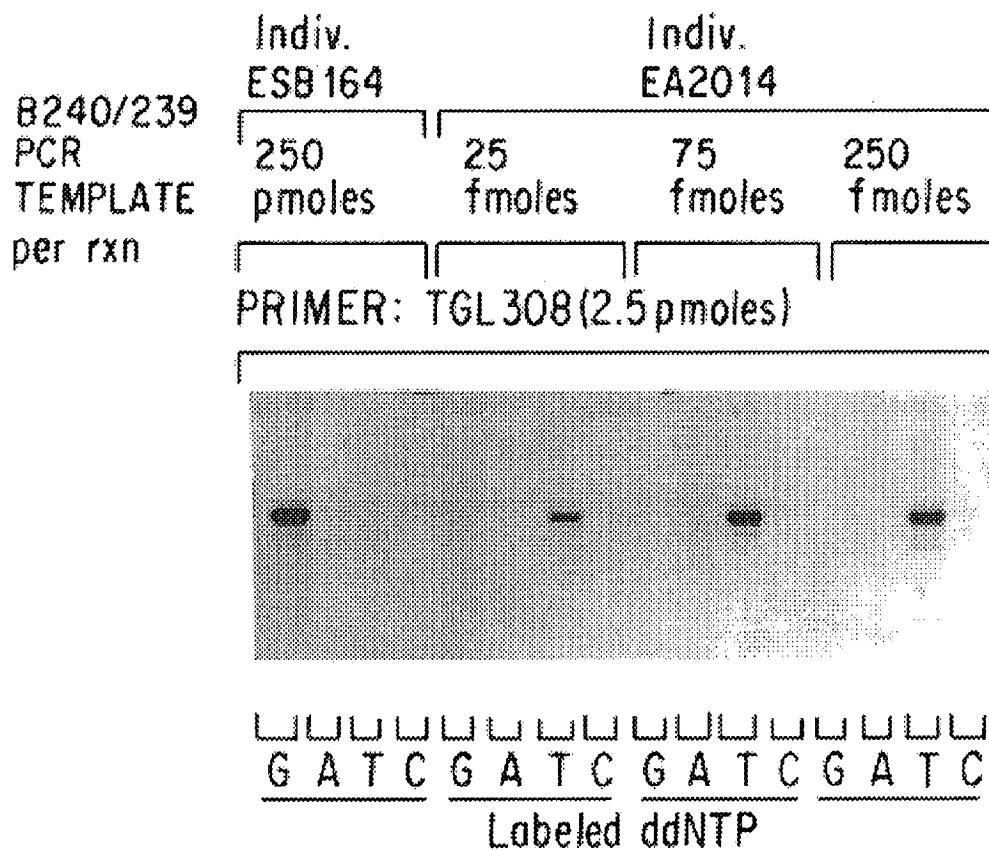


FIG.7

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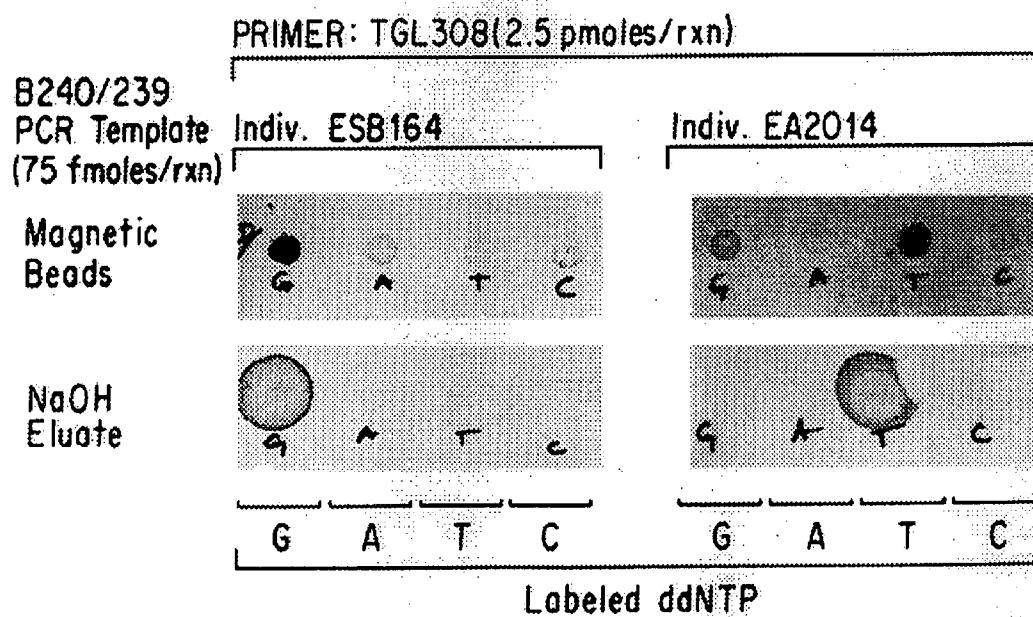


FIG.8

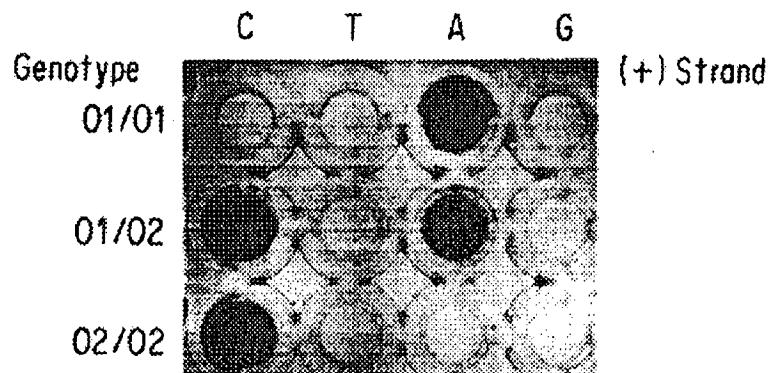
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GBA : HLA DPA1 aa 31



#517

GT~~T~~TATGTTGAATTGATGAAGATGAG AT  
CA

CAAGTACA~~AA~~ACTTAA~~ACT~~ACTTCTACTC TA  
GT

GT~~T~~TATGTTGAATTGATGAAGATGAG CAAGATA~~AC~~CTAGAC~~CT~~GTTCTTCCT

#518

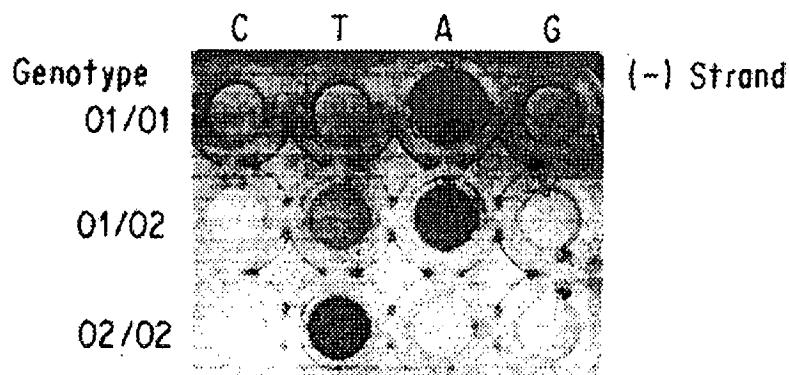


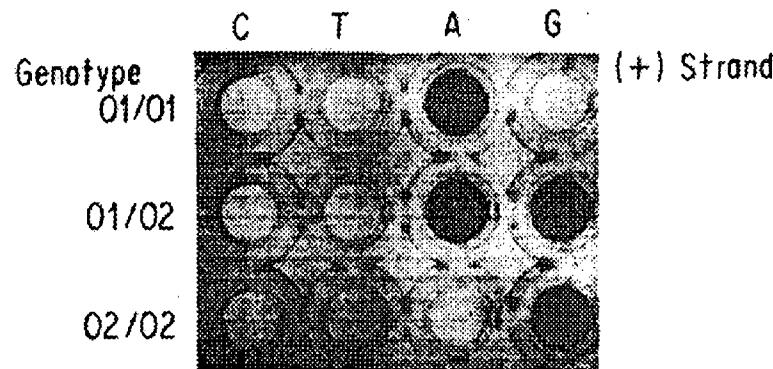
FIG.9

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GBA: HLA DPA1  $\alpha\alpha$ 50

#515 → A      ACCGTCTGGCATCTGGAGGAGTTGGCC  
G  
TGGCAGACCGTAGACCTCCTAAACCGG      T      TCGGAAAAGGAAACTCCGACTCCCGCCC  
C ← #513

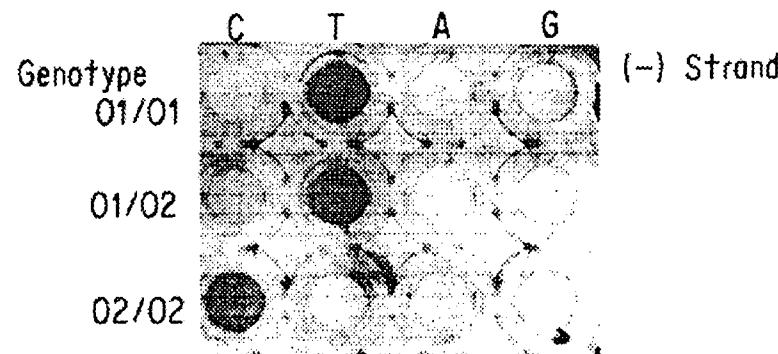


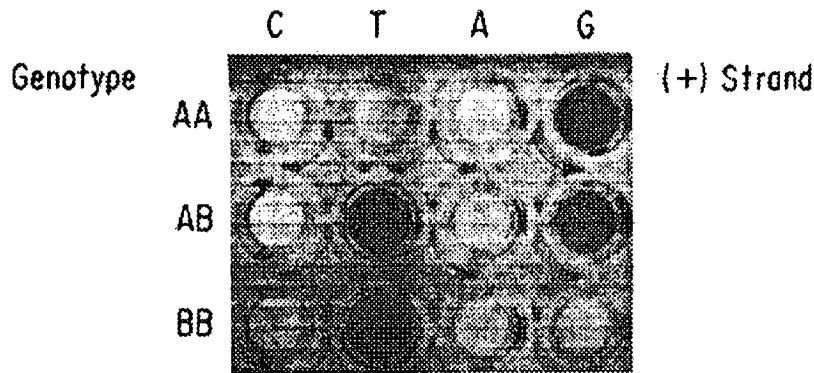
FIG.10

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GBA: Equine Locus JH85  
Nucleotide 122

#308 →  
 TGAAGCCTCAGACCAGCGTGGTGCCTGGT      G-  
     TC  
 ACTTCGGAGTCTGGCGCACACGGACCA      C-  
     AG ← #307

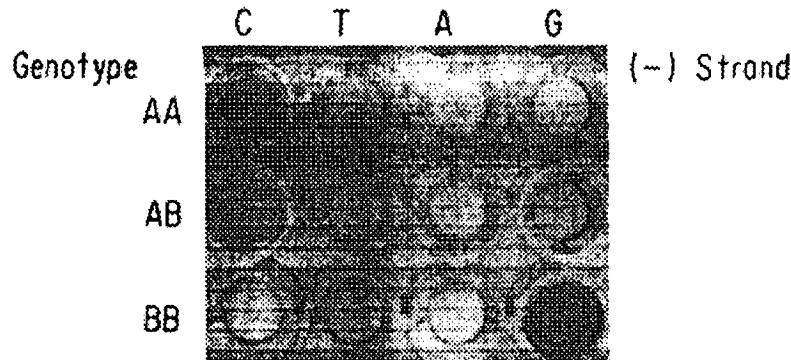


FIG. 11

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QUANTITATIVE GBA: EQUINE IOCUS JH85

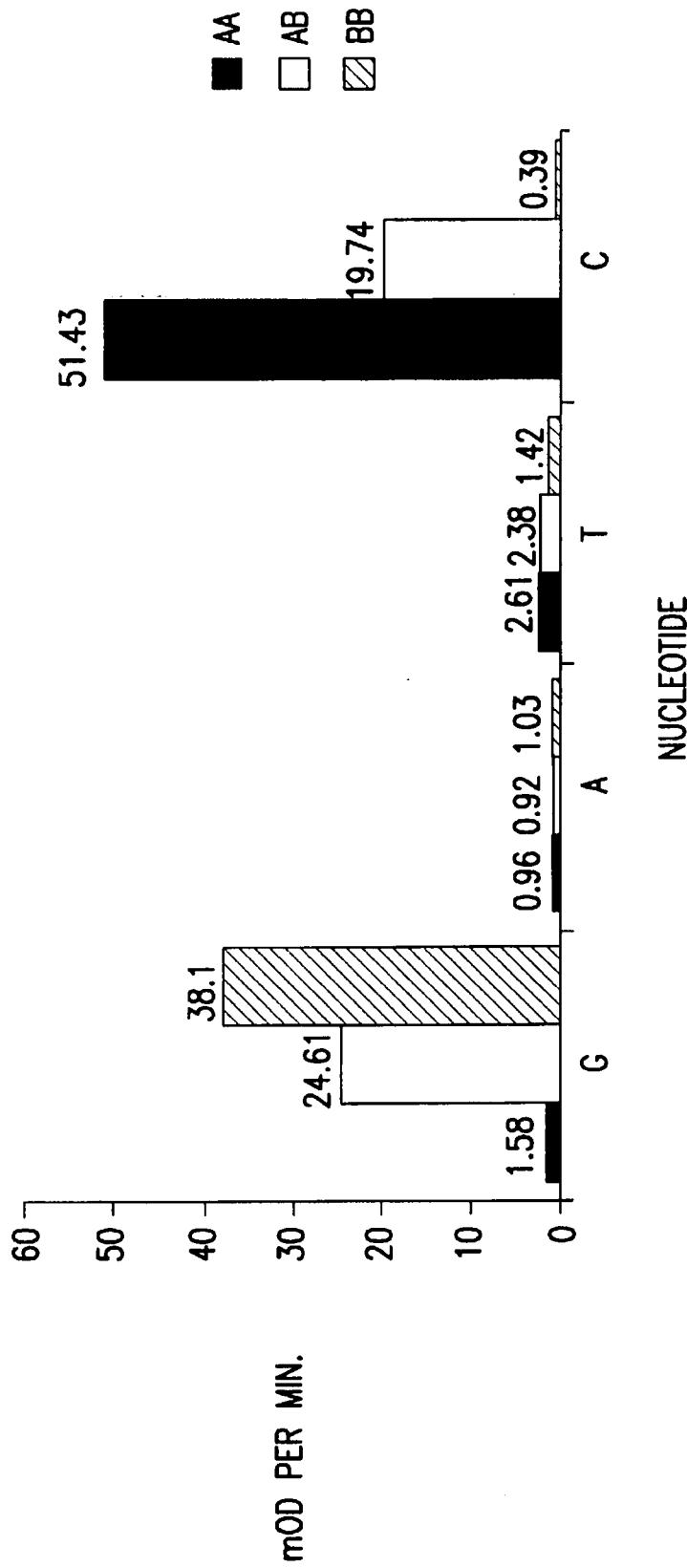


FIG. 12

6,004,744

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**METHOD FOR DETERMINING  
NUCLEOTIDE IDENTITY THROUGH  
EXTENSION OF IMMOBILIZED PRIMER**

This application is a continuation-in-part of U.S. Ser. No. 664,837 filed Mar. 5, 1991, now U.S. Pat. No. 5,888,819, the contents of which are hereby incorporated by reference into the present disclosure.

**BACKGROUND OF THE INVENTION**

This invention relates to the field of nucleic acid sequence detection. The detection of nucleic acid sequences can be used in two general contexts. First, the detection of nucleic acid sequences can be used to determine the presence or absence of a particular genetic element. Second, the detection of nucleic acid sequences can be used to determine the specific type of a particular genetic element that is present. Variant genetic elements usually exist. Many techniques have been developed (1) to determine the presence of specific nucleic acid sequences, and (2) to compare homologous segments of nucleic acid sequence to determine if the segments are identical or if they differ at one or more nucleotides. Practical applications of these techniques include genetic disease diagnoses, infectious disease diagnoses, forensic techniques, paternity determinations, and genome mapping.

In general, the detection of nucleic acids in a sample and the subtypes thereof depends on the technique of specific nucleic acid hybridization in which the oligonucleotide probe is annealed under conditions of high stringency to nucleic acids in the sample, and the successfully annealed probes are subsequently detected (see Spiegelman, S., *Scientific American*, Vol. 210, p. 48 (1964)).

The most definitive method for comparing DNA segments is to determine the complete nucleotide sequence of each segment. Examples of how sequencing has been used to study mutations in human genes are included in the publications of Engelke, et al., *Proc. Natl. Acad. Sci. U.S.A.*, 85:544-548 (1988) and Wong, et al., *Nature*, 330:384-386 (1987). At the present time, it is not practical to use extensive sequencing to compare more than just a few DNA segments because the effort required to determine, interpret, and compare sequence information is time-consuming.

A commonly used screen for DNA polymorphisms arising from DNA sequence variation consists of digesting DNA with restriction endonucleases and analyzing the resulting fragments by means of Southern blots, as described by Botstein, et al., *Am. J. Hum. Genet.*, 32:314-331 (1980) and White, et al., *Sci. Am.*, 258:40-48 (1988). Mutations that affect the recognition sequence of the endonuclease will preclude enzymatic cleavage at that site, thereby altering the cleavage pattern of that DNA. DNAs are compared by looking for differences in restriction fragment lengths. A major problem with this method (known as restriction fragment length polymorphism mapping or RFLP mapping) is its inability to detect mutations that do not affect cleavage with a restriction endonuclease. Thus, many mutations are missed with this method. One study, by Jeffreys, *Cell*, 18:1-18 (1979), was able to detect only 0.7% of the mutational variants estimated to be present in a 40,000 base pair region of human DNA. Another problem is that the methods used to detect restriction fragment length polymorphisms are very labor intensive, in particular, the techniques involved with Southern blot analysis.

A technique for detecting specific mutations in any segment of DNA is described in Wallace, et al., *Nucl. Acids*

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*Res.*, 9:879-894 (1981). It involves hybridizing the DNA to be analyzed (target DNA) with a complementary, labeled oligonucleotide probe. Due to the thermal instability of DNA duplexes containing even a single base pair mismatch, differential melting temperature can be used to distinguish target DNAs that are perfectly complementary to the probe from target DNAs that differ by as little as a single nucleotide. In a related technique, described in Landegren, et al., *Science*, 41:1077-1080 (1988), oligonucleotide probes are constructed in pairs such that their junction corresponds to the site on the DNA being analyzed for mutation. These oligonucleotides are then hybridized to the DNA being analyzed. Base pair mismatch between either oligonucleotide and the target DNA at the junction location prevents the efficient joining of the two oligonucleotide probes by DNA ligase.

**A. Nucleic acid hybridization**

The base pairing of nucleic acids in a hybridization reaction forms the basis of most nucleic acid analytical and diagnostic techniques. In practice, tests based only on parameters of nucleic acid hybridization function poorly in cases where the sequence complexity of the test sample is high. This is partly due to the small thermodynamic differences in hybrid stability, generated by single nucleotide changes, and the fact that increasing specificity by lengthening the probe has the effect of further diminishing this differential stability. Nucleic acid hybridization is, therefore, generally combined with some other selection or enrichment procedure for analytical and diagnostic purposes.

Combining hybridization with size fractionation of hybridized molecules as a selection technique has been one general diagnostic approach. Size selection can be carried out prior to hybridization. The best known prior size selection technique is Southern Blotting (see Southern, E., *Methods in Enzymology*, 69:152 (1980)). In this technique, a DNA sample is subjected to digestion with restriction enzymes which introduce double stranded breaks in the phosphodiester backbone at or near the site of a short sequence of nucleotides which is characteristic for each enzyme. The resulting heterogeneous mixture of DNA fragments is then separated by gel electrophoresis, denatured, and transferred to a solid phase where it is subjected to hybridization analysis *in situ* using a labeled nucleic acid probe. Fragments which contain sequences complementary to the labeled probe are revealed visually or densitometrically as bands of hybridized label. A variation of this method is Northern Blotting for RNA molecules. Size selection has also been used after hybridization in a number of techniques, in particular by hybrid protection techniques, by subjecting probe/nucleic acid hybrids to enzymatic digestion before size analysis.

**B. Polymerase extension of duplex primer:template complexes**

Hybrids between primers and DNA targets can be analyzed by polymerase extension of the hybrids. A modification of this methodology is the polymerase chain reaction in which the purification is produced by sequential hybridization reactions of anti-parallel primers, followed by enzymatic amplification with DNA polymerase (see Saiki, et al., *Science* 239:487-491 (1988)). By selecting for two hybridization reactions, this methodology provides the specificity lacking in techniques that depend only upon a single hybridization reaction.

It has long been known that primer-dependent DNA polymerases have, in general, a low error rate for the addition of nucleotides complementary to a template. This feature is essential in biology for the prevention of genetic

mistakes which would have detrimental effects on progeny. The specificity inherent in this enzymological reaction has been widely exploited as the basis of the "Sanger" or dideoxy chain termination sequencing methodology which is the ultimate nucleic acid typing experiment. One type of Sanger DNA sequencing method makes use of mixtures of the four deoxynucleoside triphosphates, which are normal DNA precursors, and one of the four possible dideoxynucleoside triphosphates, which have a hydrogen atom instead of a hydroxyl group attached to the 3' carbon atom of the ribose sugar component of the nucleotide. DNA chain elongation in the 5' to 3' direction ("downstream") requires this hydroxyl group. As such, when a dideoxynucleotide is incorporated into the growing DNA chain, no further elongation can occur. With one dideoxynucleotide in the mixture, DNA polymerases can, from a primer:template combination, produce a population of molecules of varying length, all of which terminate after the addition of one out of the four possible nucleotides. The series of four independent reactions, each with a different dideoxynucleotide, generates a nested set of fragments, all starting at the same 5' terminus of the priming DNA molecule and terminating at all possible 3' nucleotide positions.

Another utilization of dideoxynucleoside triphosphates and a polymerase in the analysis of DNA involves labeling the 3' end of a molecule. One prominent manifestation of this technique provides the means for sequencing a DNA molecule from its 3' end using the Maxam-Gilbert method. In this technique, a molecule with a protruding 3' end is treated with terminal transferase in the presence of radioactive dideoxy-ATP. One radioactive nucleotide is added, rendering the molecule suitable for sequencing. Both methods of DNA sequencing using labeled dideoxynucleotides require electrophoretic separation of reaction products in order to derive the typing information. Most methods require four separate gel tracks for each typing determination.

The following two patents describe other methods of typing nucleic acids which employ primer extension and labeled nucleotides. Mundy (U.S. Pat. No. 4,656,127) describes a method whereby a primer is constructed complementary to a region of a target nucleic acid of interest such that its 3' end is close to a nucleotide in which variation can occur. This hybrid is subject to primer extension in the presence of a DNA polymerase and four deoxynucleoside triphosphates, one of which is an  $\alpha$ -thionucleotide. The hybrid is then digested using an exonuclease enzyme which cannot use thio-derivatized DNA as a substrate for its nucleolytic action (for example Exonuclease III of *E. coli*). If the variant nucleotide in the template is complementary to one of the thionucleotides in the reaction mixture, the resulting extended primer molecule will be of a characteristic size and resistant to the exonuclease; hybrids without thio-derivatized DNA will be digested. After an appropriate enzyme digest to remove undervatized molecules, the thio-derivatized molecule can be detected by gel electrophoresis or other separation technology.

Vary and Diamond (U.S. Pat. No. 4,851,331) describes a method similar to that of Mundy wherein the last nucleotide of the primer corresponds to the variant nucleotide of interest. Since mismatching of the primer and the template at the 3' terminal nucleotide of the primer is counterproductive to elongation, significant differences in the amount of incorporation of a tracer nucleotide will result under normal primer extension conditions. This method depends on the use of a DNA polymerase, e.g., AMV reverse transcriptase, that does not have an associated 3' to 5' exonuclease activity.

The methods of Mundy and of Vary and Diamond have drawbacks. The method of Mundy is useful but cumbersome due to the requirements of the second, different enzymological system where the non-derivatized hybrids are digested. The method of Vary is complicated by the fact that it does not generate discrete reaction products. Any "false" priming will generate significant noise in such a system which would be difficult to distinguish from a genuine signal.

The present invention circumvents the problems associated with the methods of Mundy and of Vary and Diamond for typing nucleic acid with respect to particular nucleotides. With methods employing primer extension and a DNA polymerase, the current invention will generate a discrete molecular species one base longer than the primer itself. In many methods, particularly those employing the polymerase chain reaction, the type of reaction used to purify the nucleic acid of interest in the first step can also be used in the subsequent detection step. Finally, with terminators which are labeled with different detector moieties (for example different fluorophors having different spectral properties), it will be possible to use only one reagent for all sequence detection experiments. Furthermore, if techniques are used to separate the terminated primers post-reaction, sequence detection experiments at more than one locus can be carried out in the same tube.

A recent article by Mullis (*Scientific American*, April 1990, pp. 56-65) suggests an experiment, which apparently was not performed, to determine the identity of a targeted base pair in a piece of double-stranded DNA. Mullis suggests using four types of dideoxynucleosides triphosphate, with one type of dideoxynucleoside triphosphate being radioactively labeled.

The present invention permits analyses of nucleic acid sequences that can be useful in the diagnosis of infectious diseases, the diagnosis of genetic disorders, and in the identification of individuals and their parentage.

A number of methods have been developed for these purposes. Although powerful, such methodologies have been cumbersome and expensive, generally involving a combination of techniques such as gel electrophoresis, blotting, hybridization, and autoradiography or non-isotopic revelation. Simpler technologies are needed to allow the more widespread use of nucleic acid analysis. In addition, tests based on nucleic acids are currently among the most expensive of laboratory procedures and for this reason cannot be used on a routine basis. Finally, current techniques are not adapted to automated procedures which would be necessary to allow the analysis of large numbers of samples and would further reduce the cost.

The current invention provides a method that can be used to diagnose or characterize nucleic acids in biological samples without recourse to gel electrophoretic size separation of the nucleic acid species. This feature renders this process easily adaptable to automation and thus will permit the analysis of large numbers of samples at relatively low cost. Because nucleic acids are the essential blueprint of life, each organism or individual can be uniquely characterized by identifiable sequences of nucleic acids. It is, therefore, possible to identify the presence of particular organisms or demonstrate the biological origin of certain samples by detecting these specific nucleic acid sequences.

#### SUMMARY OF THE INVENTION

The subject invention provides a reagent composition comprising an aqueous carrier and an admixture of at least two different terminators of a nucleic acid template-

dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the unpaired nucleotide base in the template immediately adjacent to, and downstream of, the 3' end of the primer. In addition, at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and one, two, three, or four of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent as described above wherein the terminators comprise nucleotides, nucleotide analogs, dideoxynucleotides, or arabinoside triphosphates. The subject invention also provides a reagent wherein the terminators comprise one or more of dideoxyadenosine triphosphate (ddATP), dideoxycytosine triphosphate (ddCTP), dideoxyguanosine triphosphate (ddGTP), dideoxythymidine triphosphate (ddTTP), or dideoxyuridine triphosphate (ddUTP).

The subject invention also provides a method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with a stretch of nucleotide bases present in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest. Enzymatic extension of the oligonucleotide primer in the resultant duplex by one nucleotide, catalyzed, for example, by a DNA polymerase, thus depends on correct base pairing of the added nucleotide to the nucleotide base to be identified.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. The net result is that the oligonucleotide primer has been extended by one terminator. Next, the identity of the detectable marker present at the 3' end of the extended primer is determined. The identity of the detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

The subject invention also provides another method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. This additional method uses a reagent containing four terminators, only one of the terminators having a detectable marker.

The subject invention also provides a method of typing a sample of nucleic acids which comprises identifying the base or bases present at each of one or more specific positions, each such nucleotide base being identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. Each specific position in the nucleic acid of interest is determined using a different primer. The identity of each nucleotide base or bases at each position can be determined individually or the identities of the nucleotide bases at different positions can be determined simultaneously.

The subject invention further provides a method for identifying different alleles in a sample containing nucleic acids which comprises identifying the base or bases present at each of one or more specific positions. The identity of each nucleotide base is determined by the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention also provides a method for determining the genotype of an organism at one or more particular genetic loci which comprises obtaining from the organism a sample containing genomic DNA and identifying the nucleotide base or bases present at each of one or more specific positions in nucleic acids of interest. The identity of each such base is determined by using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. The identities of the nucleotide bases determine the different alleles and, thereby, determine the genotype of the organism at one or more particular genetic loci.

#### BRIEF DESCRIPTION OF THE FIGURES

FIGS. 1A-1C. Autoradiography of labeled DNA products after fractionation on a polyacrylamide/urea gel. Panel A shows products of the "A" extension reaction on oligonucleotide primer 182 directed by template oligonucleotides 180 or 181. Panel B shows products of the "B" termination reaction on oligonucleotide primer 182 annealed to template oligonucleotides 180 or 181. Panel C shows the same products as in panel B after purification on magnetic beads. Note: oligodeoxynucleotide 182 was used as supplied by Midland Certified Reagents with no further purification. The minor bands above and below the main band are presumably contaminants due to incomplete reactions or side reactions that occurred during the step-wise synthesis of the oligonucleotide. For a definition of the "A" extension reaction and the "B" termination reaction, see "A. GENERAL METHODS" in the Detailed Description of the Invention.

FIG. 2. Detection of Sequence Polymorphisms in PCR Products. Target polymorphic DNA sequence showing amplification primers [SEQ ID NO:15] and [SEQ ID NO:16], detection primers [SEQ ID NO:17] and [SEQ ID NO:18], and molecular clone (plasmid) designations. For each primer, sites of binding to one or the other strand of the target DNA sequence [SEQ ID NO:19] are indicated by underlining, and the direction of DNA synthesis is indicated by an arrow. Numbering for the target sequence is shown in the righthand margin. Polymorphic sites at positions 114 and 190 are indicated by bold lettering and a slash between the two polymorphic possibilities.

FIG. 3. Autoradiogram of gel-analyzed polymorphism test on PCR products. Templates from PCR products of p183, p624, or p814 were analyzed with the detection primers, TGL182 and TGL166, in a template-directed chain extension experiment, as described in the specification.

Reaction products were fractionated by size on a polyacrylamide/urea DNA sequencing gel, and incorporation of [<sup>35</sup>S]- $\alpha$ -thio-dideoxy adenosine monophosphate was assayed by autoradiography.

FIG. 4. Gel electrophoretic analysis of the labelled extension products of primers TGL346 and TGL391. Productive primer-template complexes of TGL346 or TGL391 with the bead-bound oligonucleotide template, TGL382, were subjected to primer extension labelling reactions with the four different [ $\alpha$ -thio-<sup>35</sup>S]dideoxynucleoside triphosphate mixes. Labelled primer DNA was released from the washed beads and electrophoresed on an 8% polyacrylamide/8 M urea DNA sequencing gel (2.5 pmoles of primer/lane), then analyzed by autoradiography. The four lanes shown for the primer TGL346 indicate that labelling occurred predominantly with the ddC mix, indicating that the next unpaired base in the TGL382 template adjacent to the 3' end of TGL346 was a G (see sequence given in Example 4). The four lanes shown for the primer TGL391 indicate that the labelling occurred predominantly with the ddT mix, indicating that the next unpaired base in the TGL382 template adjacent to the 3' end of TGL391 was an A.

FIG. 5. Autoradiographic analyses of total radioactivity bound to beads. The bead suspensions, containing the products of the extension reactions described in FIG. 5, were spotted onto filter paper (1 pmole of primer per spot) and exposed to X-ray film to assay total bead-bound radioactivity. As shown, TGL346 predominantly incorporated label from the ddC mix and TGL391 predominantly from the ddT mix.

FIG. 6. PCR-amplified polymorphic locus of mammalian DNA. Shown is a 327 basepair segment of mammalian DNA that was amplified from samples of genomic DNA using the PCR primers TGL240 [SEQ ID NO:22] (biotinylated) and TGL239 (unbiotinylated). Samples of DNA from two homozygous individuals, ESB164 (genotype AA) and EA2014 (genotype BB), were subjected to the analyses described in Example 5. The complete DNA sequence [SEQ ID NO:20] of the A allele at this locus is shown, with the polymorphic sites where the B allele sequence differs from the A allele sequence indicated by the bases underneath the A sequence. The detection primer, TGL308 [SEQ ID NO:21], is shown base-paired with the template strand extending from the biotinylated primer. For the A allele, the first unpaired template base immediately downstream of the 3' end of TGL308 is a C, and for the B allele this base is an A. Thus, the A allele should result in labelling of TGL308 by the ddG mix only, and the B allele should result in labelling by the ddT mix only.

FIG. 7. Gel electrophoretic analysis of PCR products from two different homozygous individuals. Primers TGL240 and TGL239 were used to amplify genomic DNA (obtained from blood) from two individuals, ESB164 and EA2014. The products of the extension reactions for primer TGL308, annealed to the bead-bound, PCR-generated template as outlined in FIG. 7, were analyzed by electrophoresis on an 8% polyacrylamide/8 M urea DNA sequencing gel as outlined in FIG. 5. Shown for individual ESB164 (genotype AA: labelling expected from the ddG mix) are 250 fmole of extended primer from the four different ddNTP labelling reactions. Shown for individual EA2014 (genotype BB: labelling expected from the ddT mix) are loadings of 25, 75, and 250 fmole of extended primer from the four different ddNTP labelling reactions.

FIG. 8. Autoradiographic analyses of total and NaOH-eluted radioactivity from TGL308 primer extension reac-

tions. Primer TGL308 was used to analyze the genotypes of individuals ESB164 and EA2014 as outlined in Example 5 and FIGS. 7 and 8. Total bead-associated radioactivity was determined by directly spotting a suspension of beads containing 75 fmole of primer onto filter paper followed by autoradiographic detection of the label in the spot. Radioactivity specifically associated with the TGL308 primer was determined by magnetically immobilizing the beads, eluting the primer with NaOH as described in Examples 4 and 5, and spotting on filter paper an amount corresponding to 75 fmole. Label in these spots was also detected by autoradiography.

FIG. 9. Data is shown from GBA on single stranded nucleic acid produced by asymmetric PCR from human DNA samples of different genotypes. The DNA sequence being interrogated [SEQ ID NO:23] is from the HLA DPA1 locus at the polymorphic sequence coding for amino acid 31 of the DP alpha chain (Marsh, S. G. E. and Bodmer, J. G., HLA Class II Nucleotide Sequences, 1991. Human Immunol. 31, 207-227 [1991]) and is shown in the middle of the figure. Identification of the nucleotide immediately downstream of the primer is accomplished by enzyme-linked detection and is visualized as an orange color change in the well corresponding to the nucleotide which is inserted by the T7 DNA polymerase. Homozygotes only have one positive well, heterozygotes have two. The sequence of the GBA primer [SEQ ID NO:24] is indicated by an arrow whose tail is the 5' end and head is the 3' end of the oligonucleotide.

FIG. 10. Data is shown from GBA on single stranded nucleic acid produced by asymmetric PCR from equine DNA samples of different genotypes. The DNA sequence being interrogated [SEQ ID NO:25] is from the HLA DPA1 locus at the polymorphic sequence [SEQ ID NO:26] coding for amino acid 50 of the DP alpha chain (Marsh, S. G. E. and Bodmer, J. G., HLA Class II Nucleotide Sequences, 1991. Human Immunol. 31, 207-227 [1991]) and is shown in the middle of the Figure.

FIG. 11. Data is shown from GBA on single stranded nucleic acid produced by asymmetric PCR from equine DNA samples of different genotypes. The DNA sequence being interrogated is from the anonymous locus JH85 at the polymorphic sequence at nucleotide number 122 with respect to the original cloned genomic piece (unpublished results) and is shown in the middle of the figure. At this position, the "B" allele contains one extra base. For this reason, a different nucleotide position is interrogated by primer #307 [SEQ ID NO:27] as compared to #308 [SEQ ID NO:28]. Nevertheless, the results of both strand interrogations allow for unambiguous typing.

FIG. 12. Data shown are the results of a quantitative GBA of equine locus JH85. Following addition of substrate, the microplate was read kinetically, in a "Vmax" model 96-well spectrophotometer (Molecular Devices, Inc., Menlo Park, Calif.). Values are expressed as a Vmax in milli OD units per minute. The GBA results for the AA homozygote (solid bars), the AB heterozygote (open bars), and BB homozygote (stippled bars) single stranded templates is indicated for the four biotinylated ddNTPs analyzed in separate wells. Numerical values obtained are indicated at the top of each bar.

#### DETAILED DESCRIPTION OF THE INVENTION

The subject invention provides a reagent composition comprising an aqueous carrier and an admixture of at least two different terminators of a nucleic acid template-

dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the unpaired nucleotide base in the template immediately adjacent to, and downstream of, the 3' end of the primer. In addition, at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and two, three, or four of the terminators are labeled with a different detectable marker.

The subject invention further provides a reagent as described above wherein the terminators comprise nucleotides, nucleotide analogs, dideoxynucleotides, or arabinoside triphosphates. The subject invention also provides a reagent wherein the terminators comprise one or more of dideoxyadenosine triphosphate (ddATP), dideoxycytosine triphosphate (ddCTP), dideoxyguanosine triphosphate (ddGTP), dideoxythymidine triphosphate (ddTTP), or dideoxyuridine triphosphate (ddUTP).

The subject invention further provides a reagent as described above wherein each of the detectable markers attached to the terminators is an isotopically labeled moiety, a chromophore, a fluorophore, a protein moiety, or a moiety to which an isotopically labeled moiety, a chromophore, a fluorophore, or a protein moiety can be attached. The subject invention also provides a reagent wherein each of the different detectable markers is a different fluorophore.

The subject invention also provides a reagent as described above wherein the reagent further comprises pyrophosphatase.

The invented reagent consists of two or more chain terminators with one or more of the chain terminators being identifiably tagged. This reagent can be used in a DNA polymerase primer extension reaction to type nucleic acid sequences of interest that are complementary to one or more oligonucleotide primers by chemically or physically separating the polymerase extended primers from the chain terminator reagent and analyzing the terminal additions. Any kind of terminator that inhibits further elongation can be used, for example, a dideoxynucleoside triphosphate. Several approaches can be used for the labeling and detection of terminators: (1) radioactivity and its detection by either autoradiography or scintillation counting, (2) fluorescence or absorption spectroscopy, (3) mass spectrometry, or (4) enzyme activity, using a protein moiety. The identity of each terminator can be determined individually, i.e., one at a time. In addition, methods which permit independent analyses of each of the terminators permit analysis of incorporation of up to four terminators simultaneously.

The subject invention also provides a method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest

is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with a stretch of nucleotide bases present in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest. Enzymatic extension of the oligonucleotide primer in the resultant duplex by one nucleotide, catalyzed, for example, by a DNA polymerase, thus depends on correct base pairing of the added nucleotide to the nucleotide base to be identified.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer.

The net result is that the oligonucleotide primer has been extended by one terminator. Next, the identity of the detectable marker present at the 3' end of the extended primer is determined. The identity of the detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

The subject invention also provides another method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with nucleotide bases in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four terminators, only one of the terminators having a detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. The net result is that the oligonucleotide primer has been extended by one terminator.

The original duplex of primer and the nucleic acid of interest is then contacted with three different reagents, with a different one of each of the four terminators being labeled in each of the four parallel reaction steps. Next, the products

of the four parallel template-dependent, primer extension reactions are examined to determine which of the products has a detectable marker. The product with a detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

Both of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest label the primer after hybridization between the primer and the template. If the template-dependent enzyme has no exonuclease function, the 3' end of the primer must be base paired for the labeling by a terminator to occur.

The subject invention also provides a method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids. First, the sample of nucleic acids is treated, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids. If the nucleic acids in the sample are single-stranded, this step is not necessary. Second, the sample of nucleic acids is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with the particular nucleotide sequence, if the particular nucleotide sequence is present, so as to form a duplex between the primer and the particular nucleotide sequence.

The duplex of primer and the particular nucleotide sequence, if any, is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the particular nucleotide sequence, if any, is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the unpaired template nucleotide base downstream of the 3' end of the primer, the primer being hybridized with the particular nucleotide sequence in the template, and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. Next, the absence or presence and identity of a detectable marker at the 3' end of the primer are determined. The presence or absence of the detectable marker indicates whether the primer has hybridized to the template. If a detectable marker is absent, the primer did not hybridize to the template, and, therefore, the particular nucleotide sequence is not present in the sample of nucleic acids. If a detectable marker is present, the primer did hybridize to the template, and, therefore, the particular nucleotide sequence is present in the sample of nucleic acids.

The subject invention also provides another method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids. First, the sample of nucleic acids is treated, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids. Second, the sample of nucleic acids is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with the particular nucleotide sequence, if the particular nucleotide sequence is present, so as to form a duplex between the primer and the particular nucleotide sequence.

The duplex of primer and the particular nucleotide sequence, if any, is then contacted with a reagent containing four terminators, only one of the terminators having a detectable marker. The duplex of primer and the particular nucleotide sequence, if any, is contacted with the reagent

under conditions permitting base pairing of a complementary terminator present in the reagent with the unpaired template nucleotide base downstream of the 3' end of the primer, the primer being hybridized with the particular nucleotide sequence in the template, and the occurrence of a template-dependent, primer extension reaction. The net result is the incorporation of the terminator at the 3' end of the primer.

The original duplex of primer and the particular nucleotide sequence, if any, is then contacted with three different reagents, with a different one of each of the four terminators being labeled in each of the four parallel reaction steps. Next, the products of the four parallel, template-dependent, primer extension reactions are examined to determine which, if any, of the products have detectable markers. The absence or presence and identity of the detectable marker indicates whether the primer has hybridized to the template. If no detectable marker is present in any of the products, the primer did not hybridize to the template, and, therefore, the particular nucleotide sequence was not present in the sample of nucleic acids. If a detectable marker is present in any of the products, the primer did hybridize to the template, and, therefore, the particular nucleotide sequence was present in the sample of nucleic acids.

Different versions of the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest and the method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids are possible. In the first version, the template is a deoxyribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a DNA polymerase. This version gives a DNA product. In a second version, the template is a ribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a reverse transcriptase. This version gives a DNA product. In a third version, the template is a deoxyribonucleic acid, the primer is an oligoribonucleotide, and the enzyme is an RNA polymerase. This version gives an RNA product. In a fourth version, the template is a ribonucleic acid, the primer is an oligoribonucleotide, and the template-dependent enzyme is an RNA replicase. This version gives an RNA product.

Preferably, before the primer extension reaction is performed, the template is capped by the addition of a terminator to the 3' end of the template. The terminator is capable of terminating a template-dependent, primer extension reaction. The template is capped so that no additional labeled terminator will attach at the 3' end of the template. The extension reaction should occur on the primer, not on the template. A dideoxynucleotide can be used as a terminator for capping the template.

Another modification of the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest is to separate the primer from the nucleic acid of interest after the extension reaction by using appropriate denaturing conditions. The denaturing conditions can comprise heat, alkali, formamide, urea, glyoxal, enzymes, and combinations thereof. The denaturing conditions can also comprise treatment with 2.0 N NaOH.

The nucleic acid of interest can comprise non-natural nucleotide analogs such as deoxyinosine or 7-deaza-2'-deoxyguanosine. These analogues destabilize DNA duplexes and could allow a primer annealing and extension

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reaction to occur in a double-stranded sample without completely separating the strands.

The sample of nucleic acids can be from any source. The sample of nucleic acids can be natural or synthetic (i.e., synthesized enzymatically in vitro). The sample of nucleic acids can comprise deoxyribonucleic acids, ribonucleic acids, or copolymers of deoxyribonucleic acid and ribonucleic acid. The nucleic acid of interest can be a deoxyribonucleic acid, a ribonucleic acid, or a copolymer of deoxyribonucleic acid and ribonucleic acid. The nucleic acid of interest can be synthesized enzymatically in vivo, synthesized enzymatically in vitro, or synthesized non-enzymatically. The sample containing the nucleic acid or acids of interest can comprise genomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof. The sample containing the nucleic acid or acids of interest can also comprise extragenomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof. Also, the nucleic acid or acids of interest can be synthesized by the polymerase chain reaction.

The sample can be taken from any organism. Some examples of organisms to which the method of the subject invention is applicable include plants, microorganisms, viruses, birds, vertebrates, invertebrates, mammals, human beings, horses, dogs, cows, cats, pigs, or sheep.

The nucleic acid of interest can comprise one or more moieties that permit affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer. The nucleic acid of interest can comprise biotin which permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via binding of the biotin to streptavidin which is attached to a solid support. The sequence of the nucleic acid of interest can comprise a DNA sequence that permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via base pairing to a complementary sequence present in a nucleic acid attached to a solid support. The nucleic acid of interest can be labeled with a detectable marker; this detectable marker can be different from any detectable marker present in the reagent or attached to the primer.

The oligonucleotide primer can be an oligodeoxyribonucleotide, an oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides. The oligonucleotide primer can be either natural or synthetic. The oligonucleotide primer can be synthesized either enzymatically in vivo, enzymatically in vitro, or non-enzymatically in vitro. The oligonucleotide primer can be labeled with a detectable marker; this detectable marker can be different from any detectable marker present in the reagent or attached to the nucleic acid of interest. In addition, the oligonucleotide primer must be capable of hybridizing or annealing with nucleotides present in the nucleic acid of interest, immediately adjacent to, and upstream of, the nucleotide base to be identified. One way to accomplish the desired hybridization is to have the template-dependent primer be substantially complementary or fully complementary to the known base sequence immediately adjacent to the base to be identified.

The oligonucleotide primer can comprise one or more moieties that permit affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest. The oligonucleotide primer can comprise biotin which permits affinity separation of the primer from the unincorporated reagent and/or nucleic acid of interest via binding of

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the biotin to streptavidin which is attached to a solid support. The sequence of the oligonucleotide primer can comprise a DNA sequence that permits affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest via base pairing to a complementary sequence present in a nucleic acid attached to a solid support.

The subject invention also provides a method of typing a sample of nucleic acids which comprises identifying the base or bases present at each of one or more specific positions, each such nucleotide base being identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. Each specific position in the nucleic acid of interest is determined using a different primer. The identity of each nucleotide base or bases at each position can be determined individually or the identities of the nucleotide bases at different positions can be determined simultaneously.

The subject invention also provides another method of typing a sample of nucleic acids which comprises determining the presence or absence of one or more particular nucleotide sequences, the presence or absence of each such nucleotide sequence being determined using one of the methods for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids as outlined above.

The subject invention also provides an additional method of typing a sample containing nucleic acids. First, the presence or absence of one or more particular nucleotide sequences is determined; the presence or absence of each such nucleotide sequence is determined using one of the methods for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids as outlined above. Second, the nucleotide base or bases present at each of one or more specific positions is identified; each such base is identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention further provides a method for identifying different alleles in a sample containing nucleic acids which comprises identifying the base or bases present at each of one or more specific positions. The identity of each nucleotide base is determined by the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention also provides a method for determining the genotype of an organism at one or more particular genetic loci which comprises obtaining from the organism a sample containing genomic DNA and identifying the nucleotide base or bases present at each of one or more specific positions in nucleic acids of interest. The identity of each such base is determined by using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. The identity of the nucleotide bases determine the different alleles and, thereby, determine the genotype of the organism at one or more particular genetic loci.

The chain termination reagent in combination with an appropriate oligonucleotide primer, and a DNA polymerase with or without an associated 3' to 5' exonuclease function, and an appropriate salt and cofactor mixture, can be used under appropriate hybridization conditions as a kit for diagnosing or typing nucleic acids, if appropriate primer separation techniques are used. To simplify the primer separation and the terminal nucleotide addition analysis this invention makes use of oligonucleotides that are modified in

such ways that permit affinity separation as well as polymerase extension. The 5' termini and internal nucleotides of synthetic oligonucleotides can be modified in a number of different ways to permit different affinity separation approaches, e.g., biotinylation. These affinity reagents can be used with the terminator mixture to facilitate the analysis of extended oligonucleotide(s) in two ways:

- (1) If a single affinity group is used on the oligonucleotide(s), the oligonucleotide(s) can be separated from the unincorporated terminator reagent. This eliminates the need of physical or size separation.
- (2) More than one oligonucleotide can be separated from the terminator reagent and analyzed simultaneously if more than one affinity group is used. This permits the analysis of several nucleic acid species or more nucleic acid sequence information per extension reaction.

The affinity group(s) need not be on the priming oligonucleotide but could, alternatively, be present on the template. As long as the primer remains hydrogen bonded to the template during the affinity separation step, this will allow efficient separation of the primer from unincorporated terminator reagent. This also has the additional benefit of leaving sites free on the primer for the convenient attachment of additional moieties. For example, the 5'-terminus of the primer could be modified by coupling it to a suitable fluorescent group such as rhodamine, allowing the amount of primer in the primer:template complex to be easily quantified after the affinity separation step. The amounts of 3'-terminating terminators could then be normalized to the total amount of annealed primer.

The oligonucleotide primers and template can be any length or sequence, can be DNA or RNA, or any modification thereof. It is necessary, however, that conditions are chosen to optimize stringent hybridization of the primers to the target sequences of interest.

The conditions for the occurrence of the template-dependent, primer extension reaction can be created, in part, by the presence of a suitable template-dependent enzyme. Some of the suitable template-dependent enzymes are DNA polymerases. The DNA polymerase can be of several types. The DNA polymerase must, however, be primer and template dependent. For example, *E. coli* DNA polymerase I or the "Klenow fragment" thereof, T4 DNA polymerase, T7 DNA polymerase ("Sequenase"), *T. aquaticus* DNA polymerase, or a retroviral reverse transcriptase can be used. RNA polymerases such as T3 or T7 RNA polymerase could also be used in some protocols. Depending upon the polymerase, different conditions must be used, and different temperatures ranges may be required for the hybridization and extension reactions.

The reagents of the subject invention permit the typing of nucleic acids of interest by facilitating the analysis of the 3' terminal addition of terminators to a specific primer or primers under specific hybridization and polymerase chain extension conditions. Using only the terminator mixture as the nucleoside triphosphate substrate ensures addition of only one nucleotide residue to the 3' terminus of the primer in the polymerase reaction. Using all four terminators simultaneously ensures fidelity, i.e., suppression of misreading.

By specifically labeling one or more of the terminators, the sequence of the extended primer can be deduced. In principle, more than one reaction product can be analyzed per reaction if more than one terminator is specifically labeled.

By specifically tagging the oligonucleotide primer(s), or template(s) with a moiety that does not affect the 3' extension reaction yet permits affinity separation, the extension

product(s) can be separated post-reaction from the unincorporated terminators, other components of the reagents, and/or the template strand. Several oligonucleotides can be analyzed per extension reaction if more than one affinity agent is used.

In principle, the combination of four differently labeled terminators and many primers or templates tagged with different groups permits the typing of many different nucleic acid sequences simultaneously.

Specificity in this diagnostic reaction is determined by (1) the stringency of oligonucleotide hybridization and (2) the sequence information gained by the single residue extension.

#### A. GENERAL METHODS

##### 1. Biotinylation of oligodeoxynucleotides

Oligodeoxynucleotides, terminated at their 5'-ends with a primary amino group, were ordered from Midland Certified Reagents, Midland, Tex. These were biotinylated using biotin-XX-NHS ester (Clontech Laboratories, Inc., Palo Alto, Calif.), a derivative of biotin-N-hydroxysuccinimide. Reagents used were from the Clontech biotinylation kit. Typically, the oligonucleotide (9 nanomoles) was dissolved in 100  $\mu$ l of 0.1M NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> (pH 9), and 25  $\mu$ l of N,N-dimethylformamide containing 2.5 mg biotin-XX-NHS-ester was added. The mixture was incubated overnight at room temperature. It was then passed over a 6 ml Sephadex G-25 column ("DNA grade"—Pharmacia) equilibrated with H<sub>2</sub>O. Eluate fractions containing DNA were identified by mixing 4  $\mu$ l aliquots with an equal volume of ethidium bromide (2  $\mu$ g/ml) and the DNA-induced fluorescence was monitored with a UV transilluminator. Unreacted ester was detected by UV absorption at 220 nm. The tubes containing DNA were pooled, concentrated in a Centricon-3 microconcentrator (Amicon), and passed over Sephadex again.

Inhibition of the binding of [<sup>3</sup>H]-biotin to magnetic M-280 streptavidin Dynabeads (Dynal) was used to assay quantitatively the extent of biotinylation of the oligonucleotides. Eppendorf tubes and pipet tips were siliconized. A known amount (5–10 pmoles) of biotin-labeled oligonucleotide in 10  $\mu$ l 0.1M NaCl was added to tubes containing 25  $\mu$ l of 1:4 suspension of beads in 0.1M NaCl. The tubes were rotated for one hour on a Labquake shaker (Labindustries, Inc.). Increasing amounts of [<sup>3</sup>H]-biotin (5–35 pmoles) in 20  $\mu$ l of 0.1M NaCl were added to the tubes and these were rotated again for one hour. Tubes were put on a Dynal MPC-E magnet to remove the beads from suspension, 10  $\mu$ l aliquots of the supernatant were withdrawn, and the amount of radioactivity in these was measured using a Beckman LS 5000 TD liquid scintillation counter. Counts were compared to those from tubes to which no oligonucleotide had been added. Alternatively, for some primers, biotinylation was monitored by size fractionation of the reaction products using analytical polyacrylamide gel electrophoresis in the presence of 8 M urea.

##### 2. Template-dependent primer extension/termination reactions

Approximately five pmoles of 5'-biotinylated oligodeoxynucleotide template (see above) were mixed with approximately three pmoles of primer in 1 $\times$  sequencing buffer (from Sequenase Version 2.0 kit, US Biochemical Corp.) (10  $\mu$ l final volume), the mixture was incubated at 65° C. for 2 min, then allowed to cool to room temperature in order to anneal the primer and template. The solution containing the annealed template-primer was separated into two 5  $\mu$ l

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portions, A and B, to which were added the following: Reactions A (for normalizing template concentrations)—0.5  $\mu$ l of 100 mM dithiothreitol, 1  $\mu$ l each of 10  $\mu$ M dATP, dGTP, ddCTP, 0.5  $\mu$ l of “Mn buffer” (from Sequenase Version 2.0 kit, US Biochemical Corp.), 0.5  $\mu$ l of [ $^{35}$ S]- $\alpha$ -thio-dITP (10 mCi/ml, 1180 Ci/mmole) (Dupont-NEN), 1  $\mu$ l of Sequenase (1:8 dilution, US Biochemical Corp.); Reactions B (for template-specific labeling of primer 3'-ends)—same additions as in Reactions A except the nucleotides used were ddCTP, ddGTP, ddTTP, and [ $^{35}$ S]- $\alpha$ -thio-ddATP.

Reactions were for 5 min at 37° C. Control reactions omitting the primer or the Sequenase were also performed. Aliquots were removed and analyzed by electrophoresis on a 15% polyacrylamide, 8 M urea, DNA sequencing gel (see Maniatis, T., et al., *Molecular Cloning, a Laboratory Manual*, Cold Spring Harbor Laboratory (1982)). The gel was fixed in 10% methanol, 10% acetic acid, dried down onto Whatman's 3MM paper, and exposed to Kodak X-Omat AR film. Alternatively, for purposes of analyzing the products by liquid scintillation counting, the biotinylated template or template-primer was bound to an excess of M-280 streptavidin Dynabeads (Dynal) before or after the Sequenase reaction (see above, “1. Biotinylation of oligodeoxynucleotides”, for binding conditions). Beads were washed three times with 0.1 M NaCl to remove unincorporated label, then scintillation fluid was added and the radioactivity measured by liquid scintillation counting.

### 3. Generation of templates from polymerase chain reaction products

Polymerase chain reaction (PCR) reactions were carried out where one or the other of the amplification primers flanking the target stretch of DNA were biotinylated as described above. These primers (2  $\mu$ mol final concentration) and the target DNA (up to 1  $\mu$ g) were incubated with 2.5 units of Taq polymerase (Perkin Elmer/Cetus), 200  $\mu$ M each of dATP, dCTP, dGTP, and dITP, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl<sub>2</sub>, and 0.01% gelatin (Sigma). Reaction mixtures were overlayed with paraffin oil and incubated for 30 cycles in Perkin Elmer/Cetus thermocycler. Each cycle consisted of 1 min at 94° C., 2 min at 60° C., and 3 min at 72° C. Reaction products were purified by phenol/chloroform extraction and ethanol precipitation, then analyzed by ethidium bromide staining after electrophoresis on a polyacrylamide gel. The yield of duplex PCR product was typically about 10  $\mu$ g.

Approximately 5  $\mu$ g of this PCR product was incubated with gentle agitation for 60 min with 50  $\mu$ L of a suspension of prewashed M-280 Dynabeads in 0.1 M NaCl. The beads with the bound DNA (approximately 15 pmoles) were then incubated for 5 min at 25° C. with 0.15 M NaOH. Beads were washed once with 0.15 M NaOH to remove the unbiotinylated DNA strand, then washed three times with H<sub>2</sub>O. The beads were resuspended in H<sub>2</sub>O and the strand bound to the beads via the biotin-streptavidin link was used as template for further primer extension reactions.

## B. EXAMPLES

### Example 1

Primer oligo 182: [SEQ ID NO:1] 5'-GCCTTGCGTTGTAGAA-

Template oligos

180(C)/181(T): [SEQ ID NO:2] 3'-TCGGGTCGGAACCGAACATCTT<sup>c</sup>/TATAGACTA<sup>5'</sup>

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Oligonucleotides 180 and 181 were synthesized with primary amino groups attached to their 5' termini. These were coupled with biotin as described above. Oligonucleotide 182 was annealed as a primer and extension reactions “A” and “B” (see above) were carried out. The expected template-dependent 3'-terminal extensions to oligonucleotide 182 were as follows (“\*” preceding a nucleotide signifies a radioactive label):

| Template | Reaction A         | Reaction B |
|----------|--------------------|------------|
| 180      | -dG-*dT-dA-*dT-ddC | -ddG       |
| 181      | -dA-*dT-dA-*dT-ddC | -*ddA      |

Thus, in the “A” reactions, both template oligonucleotides will direct a radioactively-labelled five nucleotide extension of the primer; the amount of labeling should be proportional to the amount of productively primed template present in the reactions. In the “B” reactions, both templates will direct a one nucleotide extension of the primer, but only for template 181 should this result in labeling of the primer. The “B” reaction, therefore, is an example of template-directed, sequence-specific labeling of an oligonucleotide via DNA polymerase-catalyzed extension of a productive primer-template complex.

The reaction products were fractionated by size on a 15% polyacrylamide/8M urea sequencing gel and visualized by autoradiography. The results (FIG. 1) show that, as expected, the “A” reactions yield labeling and extension of both primers whereas the “B” reaction results in labeling that is strongly biased in favor of template 181. Panel C in FIG. 1 shows a gel analysis of the same reaction products as in Panel B, except the reaction products were first purified as described above using M-280 streptavidin Dynabeads.

### Example 2

The experiment described in Example 1 shows template-directed labeling of oligonucleotide primer 182 in which the labeling is specific with respect to oligonucleotides or other species that migrate similarly on a polyacrylamide gel. In order to assess more generally the template-directed specific labeling of oligonucleotide 182 with respect to all other labeled species, regardless of gel mobility, a direct measurement of incorporated radioactivity was performed. In this experiment, both reactions “A” and “B” were performed, reaction products were purified using Dynabeads, and total radioactivity in the aliquots was measured by liquid scintillation counting. This procedure assesses both misincorporation of label into other species and, in addition, the efficiency of the Dynabead washing procedure with respect to unincorporated nucleotides. As a practical matter, it would be of interest to minimize [SEQ ID NO:3] both sources of non-specific label in order to have a simple, non-gel-based, procedure for assessing specific, template-directed labeling of the primer. The results of [SEQ ID NO:4] directly counting the reaction products after washing on the magnetic beads are as follows (all results expressed as cpm of [ $^{35}$ S]): [SEQ ID NO:5]

6,004,744

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## Example 4

| Reaction         | Template 180 | Template 181 |
|------------------|--------------|--------------|
| A, complete      | 325,782      | 441,823      |
| A, no polymerase | 5,187        | 5,416        |
| A, no primer     | 4,351        | 12,386       |
| B, complete      | 5,674        | 176,291      |
| B, no polymerase | 2,988        | 1,419        |
| B, no primer     | 1,889        | 1,266        |

As can be seen from these results, specific template-directed labeling of primer 182 can also be determined by measuring the total radioactivity of the reaction products after washing with magnetic beads to remove unreacted nucleotides. The background in this experiment due to nonspecific label from all other sources was approximately 3-4% (compare templates 180 and 181 in the "B, complete" reaction). Control experiments ("no polymerase" and "no primer") showed that the bulk of the background label was probably contributed by unincorporated nucleotides that were not completely removed by the washing step. The "A, complete" reactions showed that, for both templates, productive template:primer complexes were present.

## Example 3

Two amplification primers, TGL 105 and TGL 106 (FIG. 2), were used to amplify a cloned stretch of bovine DNA containing two DNA sequence polymorphisms: a C or T at position 114 and an A or G at position 190 (FIG. 2). DNAs containing these polymorphisms were molecularly cloned and available on plasmids, as follows: plasmid p183, C114 and A190; plasmid p624, T114 and A190; plasmid p814, C114 and G190. Four PCR reactions with biotinylated primers were performed to amplify and purify specific strands of these plasmids for use as templates:

| Primers                                 | Plasmids      | Detection Primers |
|---|---------------|-------------------|
| 105 biotinylated,<br>106 unbiotinylated | p183 and p624 | TGL 182           |
| 105 unbiotinylated,<br>106 biotinylated | p183 and p814 | TGL 166           |

The duplex PCR products were bound to magnetic microspheres, denatured with NaOH, and the biotinylated strand purified as described above. Templates prepared with biotinylated TGL 105 were subjected to analysis by DNA sequencing with unbiotinylated primer TGL 106 in order to measure the amount of template present. Similarly, template prepared using biotinylated TGL 106 was analyzed by sequencing with unbiotinylated TGL 105.

Approximately equal amounts of template (2 pmoles) were annealed for 5 min at 65° C. to the polymorphism detection primers, TGL 182 and TGL 166 (see above and FIG. 2). These primers hydrogen-bond to the templates in a sequence-specific fashion such that their 3'-termini are adjacent to nucleotide positions 114 and 190, respectively (FIG. 2). Template-directed primer extension reactions (reaction "B" conditions) were carried out on these primer:template complexes in the presence of the four ddNTPs, one of which (ddATP) was labeled. The products of these extension reactions were analyzed by electrophoresis on a 15% polyacrylamide/8M urea gel followed by autoradiography (FIG. 3).

Primer oligo TGL391: 5' TGTGTTGCACAAAAGCA 3'  
 5 Primer oligo TGL346: 5' GTTTTGACAAAAGCAT 3'  
 Template oligo TGL382: 3' CACAAACGTGTTTCGTAGGA 5' -  
 biotin: (streptavidin-bead)

Oligonucleotide TGL382 was purchased from the Midland Certified Reagent Company, Midland, Tex. It was biotinylated using Midland Certified Reagent Company's "Biotin dX" reagent (a biotin derivative phosphoramidite) which is suitable for use in automated DNA synthesis in the 5' terminal nucleotide position. The biotinylated oligonucleotide was then purified by anion exchange HPLC. Streptavidin-conjugated M-280 Dynabeads were washed in TNET buffer (10 mM Tris-HCl, pH 7.5, 100 mM NaCl, 1 mM EDTA, 0.1% Triton X-100) and resuspended in the same buffer at a concentration of 7×10<sup>8</sup> beads/ml. 10-100 pmoles of biotinylated oligonucleotide TGL382 was incubated with 100 µl of the Dynabead suspension in TNET for 30 minutes at 20° C. in order to allow the biotin moiety to bind to the streptavidin. The beads were then washed (using a magnet to immobilize them) three times with 200 µl of TNET and resuspended in 100 µl of TNET. For annealing, 25 µl of this suspension of the Dynabeads with the attached template oligonucleotide was immobilized with the magnet, the TNET withdrawn, and 25 µl of 40 mM Tris-HCl, pH 7.5, 20 mM MgCl<sub>2</sub>, 50 mM NaCl, containing 2 µM of oligonucleotide primers 346 or 391, was added. The template and each primer were annealed by incubating them for 5 minutes at 65° C., followed by slow cooling over a period of 20 minutes to room temperature. Beads containing the bound template-primer complexes were washed twice with 200 µl TNET, followed by resuspension in 25 µl of 40 mM Tris-HCl, pH 7.5, 20 mM MgCl<sub>2</sub>, 50 mM NaCl.

The following ddNTP mixes were used:

<sup>35</sup>S-labelled dideoxynucleoside triphosphate mixes (labelled nucleotide indicated in the form ddN\*TP):

|    |          |                            |                            |             |
|----|----------|----------------------------|----------------------------|-------------|
| 40 | ddG Mix: | 5 µM ddG*TP<br>10 µM ddCTP | 10 µM ddATP                | 10 µM ddTTP |
|    | ddA Mix: | 10 µM ddGTP<br>10 µM ddCTP | 5 µM ddA*TP<br>10 µM ddITP |             |
|    | ddT Mix: | 10 µM ddGTP<br>10 µM ddCTP | 10 µM ddATP                | 5 µM ddT*TP |
| 45 | ddC Mix: | 10 µM ddGTP<br>5 µM ddC*TP | 10 µM ddATP                | 10 µM ddTTP |

The ddN\*TPs were the four respective [α-thio-<sup>35</sup>S] dideoxynucleoside triphosphates (purchased from New England Nuclear).

For each bead-bound, template-primer complex, four extension reactions were carried out, one reaction for each of the four ddNTP mixes. Extension reactions contained the following components: 5.0 µl bead suspension containing the annealed template-primer complex, 0.5 µl of 100 mM dithiothreitol, 0.5 µl of "Mn<sup>++</sup> solution" (100 mM MnCl<sub>2</sub>, 150 mM DL-isocitrate, pH 7.0; purchased from U.S. Biochemicals, Cleveland, Ohio), 1.0 µl of ddG, ddA, ddT, or ddC mix, 2.0 µl of H<sub>2</sub>O, and 1.0 µl of T7 DNA polymerase ("Sequenase", version 2.0, US Biochemicals, 1625 units/ml in 50 mM Tris-HCl, pH 7.5, 10 mM 2-mercaptoethanol, 1 mg/ml bovine serum albumin).

Reactions were allowed to proceed for 15 minutes at 20° C., then stopped by washing the magnetically immobilized beads three times with 500 µl TNET. Beads were resuspended in final volume of 25 µl TNET prior to the detection assays.

EXHIBIT B-Page 23

EXHIBIT B-Page 24

## Solid phase immobilization of nucleic acids

GBA reactions were performed in 96-well plates (Nunc Nunc隆 plates, Roskilde, Denmark). The GBA primer was covalently coupled to the plate by incubating 10 pmoles of primer having a 5' amino group per well in 50  $\mu$ l of 3 mM sodium phosphate buffer, pH 6, 20 mM 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide (EDC) overnight at room temperature. After coupling, the plate was washed three times with 10 mM Tris pH 7.5/150 mM NaCl/0.05% Tween-20 (TNTw).

## Biotinylated ddNTPs

Biotinylated ddNTPs were synthesized according to U.S. Pat. No. 5,047,519.

## GBA in Microwell Plates

Hybridization of single-stranded DNA to primers covalently coupled to 96-well plates was accomplished by adding an equal volume of 3M NaCl/50 mM EDTA to the second round asymmetric PCR and incubating each well with 20  $\mu$ l of this mixture at 55°C. for 30 minutes. The plate was subsequently washed three times with TNTw. Twenty (20)  $\mu$ l of polymerase extension mix containing ddNTPs (3  $\mu$ M each, one of which was biotinylated/5 mM D'IT/7.5 mM sodium isocitrate/5 mM MnCl<sub>2</sub>/0.04 units per  $\mu$ l of modified T7 DNA polymerase and incubated for 5 minutes at room temperature. Following the extension reaction, the plate was washed once with TNTw. Template strands were removed by incubating wells with 50  $\mu$ l 0.2N NaOH for 5 minutes at room temperature, then washing the wells with another 50  $\mu$ l 0.2N NaOH. The plate was then washed three times with TNTw. Incorporation of biotinylated ddNTPs was measured by an enzyme-linked assay. Each well was incubated with 20  $\mu$ l of streptavidin-conjugated horseradish peroxidase (1/100 dilution in TNTw of product purchased from BRL, Gaithersburg, Md.) with agitation for 30 minutes at room temperature. After washing 5 times with TNTw, 100  $\mu$ l of o-phenylenediamine (OPD, 1 mg/ml in 0.1 M Citric acid, pH 4.5) (BRL) containing 0.012% H<sub>2</sub>O<sub>2</sub> was added to each well. The amount of bound enzyme was determined by photographing the plate after stopping the reaction or quantitatively using a Molecular Devices model "Vmax" 96-well spectrophotometer.

In order to demonstrate the generality of the procedure, the ability to type three different sites located on two different template molecules is shown. In the middle of FIGS. 9 through 11 is shown the polymorphic region of these loci together with the sequence of the GBA primers used to genotype the DNA samples. The genotype of the test DNA samples was previously determined by restriction analysis and gel electrophoresis (equine samples) or by allele specific hybridization (human samples).

At the top and bottom of FIGS. 9 through 11 are photographs of the non-radioactive GBA analysis of these sites. Analysis of the "plus" strand (which corresponds to the mRNA for the HLA DPA1 but is arbitrarily chosen for the equine locus JH85) is shown at the top of the figure, analysis of the "minus" strand is shown in the lower photograph. Using horseradish peroxidase activity genotyping data was observed visually. Because both strands were suitable templates for GBA, it was possible to get genotypic confirmation by using two different primers. For the HLA DPA1 locus, two sites of variation were typed (FIGS. 9 and 10). Identical results were achieved. Spectrophotometric quantitation of a separate experiment involving the equine locus JH85 is shown in FIG. 12. The average ratio of signals obtained with expected vs. inappropriate base incorporation was 62.2.

## C. EMBODIMENTS

An example of one method to practice the present invention involves obtaining from a convenient source, such as blood, epithelium, hair, or other tissue, samples of DNA or RNA, then amplifying in vitro specific regions of the nucleic acid using the polymerase chain reaction, transcription-based amplification (see Kwok, et al., Proc. Natl. Acad. Sci. 80:1173 (1989)), etc. Amplification is accomplished using specific primers flanking the region of interest, with one or more of the primers being modified by having an attached affinity group (although in any given reaction only one such primer is modified at a time). A preferred modification is attachment of biotin moieties to the 5'-termini of the primers. A sample (typically, 0.5–5 pmoles) of the amplified DNA is then bound to streptavidin-conjugated magnetic microspheres (e.g., Dynal M-280 "Dynabeads") via the attached biotin moiety on the amplification primer. The DNA is denatured by adjusting the aqueous suspension containing the microspheres to a sufficiently alkaline pH, and the strand bound to the microspheres via the biotin-streptavidin link is separated from the complementary strand by washing under similar alkaline conditions. To accomplish this, the microspheres are centrifuged or immobilized by the application of a magnetic field. The microsphere-bound strand is then used as a template in the remaining manipulations.

To the template strand, generated as described above, a specific primer oligonucleotide is bound under high stringency annealing conditions, the sequence of the primer being consistent with unique binding to a site on the template strand immediately adjacent to a known DNA sequence polymorphism. A preferred sequence and mode of binding for the primer ensures that the primer forms a duplex with the template such that the 3'-terminal nucleotide of the primer forms a Watson-Crick basepair with the template nucleotide immediately adjacent to the site of the first nucleotide in the sequence polymorphism, without the duplex overlapping any of the polymorphic sequence to be analyzed. This arrangement causes the nucleotides added via template-directed, DNA polymerase-catalyzed, extension of the primer to be determined unambiguously by the polymorphic nucleotide sequence in the template.

The above-described primer:template complex is contacted, under conditions of salt, pH, and temperature compatible with template-directed DNA synthesis, with a suitable DNA polymerase and four different chain-terminating nucleotide analogues known to form specific base pairs with the bases in the template. Most likely, but not necessarily, the bases in the template as well as the chain-terminating analogues are based on the common nucleosides: adenosine, cytosine, guanine or inosine, thymidine or uridine. A preferred set of chain-terminating analogues are the four dideoxynucleoside triphosphates, ddATP, ddCTP, ddGTP, and ddTTP, where each of the four ddNTPs has been modified by attachment of a different fluorescent reporter group. These fluorescent tags would have the property of having spectroscopically distinguishable emission spectra, and in no case would the dideoxynucleoside triphosphate modification render the chain-terminating analogue unsuitable for DNA polymerase-catalyzed incorporation onto primer 3'-termini. The result of DNA polymerase-catalyzed chain extension in such a mixture with such a primer:template complex is the quantitative, specific and unambiguous incorporation of a fluorescent chain-terminating analogue onto the 3'-terminus of the primer, the particular fluorescent nucleotide added being solely dictated by the sequence of the polymorphic nucleotides in the template.

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-continued

(J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

AGGATGCTTT TGTGGCAAAA CAC

23

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

AGATGATGCT TTTGTGCAAA ACAC

24

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

TCAATACCTG AGTCCCGACA CCCTG

25

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

AGCCTCAGAC CGCGTGGTGC CTGGT

25

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992

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(J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CGTCTGCAGA ATCCACTGGC TTCTTGAG

28

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

GCAGGATCCT GGAAC TACTC ATTTGCCT

28

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TCAATACCTG AGTCCCGACA CCCTG

25

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

AGGATGATGC TTTTGTGCAA AACAC

25

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992

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(J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GCGGACCATG TGTCAACTTA T

21

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GCCTGAGTGT GGTTGGAACT G

21

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TTCTTCTTGC ATCTATGTTC G

21

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

TTAACGCACCA CCACAGGTCC T

21

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992

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(J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GCCTTGGCGT TGTAGAA

17

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

AGAGAACAA TTTCAAG

17

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 240 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 114
- (D) OTHER INFORMATION: /label= n  
/note= "at residue 114, n = either c or t"

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 190
- (D) OTHER INFORMATION: /label= n  
/note= "at residue 190, n = either a or g"

(x) PUBLICATION INFORMATION:

- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

TTCTTCTTG CATCTATGTT CGTTTTCTT ATTGCTACAA ATGCCTATGC ACGGCCTGAC

60

TTCTGCCTAG AGCCTCCATA TACGGGTCCC TGCAAGGCCA GAATTATCG ATANTTCTAC

120

AACGCCAAGG CTGGGCTCTG CCAGACCTTT GTATAATGGTG GCTGCAGAGC TAAGAGAAC

180

AATTTCAAGN GCGCAGAGGA CTGCATGAGG ACCTGTGGTG GTGCTTAAGG GCCCCGGAA

240

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 327 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

- (x) PUBLICATION INFORMATION:
- (H) DOCUMENT NUMBER: WO PCT/US92/01905
- (I) FILING DATE: 04-MAR-1992
- (J) PUBLICATION DATE: 17-SEP-1992

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## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

|   |     |
|---|-----|
| AGATGATGCT TTTGTGCAAA ACACTTTTA ACACCTCTT TAAAATTCTT TTCAAATTCT   | 60  |
| ACGGCATTTT TTTCCTGAAA ATGCTTCGGT TTTAGGTCAA AGCTTTATTC TCCTAAGAAC | 120 |
| CTAACTCCCA CTGGTCTCAG GCGCCCTCTG GGAGCCCTCG GGGAGTCTTT GCCCCCCAAT | 180 |
| CTTGGCATTC TCCCCTGACA CTCGCCAAAG GCCCCTAACC TGCACCCGGG CACCAGGCAC | 240 |
| CACGCGGTCT GAGGCTTCAG CAGGGAAAGGC CTGCTCTCCG TTCACACTGC TTTCAGGCC | 300 |
| GGCAGGGTGT CGGGACTCAG GTATTGA                                     | 327 |

## (2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

|                             |    |
|-----------------------------|----|
| AGCCTCAGAC CGCGTGGTGC CTGGT | 25 |
|-----------------------------|----|

## (2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

|                             |    |
|-----------------------------|----|
| TCAATACCTG AGTCCCGACA CCCTG | 25 |
|-----------------------------|----|

## (2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 57 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 29..30
  - (D) OTHER INFORMATION: /label= n  
/note= "at residue 29, n = either a or c; at residue 30, n = either t or a"

- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

|  |    |
|--|----|
| GTTTATGTTT GAATTTGATG AAGATGAGNN GTTCTATGTG GATCTGGACA AGAAGGA | 57 |
|--|----|

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## (2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 57 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 28..29
  - (D) OTHER INFORMATION: /label= n  
/note= "at residue 28, n = either a or t; at residue 29, n = either t or g"
- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

TCCTTCTTGT CCAGATCCAC ATAGAACNNC TCATCTTCAT CAAATTCAAA CATGAAC

57

## (2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 57 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 29
  - (D) OTHER INFORMATION: /label= n  
/note= "at residue 29, n = either a or g"
- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

ACCGTCTGGC ATCTGGAGGA GTTTGGCCNA GCCTTTCTT TTGAGGCTCA GGGGGGG

57

## (2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 57 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 29
  - (D) OTHER INFORMATION: /label= n  
/note= "at residue 29, n = either t or c"
- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

CCCGCCCTGA GCCTCAAAGG AAAAGGCTNG GCCAAACTCC TCCAGATGCC AGACGGT

57

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-continued

## (2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 57 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 29..30
  - (D) OTHER INFORMATION: /label= n  
/note= "at residue 29, n = either g or t; at residue 30, n = either no base or c"
- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

TGAAGCCTCA GACCGCGTGG TGCCTGGTNN CCCGGGTCCA GGTTAGGGGC CTTGGGC

57

## (2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 57 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 28..29
  - (D) OTHER INFORMATION: /label= n  
/note= "at residue 28, n = either no base or g; at residue 29, n = either c or a"
- (x) PUBLICATION INFORMATION:
  - (H) DOCUMENT NUMBER: WO PCT/US92/01905
  - (I) FILING DATE: 04-MAR-1992
  - (J) PUBLICATION DATE: 17-SEP-1992
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

GCCCAAGGCC CCTAACCTGC ACCCGGGNNAA CCAGGCACCA CGCGGTCTGA GGCTTCA

57

What is claimed is:

1. A reagent composition which comprises an aqueous carrier, a primer nucleic acid molecule, a template nucleic acid of interest hybridized thereto, and an admixture of at least two different terminators of a nucleic acid template-dependent, primer extension reaction, each of the terminators being capable of specifically terminating an extension reaction of said primer hybridized to said template nucleic acid molecule of interest in a manner strictly dependent on the identity of the unpaired nucleotide base in said template nucleic acid molecule of interest immediately adjacent to, and downstream of, the 3' end of said primer, and at least one of the terminators being labeled with a detectable marker, and wherein said reagent composition lacks dATP, dCTP, dGTP, and dTTP.

2. A reagent of claim 1, wherein the reagent comprises four different terminators.

3. A reagent of claim 2, wherein two of the terminators are labeled, each with a different detectable marker.

4. A reagent of claim 2, wherein three of the terminators are labeled, each with a different detectable marker.

5. A reagent of claim 2, wherein the four terminators are labeled, each with a different detectable marker.

6. A reagent of any of claims 1-5, wherein the terminator(s) comprise(s) a nucleotide or nucleotide analog.

7. A reagent of claim 6, wherein the terminator(s) comprise(s) dideoxynucleotides.

8. A reagent of claim 6, wherein the terminator(s) comprise(s) arabinoside triphosphates.

9. A reagent of claim 7, wherein the terminator(s) comprise(s) one or more of ddATP, ddCTP, ddGTP or ddTTP.

10. A reagent of any of claims 1-5, wherein each of the different detectable markers is an isotopically labeled moiety, a chromophore, a fluorophore, a protein moiety, or a moiety to which an isotopically labeled moiety, a chromophore, a fluorophore, or a protein moiety can be attached.

11. A reagent of claim 10, wherein each of the different detectable markers is a different fluorophore.

12. A reagent of any of claims 1-5, wherein the reagent further comprises pyrophosphatase.

13. A method of determining the identity of a nucleotide base at a specific position in a nucleic acid of interest which comprises:

- (a) treating a sample containing the nucleic acid of interest, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position, or directly employing step (b) if the nucleic acid of interest is single-stranded; 5
- (b) contacting the sample from step (a), with an oligonucleotide primer, said primer being immobilized to a solid support wherein said primer is fully complementary to and hybridizes specifically to a stretch of nucleotide bases present in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, under high stringency hybridization conditions, so as to form a duplex between the immobilized primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in said duplex; 10
- (c) contacting the duplex from step (b) in the absence of dATP, dCTP, dGTP, or dTTP, with at least two different terminators of a nucleic acid template-dependent, primer extension reaction capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the unpaired nucleotide base in the template nucleic acid of interest immediately downstream of the 3' end of the immobilized primer; wherein one of said terminators is complementary to said nucleotide base to be identified and wherein at least one of said terminators is labeled with a detectable marker, wherein if more than one terminator is labeled, a different label is used to label each such labeled terminator; wherein said contacting is under conditions sufficient to permit base pairing of said complementary terminator with the nucleotide base to be identified and occurrence of a template-dependent primer extension reaction sufficient to incorporate said complementary terminator onto the 3' end of the primer to thereby extend said 3' end of said primer by one terminator; 15
- (d) determining the presence and identity of the nucleotide base at the specific position in the nucleic acid of interest by detecting the detectable marker of said incorporated terminator while said terminator is incorporated at the 3' end of the extended primer, and wherein said detection is conducted in the absence of non-terminator nucleotides. 20

14. A method of determining the presence or absence of a desired polynucleotide having a particular nucleotide sequence in a sample of nucleic acids which comprises:

- (a) treating a sample of nucleic acids, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids, or directly employing step (b) if the sample of nucleic acids contains only single-stranded nucleic acids; 45
- (b) contacting the sample from step (a), with an oligonucleotide primer, said primer being immobilized to a solid support; wherein said primer is fully complementary to and hybridizes specifically to a stretch of nucleotide bases of the particular nucleotide sequence, if the desired polynucleotide having said particular nucleotide sequence is present, said contacting being under high stringency hybridization conditions, so as to form a duplex between the immobilized primer and the particular nucleotide sequence; 50

(c) contacting the duplex, if any, from step (b) in the absence of dATP, dCTP, dGTP or dTTP, with at least two different terminators of a nucleic acid template-dependent, primer extension reaction capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the unpaired nucleotide base in the template immediately downstream of the 3' end of the immobilized primer; wherein one of said terminators is complementary to said unpaired nucleotide base and wherein at least one of said terminators is labeled with a detectable marker, wherein if more than one terminator is labeled, a different label is used to label each such labeled terminator; wherein said contacting is under conditions sufficient to permit base pairing of said complementary terminator with said unpaired nucleotide base immediately downstream of the 3' end of said immobilized hybridized primer and occurrence of a template-dependent, primer extension reaction sufficient to incorporate said complementary terminator onto the 3' end of the immobilized primer to thereby extend said 3' end of said primer by one terminator; and 5

(d) determining the presence of the target nucleic acid molecule by detecting the detectable marker of said incorporated terminator while said terminator is incorporated at the 3' end of the extended primer from step (c) and wherein said detection is conducted in the absence of non-terminator nucleotides. 10

15. A method of determining the identity of a nucleotide base at a specific position in a nucleic acid of interest which comprises:

- (a) (1) incubating a sample containing the nucleic acid of interest with at least two oligonucleotide primers, and a polymerase, said primers being sufficient to mediate a polymerase chain reaction amplification of said nucleic acid of interest, wherein said incubation is conducted under conditions sufficient to permit said amplification to occur; 25
- (2) treating a sample containing said amplified nucleic acid of interest, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position, or directly employing step (b) if the nucleic acid of interest is single-stranded; 30
- (b) contacting the sample from step (a2), under hybridizing conditions, with an oligonucleotide primer, said primer being immobilized to a solid support wherein said primer is fully complementary to and hybridizes specifically to a stretch of nucleotide bases present in the nucleic acid of interest immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the immobilized primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in said duplex; 35
- (c) contacting the duplex from step (b), in the absence of dATP, dCTP, dGTP, or dTTP, with at least two different terminators of a nucleic acid template-dependent, primer extension reaction capable of specifically terminating the extension reaction in a manner strictly dependent upon the identity of the unpaired nucleotide base in the template nucleic acid of interest immediately downstream of the 3' end of the immobilized primer; wherein one of said terminators is complementary to said nucleotide base to be identified and wherein at least one of said terminators is labeled with a detectable marker, wherein if more than one terminator 40





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(12) **United States Patent**  
**Goelet et al.**

(10) **Patent No.:** **US 6,537,748 B1**  
**(45) Date of Patent:** **\*Mar. 25, 2003**

(54) **REAGENT FOR NUCLEIC ACID TYPING BY PRIMER EXTENSION**

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(73) Assignee: **Orchid BioSciences, Inc.**, Princeton, NJ (US)

(\*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

This patent is subject to a terminal disclaimer.

(21) Appl. No.: **09/258,133**

(22) Filed: **Feb. 26, 1999**

**Related U.S. Application Data**

(63) Continuation of application No. 07/775,786, filed on Oct. 11, 1991, now Pat. No. 6,004,744, and a continuation-in-part of application No. 07/664,837, filed on Mar. 5, 1991, now Pat. No. 5,888,819.

(51) **Int. Cl.**<sup>7</sup> ..... **C12Q 1/68; C12P 19/34; C07H 21/02; C07H 21/04**

(52) **U.S. Cl.** ..... **435/6; 435/5; 435/91.2; 536/23.1; 536/24.33; 536/26.1; 536/26.12**

(58) **Field of Search** ..... **435/6, 91.2, 5; 536/23.1, 26.1, 24.33, 26.12**

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**Primary Examiner—Carla J. Myers**

(74) **Attorney, Agent, or Firm—Kalow & Springut LLP**

(57) **ABSTRACT**

This invention concerns a reagent composition comprising at least two different terminators of a nucleic acid template-dependent, primer extension reaction. This invention also concerns a method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. This invention further concerns a method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids. This invention further concerns a method for identifying different alleles in a sample containing nucleic acids. This invention further concerns a method for determining the genotype of an organism at one or more particular genetic loci.

**11 Claims, 12 Drawing Sheets**

**EXHIBIT C-Page 1**

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**FIGURE 1**

**A**            **B**            **C**  
180 181    180 181    180 181



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**FIGURE 2.****I. Amplification primers**

TGL 105: 5'-TTCTTCTTGCATCTATGTTCTG-3'

TGL 106: 5'-TTAAGCACCACACAGGTCCCT-3'

**II. Polymorphism detection primers**

TGL 182: 5'-GCCTTGGCGTTGTAGAA-3'

TGL 166: 5'-AGAGAAACAATTCAAG-3'

**III. Target sequence**5' ... TTTCTTCTTG CATCTATGTT CGTTTTTCT ATTGCTACAA 40  
TGL 105 ----->

ATGCCTATGC ACGGCCTGAC TTCTGCCTAG AGCCTCCATA 80

TACGGTCCC TGCAAGGCCA GAATTATCAG ATA<sup>C</sup>/TTTCTAC 120AACGCCAAGG CTGGGCTCTG CCAGACCTTT GTATATGGTG 160  
<----- TGL 182GCTGCAGAGC TAAGAGAAAC AATTCAAG<sup>A</sup>/G GCGCAGAGGA 200  
TGL 166 ----->CTGCATGAGG ACCTGTGGTG GTGCTTAAGG GCCCCGGGAA..3' 240  
<----- TGL 106**IV. Polymorphisms**

| <u>Plasmid</u> | <u>Nucleotide 114</u> | <u>Nucleotide 190</u> |
|----------------|-----------------------|-----------------------|
| p183           | C                     | A                     |
| p624           | T                     | A                     |
| p814           | C                     | G                     |

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**FIGURE 3**

**A B C D**



**A: TGL 182/**

**p183**

**B: TGL 182/**

**p624**

**C: TGL 166/**

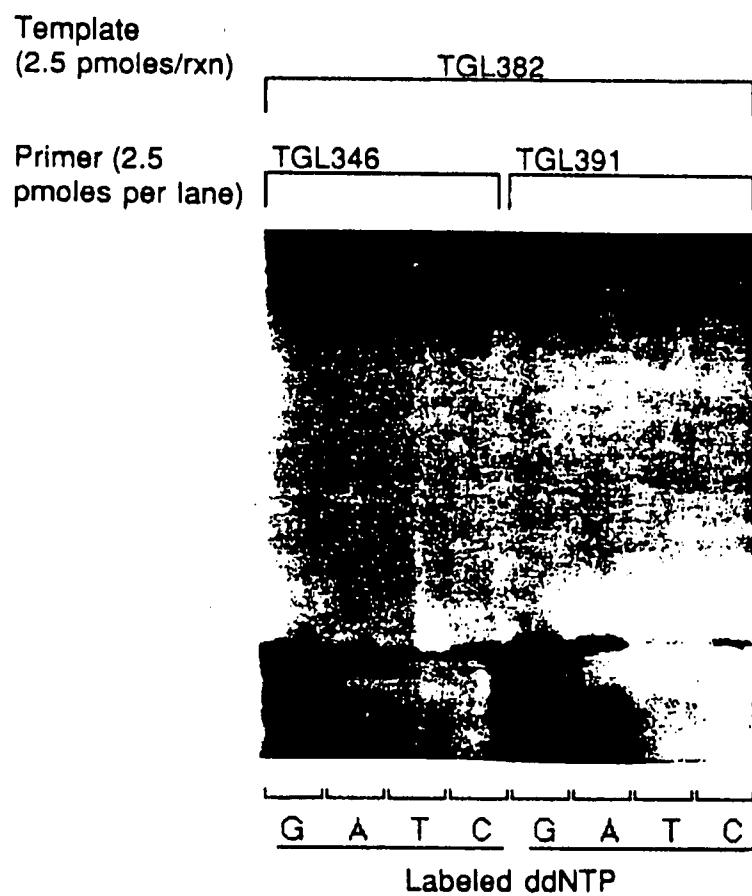
**p183**

**D: TGL 166/**

**p814**

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**FIGURE 4**



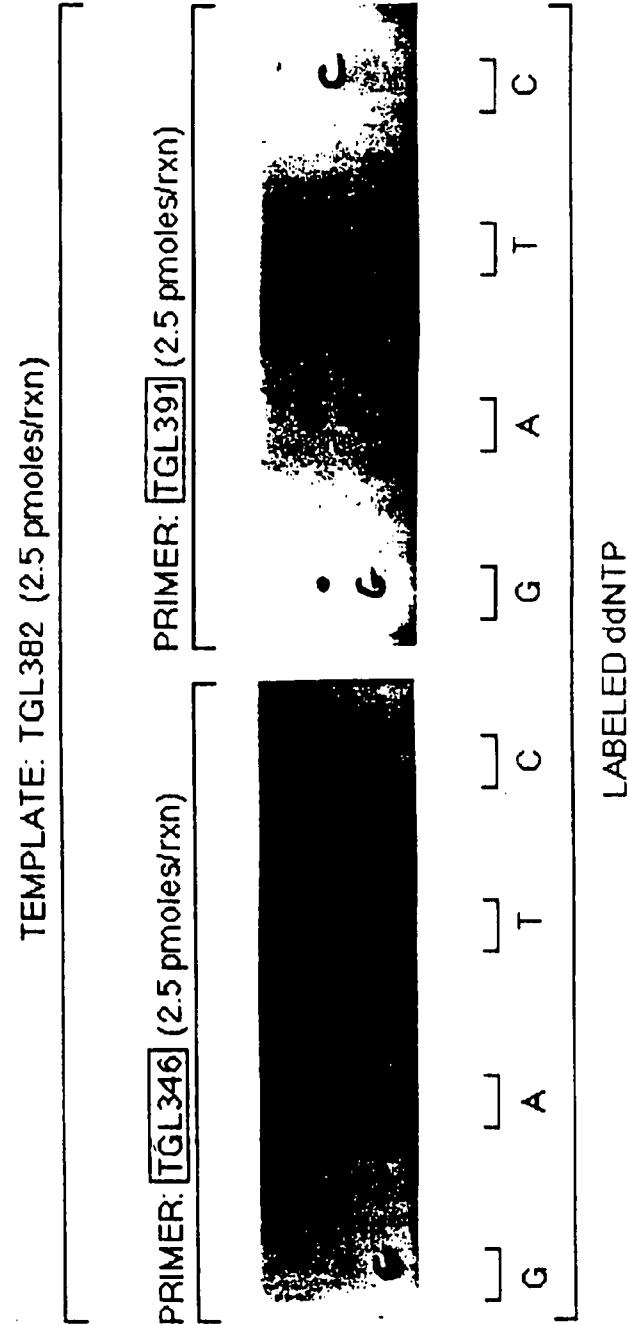
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FIGURE 5



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FIGURE 6

Attached to Bead

5' AGATGATGCT TTTGTGCAA ACACTTTTA ACACCTCTT TAAAATTCT TTCAAATTCT ACGGCATTT  
TGL240 (PCR Primer with Biotin)

TTTCCTGAAA ATGCTTCGGT TTTAGGTCAA AGCTTTATTC TCCTAAGAAC CTAACTCCCA CTGGTCTCAG

GCGCCCTCTC GGAGCCCTCG GGGAGTCCTT GCCCCCCAAT CTTGGCATTC TCCCCTGACA CTGGCCCAAG

CCCCCTAACCG TGCACCCGGG 3' TGGTCCGT GGTGGGCCAG ACTCCGA 5'  
GA CACCAAGGCA CCACGGCGTC TGAGGCTTCA GCAGGGAAAGG CCTGCTCTCC  
T C B allele  
differences  
TGL239 (Non-biotinylated PCR Primer)  
3' GTCACACTG CTTTCAGGCC CGGCAGGGTG TCGGGACTCA GGTATTGA 5'

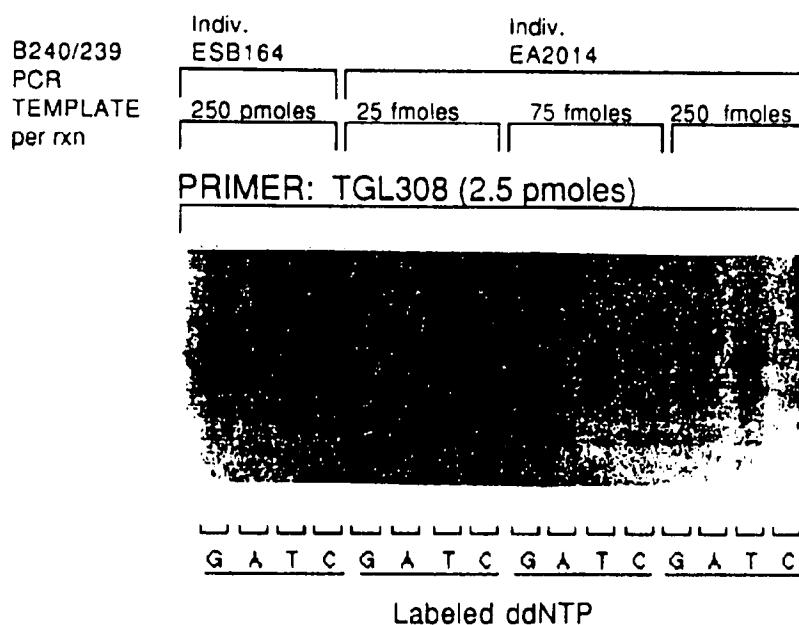
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**FIGURE 7**



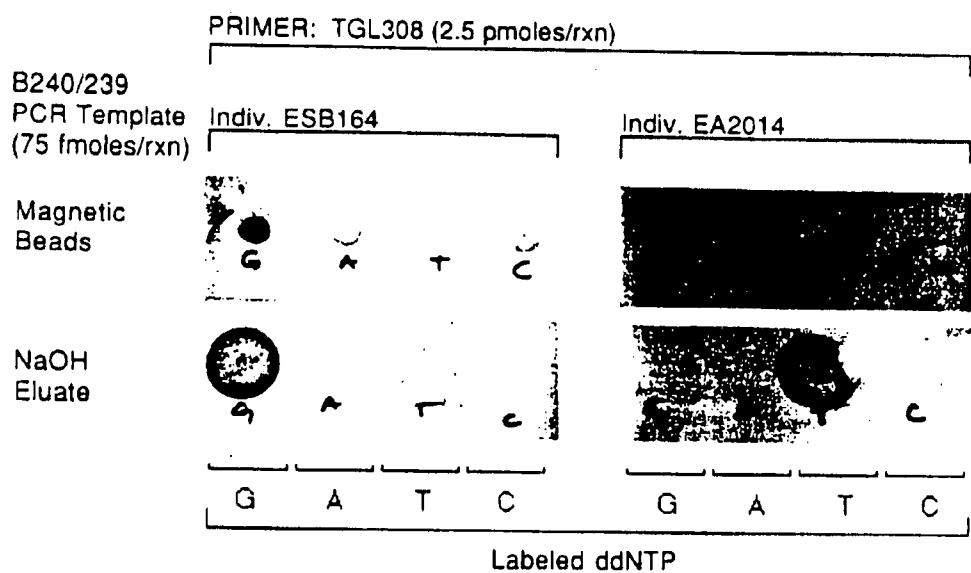
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FIGURE 8



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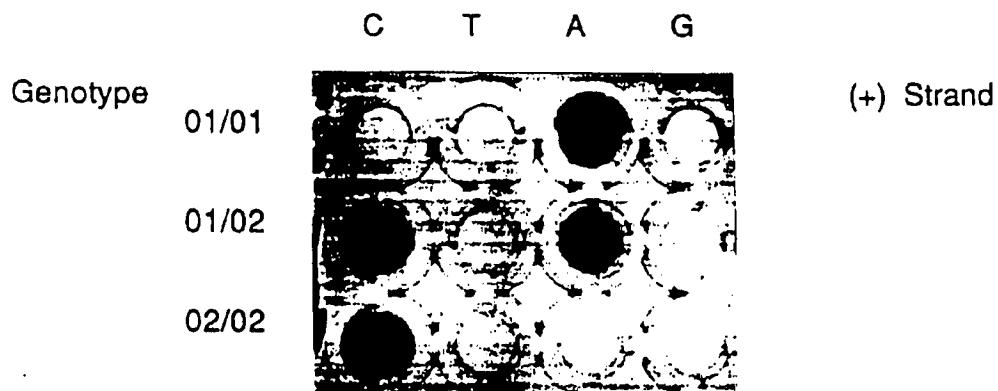
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FIGURE 9

## GBA: HLA DPA1 aa 31



#517 → AT      CA      GTTCTATGTGGATCTGGACAAGAAGGA

CAAGTACAAACTAAACTACTTCTACTC      TA      CAAGATACACCTAGACCTGTTCTCCT

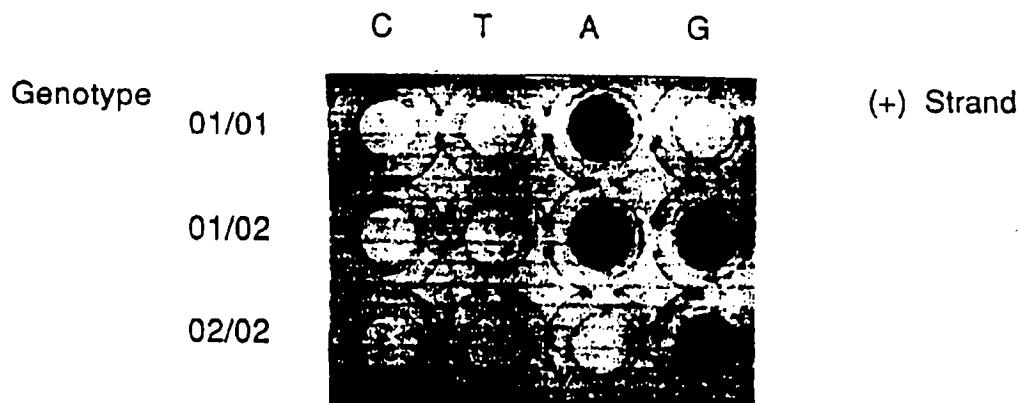
GT ← #518



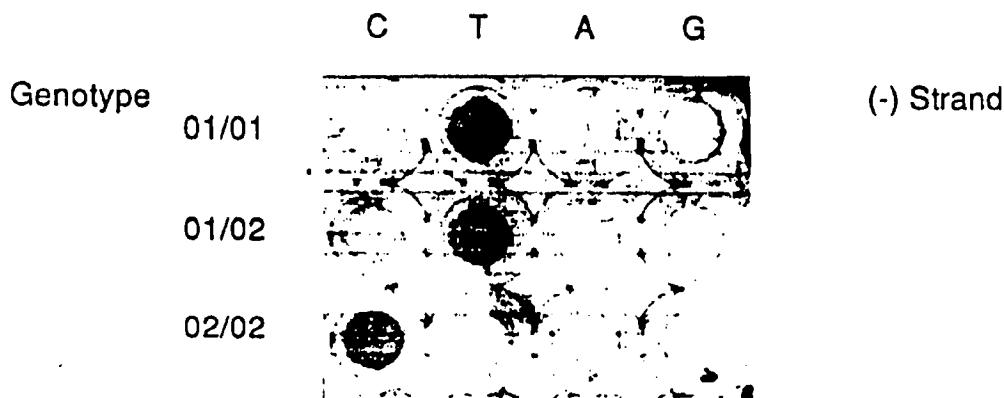
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FIGURE 10

## GBA: HLA DPA1 aa50



#515 →  
 ACCGTCTGGCATCTGGAGGAGTTGGCC A: AGCCTTTCTTGAGGCTCAGGGCGGG  
 TGGCAGACCGTAGACCTCCTCAAACCGG T: TCGGAAAAGGAAACTCCGAGTCCCGCCC  
 C ← #513



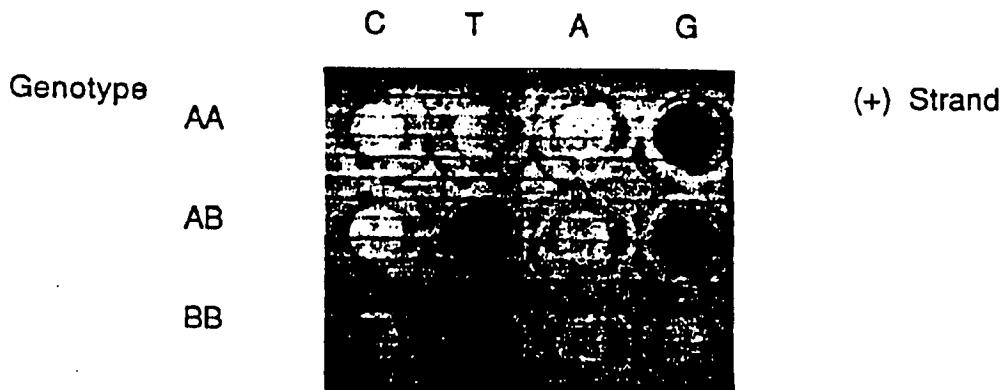
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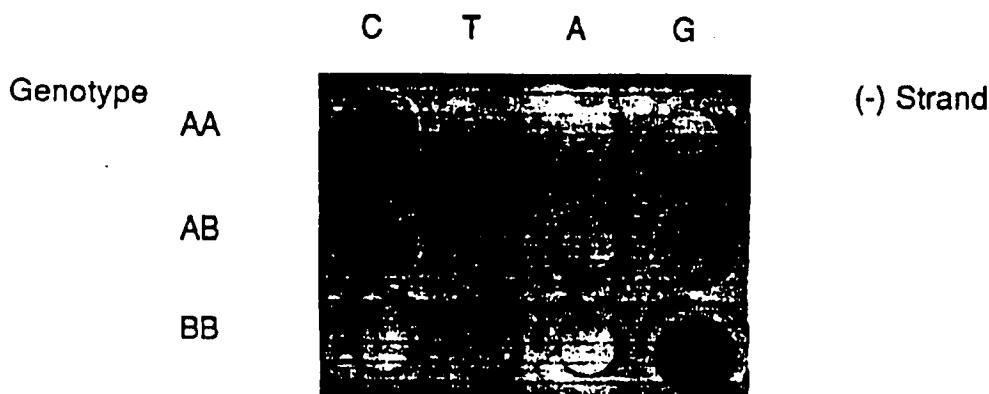
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FIGURE 11

GBA: Equine Locus JH85  
Nucleotide 122

#308 →  
 TGAAGCCTCAGACCGCGTGGTGCCTGGT G- CCCGGGTGCAGGTTAGGGGCCTGGGC  
 TC  
 ACTTCGGAGTCTGGCGCACACGGACCA C- GGGCCCACGTCCAATCCCCGGAACCCG  
 AG ← #307

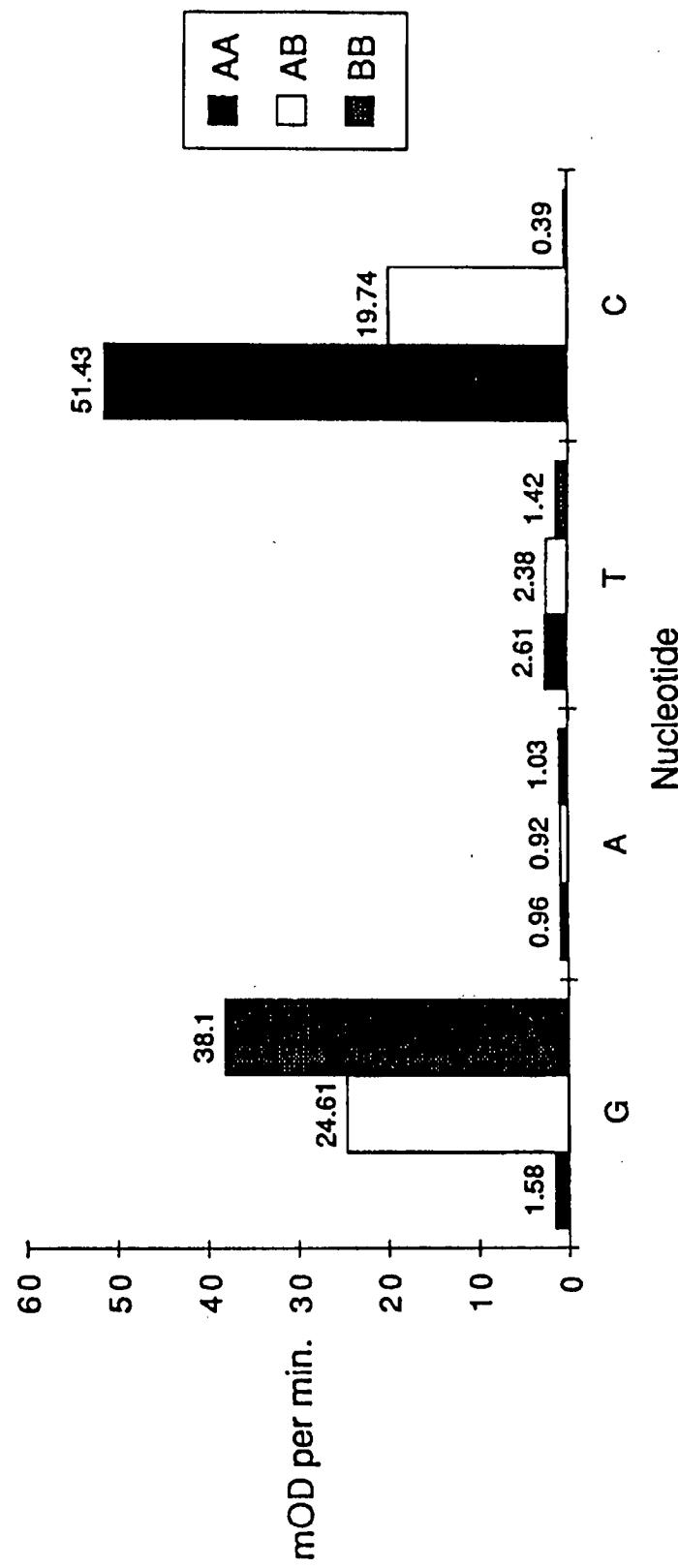


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FIGURE 12

QUANTITATIVE GBA: Equine locus JH85



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## REAGENT FOR NUCLEIC ACID TYPING BY PRIMER EXTENSION

This application is a continuation of application Ser. No. 07/775,786, filed Oct. 11, 1991 U.S. Pat. No. 6,004,744 and a continuation in part of Ser. No. 07/664,837, filed Mar. 5, 1991 which is now U.S. Pat. No. 5,888,819. Both application Ser. No. 07/775,786 and U.S. Pat. No. 5,888,819 are incorporated herein by reference.

## BACKGROUND OF THE INVENTION

This invention relates to the field of nucleic acid sequence detection. The detection of nucleic acid sequences can be used in two general contexts. First, the detection of nucleic acid sequences can be used to determine the presence or absence of a particular genetic element. Second, the detection of nucleic acid sequences can be used to determine the specific type of a particular genetic element that is present. Variant genetic elements usually exist. Many techniques have been developed (1) to determine the presence of specific nucleic acid sequences, and (2) to compare homologous segments of nucleic acid sequence to determine if the segments are identical or if they differ at one or more nucleotides. Practical applications of these techniques include genetic disease diagnoses, infectious disease diagnoses, forensic techniques, paternity determinations, and genome mapping.

In general, the detection of nucleic acids in a sample and the subtypes thereof depends on the technique of specific nucleic acid hybridization in which the oligonucleotide probe is annealed under conditions of high stringency to nucleic acids in the sample, and the successfully annealed probes are subsequently detected (see Spiegelman, S., *Scientific American*, Vol. 210, p. 48 (1964)).

The most definitive method for comparing DNA segments is to determine the complete nucleotide sequence of each segment. Examples of how sequencing has been used to study mutations in human genes are included in the publications of Engelke, et al., *Proc. Natl. Acad. Sci. U.S.A.*, 85:544-548 (1988) and Wong, et al., *Nature*, 330:384-386 (1987). At the present time, it is not practical to use extensive sequencing to compare more than just a few DNA segments because the effort required to determine, interpret, and compare sequence information is time-consuming.

A commonly used screen for DNA polymorphisms arising from DNA sequence variation consists of digesting DNA with restriction endonucleases and analyzing the resulting fragments by means of Southern blots, as described by Botstein, et al., *Am. J. Hum. Genet.*, 32:314-331 (1980) and White, et al., *Sci. Am.*, 258:40-48 (1988). Mutations that affect the recognition sequence of the endonuclease will preclude enzymatic cleavage at that site, thereby altering the cleavage pattern of that DNA. DNAs are compared by looking for differences in restriction fragment lengths. A major problem with this method (known as restriction fragment length polymorphism mapping or RFLP mapping) is its inability to detect mutations that do not affect cleavage with a restriction endonuclease. Thus, many mutations are missed with this method. One study, by Jeffreys, *Cell*, 18:1-18 (1979), was able to detect only 0.7% of the mutational variants estimated to be present in a 40,000 base pair region of human DNA. Another problem is that the methods used to detect restriction fragment length polymorphisms are very labor intensive, in particular, the techniques involved with Southern blot analysis.

A technique for detecting specific mutations in any segment of DNA is described in Wallace, et al., *Nucl. Acids*

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*Res.*, 9:879-894 (1981). It involves hybridizing the DNA to be analyzed (target DNA) with a complementary, labeled oligonucleotide probe. Due to the thermal instability of DNA duplexes containing even a single base pair mismatch, differential melting temperature can be used to distinguish target DNAs that are perfectly complementary to the probe from target DNAs that differ by as little as a single nucleotide. In a related technique, described in Landegren, et al., *Science*, 41:1077-1080 (1988), oligonucleotide probes are constructed in pairs such that their junction corresponds to the site on the DNA being analyzed for mutation. These oligonucleotides are then hybridized to the DNA being analyzed. Base pair mismatch between either oligonucleotide and the target DNA at the junction location prevents the efficient joining of the two oligonucleotide probes by DNA ligase.

## A. Nucleic Acid Hybridization

The base pairing of nucleic acids in a hybridization reaction forms the basis of most nucleic acid analytical and diagnostic techniques. In practice, tests based only on parameters of nucleic acid hybridization function poorly in cases where the sequence complexity of the test sample is high. This is partly due to the small thermodynamic differences in hybrid stability, generated by single nucleotide changes, and the fact that increasing specificity by lengthening the probe has the effect of further diminishing this differential stability. Nucleic acid hybridization is, therefore, generally combined with some other selection or enrichment procedure for analytical and diagnostic purposes.

Combining hybridization with size fractionation of hybridized molecules as a selection technique has been one general diagnostic approach. Size selection can be carried out prior to hybridization. The best known prior size selection technique is Southern Blotting (see Southern, E., *Methods in Enzymology*, 69:152 (1980)). In this technique, a DNA sample is subjected to digestion with restriction enzymes which introduce double stranded breaks in the phosphodiester backbone at or near the site of a short sequence of nucleotides which is characteristic for each enzyme. The resulting heterogeneous mixture of DNA fragments is then separated by gel electrophoresis, denatured, and transferred to a solid phase where it is subjected to hybridization analysis *in situ* using a labeled nucleic acid probe. Fragments which contain sequences complementary to the labeled probe are revealed visually or densitometrically as bands of hybridized label. A variation of this method is Northern Blotting for RNA molecules. Size selection has also been used after hybridization in a number of techniques, in particular by hybrid protection techniques, by subjecting probe/nucleic acid hybrids to enzymatic digestion before size analysis.

## B. Polymerase Extension of Duplex Primer:template Complexes

Hybrids between primers and DNA targets can be analyzed by polymerase extension of the hybrids. A modification of this methodology is the polymerase chain reaction in which the purification is produced by sequential hybridization reactions of anti-parallel primers, followed by enzymatic amplification with DNA polymerase (see Saiki, et al., *Science* 239:487-491 (1988)). By selecting for two hybridization reactions, this methodology provides the specificity lacking in techniques that depend only upon a single hybridization reaction.

It has long been known that primer-dependent DNA polymerases have, in general, a low error rate for the addition of nucleotides complementary to a template. This feature is essential in biology for the prevention of genetic

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mistakes which would have detrimental effects on progeny. The specificity inherent in this enzymological reaction has been widely exploited as the basis of the "Sanger" or dideoxy chain termination sequencing methodology which is the ultimate nucleic acid typing experiment. One type of Sanger DNA sequencing method makes use of mixtures of the four deoxynucleoside triphosphates, which are normal DNA precursors, and one of the four possible dideoxynucleoside triphosphates, which have a hydrogen atom instead of a hydroxyl group attached to the 3' carbon atom of the ribose sugar component of the nucleotide. DNA chain elongation in the 5' to 3' direction ("downstream") requires this hydroxyl group. As such, when a dideoxynucleotide is incorporated into the growing DNA chain, no further elongation can occur. With one dideoxynucleotide in the mixture, DNA polymerases can, from a primer/template combination, produce a population of molecules of varying length, all of which terminate after the addition of one out of the four possible nucleotides. The series of four independent reactions, each with a different dideoxynucleotide, generates a nested set of fragments, all starting at the same 5' terminus of the priming DNA molecule and terminating at all possible 3' nucleotide positions.

Another utilization of dideoxynucleoside triphosphates and a polymerase in the analysis of DNA involves labeling the 3' end of a molecule. One prominent manifestation of this technique provides the means for sequencing a DNA molecule from its 3' end using the Maxam-Gilbert method. In this technique, a molecule with a protruding 3' end is treated with terminal transferase in the presence of radioactive dideoxy-ATP. One radioactive nucleotide is added, rendering the molecule suitable for sequencing. Both methods of DNA sequencing using labeled dideoxynucleotides require electrophoretic separation of reaction products in order to derive the typing information. Most methods require four separate gel tracks for each typing determination.

The following two patents describe other methods of typing nucleic acids which employ primer extension and labeled nucleotides. Mundy (U.S. Pat. No. 4,656,127) describes a method whereby a primer is constructed complementary to a region of a target nucleic acid of interest such that its 3' end is close to a nucleotide in which variation can occur. This hybrid is subject to primer extension in the presence of a DNA polymerase and four deoxynucleoside triphosphates, one of which is an  $\alpha$ -thionucleotide. The hybrid is then digested using an exonuclease enzyme which cannot use thio-derivatized DNA as a substrate for its nucleolytic action (for example Exonuclease III of *E. coli*). If the variant nucleotide in the template is complementary to one of the thionucleotides in the reaction mixture, the resulting extended primer molecule will be of a characteristic size and resistant to the exonuclease; hybrids without thio-derivatized DNA will be digested. After an appropriate enzyme digest to remove underivatized molecules, the thio-derivatized molecule can be detected by gel electrophoresis or other separation technology.

Vary and Diamond (U.S. Pat. No. 4,851,331) describes a method similar to that of Mundy wherein the last nucleotide of the primer corresponds to the variant nucleotide of interest. Since mismatching of the primer and the template at the 3' terminal nucleotide of the primer is counterproductive to elongation, significant differences in the amount of incorporation of a tracer nucleotide will result under normal primer extension conditions. This method depends on the use of a DNA polymerase, e.g., AMV reverse transcriptase, that does not have an associated 3' to 5' exonuclease activity.

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The methods of Mundy and of Vary and Diamond have drawbacks. The method of Mundy is useful but cumbersome due to the requirements of the second, different enzymological system where the non-derivatized hybrids are digested. The method of Vary is complicated by the fact that it does not generate discrete reaction products. Any "false" priming will generate significant noise in such a system which would be difficult to distinguish from a genuine signal.

10 The present invention circumvents the problems associated with the methods of Mundy and of Vary and Diamond for typing nucleic acid with respect to particular nucleotides. With methods employing primer extension and a DNA polymerase, the current invention will generate a discrete molecular species one base longer than the primer itself. In many methods, particularly those employing the polymerase chain reaction, the type of reaction used to purify the nucleic acid of interest in the first step can also be used in the subsequent detection step. Finally, with terminators which 15 are labeled with different detector moieties (for example different fluorophors having different spectral properties), it will be possible to use only one reagent for all sequence detection experiments. Furthermore, if techniques are used to separate the terminated primers post-reaction, sequence 20 detection experiments at more than one locus can be carried out in the same tube.

25 A recent article by Mullis (*Scientific American*, April 1990, pp. 56-65) suggests an experiment, which apparently was not performed, to determine the identity of a targeted 30 base pair in a piece of double-stranded DNA. Mullis suggests using four types of dideoxynucleosides triphosphate, with one type of dideoxynucleoside triphosphate being radioactively labeled.

35 The present invention permits analyses of nucleic acid sequences that can be useful in the diagnosis of infectious diseases, the diagnosis of genetic disorders, and in the identification of individuals and their parentage.

40 A number of methods have been developed for these purposes. Although powerful, such methodologies have been cumbersome and expensive, generally involving a combination of techniques such as gel electrophoresis, blotting, hybridization, and autoradiography or non-isotopic 45 revelation. Simpler technologies are needed to allow the more widespread use of nucleic acid analysis. In addition, tests based on nucleic acids are currently among the most expensive of laboratory procedures and for this reason 50 cannot be used on a routine basis. Finally, current techniques are not adapted to automated procedures which would be necessary to allow the analysis of large numbers of samples and would further reduce the cost.

55 The current invention provides a method that can be used to diagnose or characterize nucleic acids in biological samples without recourse to gel electrophoretic size separation of the nucleic acid species. This feature renders this 60 process easily adaptable to automation and thus will permit the analysis of large numbers of samples at relatively low cost. Because nucleic acids are the essential blueprint of life, each organism or individual can be uniquely characterized by identifiable sequences of nucleic acids. It is, therefore, possible to identify the presence of particular organisms or demonstrate the biological origin of certain samples by detecting these specific nucleic acid sequences.

#### SUMMARY OF THE INVENTION

65 The subject invention provides a reagent composition comprising an aqueous carrier and an admixture of at least two different terminators of a nucleic acid template-

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dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the unpaired nucleotide base in the template immediately adjacent to, and downstream of, the 3' end of the primer. In addition, at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and one, two, three, or four of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent as described above wherein the terminators comprise nucleotides, nucleotide analogs, dideoxynucleotides, or arabinoside triphosphates. The subject invention also provides a reagent wherein the terminators comprise one or more of dideoxyadenosine triphosphate (ddATP), dideoxycytosine triphosphate (ddCTP), dideoxyguanosine triphosphate (ddGTP), dideoxythymidine triphosphate (ddTTP), or dideoxyuridine triphosphate (ddUTP).

The subject invention also provides a method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with a stretch of nucleotide bases present in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest. Enzymatic extension of the oligonucleotide primer in the resultant duplex by one nucleotide, catalyzed, for example, by a DNA polymerase, thus depends on correct base pairing of the added nucleotide to the nucleotide base to be identified.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. The net result is that the oligonucleotide primer has been extended by one terminator. Next, the identity of the detectable marker present at the 3' end of the extended primer is determined. The identity of the detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

The subject invention also provides another method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. This additional method uses a reagent containing four terminators, only one of the terminators having a detectable marker.

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The subject invention also provides a method of typing a sample of nucleic acids which comprises identifying the base or bases present at each of one or more specific positions, each such nucleotide base being identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. Each specific position in the nucleic acid of interest is determined using a different primer. The identity of each nucleotide base or bases at each position can be determined individually or the identities of the nucleotide bases at different positions can be determined simultaneously.

The subject invention further provides a method for identifying different alleles in a sample containing nucleic acids which comprises identifying the base or bases present at each of one or more specific positions. The identity of each nucleotide base is determined by the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention also provides a method for determining the genotype of an organism at one or more particular genetic loci which comprises obtaining from the organism a sample containing genomic DNA and identifying the nucleotide base or bases present at each of one or more specific positions in nucleic acids of interest. The identity of each such base is determined by using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. The identities of the nucleotide bases determine the different alleles and, thereby, determine the genotype of the organism at one or more particular genetic loci.

#### BRIEF DESCRIPTION OF THE FIGURES

35 FIG. 1. Autoradiography of labeled DNA products after fractionation on a polyacrylamide urea gel. Panel A shows products of the "A" extension reaction on oligonucleotide primer 182 [SEQ ID NO: 1] directed by template oligonucleotides 180 or 181. [SEQ ID NO: 2]. Panel B shows products of the "B" termination reaction on oligonucleotide primer 182 annealed to template oligonucleotides 180 or 181. Panel C shows the same products as in panel B after purification on magnetic beads. Note: oligodeoxynucleotide 182 was used as supplied by Midland Certified Reagents with no further purification. The minor bands above and below the main band are presumably contaminants due to incomplete reactions or side reactions that occurred during the step-wise synthesis of the oligonucleotide. For a definition of the "A" extension reaction and the "B" termination reaction, see "A GENERAL METHODS" in the Detailed Description of the Invention.

45 FIG. 2. Detection of Sequence Polymorphisms in PCR Products. Target polymorphic DNA sequence showing amplification primers [SEQ ID NO 15] and [SEQ ID NO 16], detection primers [SEQ ID NO: 17] and [SEQ NO: 18] and molecular clone (plasmid) designations. For each primer, sites of binding to one or the other strand of the target DNA sequence [SEQ ID NO: 19] are indicated by underlining, and the direction of DNA synthesis is indicated by an arrow. Numbering for the target sequence is shown in the righthand margin. Polymorphic sites at positions 114 and 190 are indicated by bold lettering and a slash between the two polymorphic possibilities.

55 60 FIG. 3. Autoradiogram of gel-analyzed polymorphism test on PCR products. Templates from PCR products of p183, p624, or p814 were analyzed with the detection primers, TGL182 [SEQ ED NO: 17] and TGL166 [SEQ ID

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NO: 18] in a template-directed chain extension experiment, as described in the specification. Reaction products were fractionated by size on a polyacrylamide/urea DNA sequencing gel, and incorporation of [ $^{35}\text{S}$ ]- $\alpha$ -thio-dideoxy adenosine monophosphate was assayed by autoradiography.

FIG. 4. Gel electrophoretic analysis of the labeled extension products of primers TGL346 [SEQ ID NO: 4] and TGL391 [SEQ ID NO: 3]. Productive primer-template complexes of TGL346 or TGL391 with the bead-bound oligonucleotide template, TGL382 [SEQ ID NO: 5], were subjected to primer extension labeling reactions with the four different [ $\alpha$ -thio  $^{35}\text{S}$ ] dideoxynucleoside triphosphate mixes. Labelled primer DNA was released from the washed beads and electrophoresed on an 8% polyacrylamide/8 M urea DNA sequencing gel (2.5 moles of primer/lane), then analyzed by autoradiography. The four lanes shown for the primer TGL346 indicate that labeling occurred predominantly with the ddC mix, indicating that the next unpaired based in the TGL382 template adjacent to the 3' end of TGL346 was a G (see sequence given in Example 4). The four lanes shown for the primer TGL391 indicate that the labeling occurred predominantly with the ddT mix, indicating that the next unpaired base in the TGL382 template adjacent to the 3' end of TGL391 was an A.

FIG. 5. Autoradiographic analyses of total radioactivity bound to beads. The bead suspensions, containing the products of the extension reactions described in FIG. 5, were spotted onto filter paper (1 pmole of primer per spot) and exposed to X-ray film to assay total bead-bound radioactivity. As shown, TGL346 predominantly incorporated label from the ddC mix and TGL391 predominantly from the ddT mix.

FIG. 6. PCR-amplified polymorphic locus of mammalian DNA. Shown is a 327 basepair segment of mammalian DNA that was amplified from samples of genomic DNA using the PCR primers TGL240 [SEQ ID NO: 6] (biotinylated) and TGL239 (unbiotinylated). Samples of DNA from two homozygous individuals, ESB164 (genotype AA) and EA2014 (genotype BB), [SEQ ID NO: 21] were subjected to the analyses described in Example 5. The complete DNA sequence [SEQ ID NO: 20] of the A allele at this locus is shown, with the polymorphic sites where the B allele sequence differs from the A allele sequence indicated by the bases underneath the A sequence. The detection primer, TGL308 [SEQ ID NO: 8], is shown base-paired with the template strand extending from the biotinylated primer. For the A allele, the first unpaired template base immediately downstream of the 3' end of TGL308 is a C, and for the B allele this base is an A. Thus, the A allele should result in labeling of TGL308 by the ddG mix only, and the B allele should result in labeling by the ddT mix only.

FIG. 7. Gel electrophoretic analysis of PCR products from two different homozygous individuals. Primers TGL240 [SEQ ID NO: 6] and TGL239 [SEQ ID NO: 7] were used to amplify genomic DNA (obtained from blood) from two individuals, ESB164 and EA2014. The products of the extension reactions for primer TGL308 [SEQ ID NO: 8] annealed to the bead-bound, PCR-generated template as outlined in FIG. 7, were analyzed by electrophoresis on an 8% polyacrylamide/8 M urea DNA sequencing gel as outlined in FIG. 5. Shown for individual ESB164 (genotype AA: labeling expected from the ddG mix) are 250 fmoles of extended primer from the four different ddNTP labelling reactions. Shown for individual EA2014 (genotype BB: labeling expected from the ddT mix) are loadings of 25, 75, and 250 fmoles of extended primer from the four different ddNTP labeling reactions.

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FIG. 8. Autoradiographic analyses of total and NaOH- eluted radioactivity from TGL308 [SEQ ID NO: 8] primer extension reactions. Primer TGL308 was used to analyze the genotypes of individuals ESB164 and EA2014 as outlined in Example 5 and FIGS. 7 and 8. Total bead-associated radioactivity was determined by directly spotting a suspension of beads containing 75 fmoles of primer onto filter paper followed by autoradiographic detection of the label in the spot. Radioactivity specifically associated with the TGL308 primer was determined by magnetically immobilizing the beads, eluting the primer with NaOH as described in Examples 4 and 5, and spotting on filter paper an amount corresponding to 75 fmoles. Label in these spots was also detected by autoradiography.

FIG. 9. Data is shown from GBA on single stranded nucleic acid produced by asymmetric PCR from human DNA samples of different genotypes. The DNA sequence being interrogated [SEQ ID NO: 23] is from the HLA DPA1 locus at the polymorphic sequence coding for amino acid 31 of the DP alpha chain (Marsh, S. G. E. and Bodmer, J. G., HLA Class II Nucleotide Sequences, 1991. Human Immunol. 31, 207-227 [1991]) and is shown in the middle of the figure. Identification of the nucleotide immediately downstream of the primer is accomplished by enzyme-linked detection and is visualized as an orange color change in the well corresponding to the nucleotide which is inserted by the T7 DNA polymerase. Homozygotes only have one positive well, heterozygotes have two. The sequence of the GBA primer [SEQ ID NO: 25 and 26] is indicated by an arrow whose tail is the 5' end and head is the 3' end of the oligonucleotide.

FIG. 10. Data is shown from GBA on single stranded nucleic acid produced by asymmetric PCR from equine DNA samples of different genotypes. The DNA sequence being interrogated [SEQ ID NO: 25] is from the HLA DPA1 locus at the polymorphic sequence coding for amino acid 50 of the DP alpha chain Marsh, S. G. E. and Bodmer, J. G., HLA Class II Nucleotide Sequences, 1991. Human Immunol. 31, 207-227 [1991]) and is shown in the middle of the Figure.

FIG. 11. Data is shown from GBA on single stranded nucleic acid produced by asymmetric PCR from equine DNA samples of different genotypes. The DNA sequence being interrogated is from the anonymous locus JH85 [SEQ ID NO: 24] at the polymorphic sequence at nucleotide number 122 with respect to the original cloned genomic piece (unpublished results) and is shown in the middle of the figure. At this position, the "B" allele contains one extra base. For this reason, a different nucleotide position is interrogated by primer #307 [SEQ ID NO: 29] as compared to #308 [SEQ ID NO: 30]. Nevertheless, the results of both strand interrogations allow for unambiguous typing.

FIG. 12. Data shown are the results of a quantitative GBA of equine locus JH85. Following addition of substrate, the microplate was read kinetically, in a "Vmax" model 96-well spectrophotometer (Molecular Devices, Inc., Menlo Park, Calif.). Values are expressed as a Vmax in milli OD units per minute. The GBA results for the AA homozygote (solid bars), the AB heterozygote (open bars), and BB homozygote (stippled bars) single stranded templates is indicated for the four biotinylated ddNTPs analyzed in separate wells. Numerical values obtained are indicated at the top of each bar.

#### DETAILED DESCRIPTION OF THE INVENTION

The subject invention provides a reagent composition comprising an aqueous carrier and an admixture of at least

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two different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction in a manner strictly dependent on the identity of the unpaired nucleotide base in the template immediately adjacent to, and downstream of, the 3' end of the primer. In addition, at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and at least one of the terminators is labeled with a detectable marker.

The subject invention further provides a reagent composition comprising an aqueous carrier and an admixture of four different terminators of a nucleic acid template-dependent, primer extension reaction. Each of the terminators is capable of specifically terminating the extension reaction as above and two, three, or four of the terminators are labeled with a different detectable marker.

The subject invention further provides a reagent as described above wherein the terminators comprise nucleotides, nucleotide analogs, dideoxynucleotides, or arabinoside triphosphates. The subject invention also provides a reagent wherein the terminators comprise one or more of dideoxyadenosine triphosphate (ddATP), dideoxycytosine triphosphate (ddCTP), dideoxyguanosine triphosphate (ddGTP), dideoxythymidine triphosphate (ddTTP), or dideoxyuridine triphosphate (ddUTP).

The subject invention further provides a reagent as described above wherein each of the detectable markers attached to the terminators is an isotopically labeled moiety, a chromophore, a fluorophore, a protein moiety, or a moiety to which an isotopically labeled moiety, a chromophore, a fluorophore, or a protein moiety can be attached. The subject invention also provides a reagent wherein each of the different detectable markers is a different fluorophore.

The subject invention also provides a reagent as described above wherein the reagent further comprises pyrophosphatase.

The invented reagent consists of two or more chain terminators with one or more of the chain terminators being identifiably tagged. This reagent can be used in a DNA polymerase primer extension reaction to type nucleic acid sequences of interest that are complementary to one or more oligonucleotide primers by chemically or physically separating the polymerase extended primers from the chain terminator reagent and analyzing the terminal additions. Any kind of terminator that inhibits further elongation can be used, for example, a dideoxynucleoside triphosphate. Several approaches can be used for the labeling and detection of terminators: (1) radioactivity and its detection by either autoradiography or scintillation counting, (2) fluorescence or absorption spectroscopy, (3) mass spectrometry, or (4) enzyme activity, using a protein moiety. The identity of each terminator can be determined individually, i.e., one at a time. In addition, methods which permit independent analyses of each of the terminators permit analysis of incorporation of up to four terminators simultaneously.

The subject invention also provides a method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases

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spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with a stretch of nucleotide bases present in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest. Enzymatic extension of the oligonucleotide primer in the resultant duplex by one nucleotide, catalyzed, for example, by a DNA polymerase, thus depends on correct base pairing of the added nucleotide to the nucleotide base to be identified.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer.

The net result is that the oligonucleotide primer has been extended by one terminator. Next, the identity of the detectable marker present at the 3' end of the extended primer is determined. The identity of the detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

The subject invention also provides another method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest. First, a sample containing the nucleic acid of interest is treated, if such nucleic acid is double-stranded, so as to obtain unpaired nucleotide bases spanning the specific position. If the nucleic acid of interest is single-stranded, this step is not necessary. Second, the sample containing the nucleic acid of interest is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with nucleotide bases in the nucleic acid of interest, immediately adjacent to the nucleotide base to be identified, so as to form a duplex between the primer and the nucleic acid of interest such that the nucleotide base to be identified is the first unpaired base in the template immediately downstream of the 3' end of the primer in the duplex of primer and the nucleic acid of interest.

The duplex of primer and the nucleic acid of interest is then contacted with a reagent containing four terminators, only one of the terminators having a detectable marker. The duplex of primer and the nucleic acid of interest is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the nucleotide base to be identified and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. The net result is that the oligonucleotide primer has been extended by one terminator.

The original duplex of primer and the nucleic acid of interest is then contacted with three different reagents, with a different one of each of the four terminators being labeled

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in each of the four parallel reaction steps. Next, the products of the four parallel template-dependent, primer extension reactions are examined to determine which of the products has a detectable marker. The product with a detectable marker indicates which terminator has base paired to the next base in the nucleic acid of interest. Since the terminator is complementary to the next base in the nucleic acid of interest, the identity of the next base in the nucleic acid of interest is thereby determined.

Both of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest label the primer after hybridization between the primer and the template. If the template-dependent enzyme has no exonuclease function, the 3' end of the primer must be base paired for the labeling to occur.

The subject invention also provides a method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids. First, the sample of nucleic acids is treated, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids. If the nucleic acids in the sample are single-stranded, this, step is not necessary. Second, the sample of nucleic acids is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with the particular nucleotide sequence, if the particular nucleotide sequence is present, so as to form a duplex between the primer and the particular nucleotide sequence.

The duplex of primer and the particular nucleotide sequence, if any, is then contacted with a reagent containing four labeled terminators, each terminator being labeled with a different detectable marker. The duplex of primer and the particular nucleotide sequence, if any, is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the unpaired template nucleotide base downstream of the 3' end of the primer, the primer being hybridized with the particular nucleotide sequence in the template, and the occurrence of a template-dependent, primer extension reaction so as to incorporate the terminator at the 3' end of the primer. Next, the absence or presence and identity of a detectable marker at the 3' end of the primer are determined. The presence or absence of the detectable marker indicates whether the primer has hybridized to the template. If a detectable marker is absent, the primer did not hybridize to the template, and, therefore, the particular nucleotide sequence is not present in the sample of nucleic acids. If a detectable marker is present, the primer did hybridize to the template, and, therefore, the particular nucleotide sequence is present in the sample of nucleic acids.

The subject invention also provides another method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids. First, the sample of nucleic acids is treated, if such sample of nucleic acids contains double-stranded nucleic acids, so as to obtain single-stranded nucleic acids. Second, the sample of nucleic acids is contacted with an oligonucleotide primer under hybridizing conditions. The oligonucleotide primer is capable of hybridizing with the particular nucleotide sequence, if the particular nucleotide sequence is present, so as to form a duplex between the primer and the particular nucleotide sequence.

The duplex of primer and the particular nucleotide sequence, if any, is then contacted with a reagent containing four terminators, only one of the terminators having a detectable marker. The duplex of primer and the particular

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nucleotide sequence, if any, is contacted with the reagent under conditions permitting base pairing of a complementary terminator present in the reagent with the unpaired template nucleotide base downstream of the 3' end of the primer, the primer being hybridized with the particular nucleotide sequence in the template, and the occurrence of a template-dependent, primer extension reaction. The net result is the incorporation of the terminator at the 3' end of the primer.

The original duplex of primer and the particular nucleotide sequence, if any, is then contacted with three different reagents, with a different one of each of the four terminators being labeled in each of the four parallel reaction steps. Next, the products of the four parallel, template-dependent, primer extension reactions are examined to determine which, if any, of the products have detectable markers. The absence or presence and identity of the detectable marker indicates whether the primer has hybridized to the template. If no detectable marker is present in any of the products, the primer did not hybridize to the template, and, therefore, the particular nucleotide sequence was not present in the sample of nucleic acids. If a detectable marker is present in any of the products, the primer did hybridize to the template, and, therefore, the particular nucleotide sequence was present in the sample of nucleic acids.

Different versions of the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest and the method for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids are possible. In the first version, the template is a deoxyribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a DNA polymerase. This version gives a DNA product. In a second version, the template is a ribonucleic acid, the primer is an oligodeoxyribonucleotide, oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides, and the template-dependent enzyme is a reverse transcriptase. This version gives a DNA product. In a third version, the template is a deoxyribonucleic acid, the primer is an oligoribonucleotide, and the enzyme is an RNA polymerase. This version gives an RNA product. In a fourth version, the template is a ribonucleic acid, the primer is an oligoribonucleotide, and the template-dependent enzyme is an RNA replicase. This version gives an RNA product.

Preferably, before the primer extension reaction is performed, the template is capped by the addition of a terminator to the 3' end of the template. The terminator is capable of terminating a template-dependent, primer extension reaction. The template is capped so that no additional labeled terminator will attach at the 3' end of the template. The extension reaction should occur on the primer, not on the template. A dideoxynucleotide can be used as a terminator for capping the template.

Another modification of the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest is to separate the primer from the nucleic acid of interest after the extension reaction by using appropriate denaturing conditions. The denaturing conditions can comprise heat, alkali, formamide, urea, glyoxal, enzymes, and combinations thereof. The denaturing conditions can also comprise treatment with 2.0 N NaOH.

The nucleic acid of interest can comprise non-natural nucleotide analogs such as deoxyinosine or 7-deaza-2'-deoxyguanosine. These analogues destabilize DNA

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duplexes and could allow a primer annealing and extension reaction to occur in a double-stranded sample without completely separating the strands.

The sample of nucleic acids can be from any source. The sample of nucleic acids can be natural or synthetic (i.e., synthesized enzymatically in vitro). The sample of nucleic acids can comprise deoxyribonucleic acids, ribonucleic acids, or copolymers of deoxyribonucleic acid and ribonucleic acid. The nucleic acid of interest can be a deoxyribonucleic acid, a ribonucleic acid, or a copolymer of deoxyribonucleic acid and ribonucleic acid. The nucleic acid of interest can be synthesized enzymatically in vivo, synthesized enzymatically in vitro, or synthesized non-enzymatically. The sample containing the nucleic acid or acids of interest can comprise genomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof. The sample containing the nucleic acid or acids of interest can also comprise extragenomic DNA from an organism, RNA transcripts thereof, or cDNA prepared from RNA transcripts thereof. Also, the nucleic acid or acids of interest can be synthesized by the polymerase chain reaction.

The sample can be taken from any organism. Some examples of organisms to which the method of the subject invention is applicable include plants, microorganisms, viruses, birds, vertebrates, invertebrates, mammals, human beings, horses, dogs, cows, cats, pigs, or sheep.

The nucleic acid of interest can comprise one or more moieties that permit affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer. The nucleic acid of interest can comprise biotin which permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via binding of the biotin to streptavidin which is attached to a solid support. The sequence of the nucleic acid of interest can comprise a DNA sequence that permits affinity separation of the nucleic acid of interest from the unincorporated reagent and/or the primer via base pairing to a complementary sequence present in a nucleic acid attached to a solid support. The nucleic acid of interest can be labeled with a detectable marker; this detectable marker can be different from any detectable marker present in the reagent or attached to the primer.

The oligonucleotide primer can be an oligodeoxyribonucleotide, an oligoribonucleotide, or a copolymer of deoxyribonucleotides and ribonucleotides. The oligonucleotide primer can be either natural or synthetic. The oligonucleotide primer can be synthesized either enzymatically in vivo, enzymatically in vitro, or non-enzymatically in vitro. The oligonucleotide primer can be labeled with a detectable marker; this detectable marker can be different from any detectable marker present in the reagent or attached to the nucleic acid of interest. In addition, the oligonucleotide primer must be capable of hybridizing or annealing with nucleotides present in the nucleic acid of interest, immediately adjacent to, and upstream of, the nucleotide base to be identified. One way to accomplish the desired hybridization is to have the template-dependent primer be substantially complementary or fully complementary to the known base sequence immediately adjacent to the base to be identified.

The oligonucleotide primer can comprise one or more moieties that permit affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest. The oligonucleotide primer can comprise biotin which permits affinity separation of the primer from the unincor-

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porated reagent and/or nucleic acid of interest via binding of the biotin to streptavidin which is attached to a solid support. The sequence of the oligonucleotide primer can comprise a DNA sequence that permits affinity separation of the primer from the unincorporated reagent and/or the nucleic acid of interest via base pairing to a complementary sequence present in a nucleic acid attached to a solid support.

The subject invention also provides a method of typing a sample of nucleic acids which comprises identifying the base or bases present at each of one or more specific positions, each such nucleotide base being identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. Each specific position in the nucleic acid of interest is determined using a different primer. The identity of each nucleotide base or bases at each position can be determined individually or the identities of the nucleotide bases at different positions can be determined simultaneously.

20 The subject invention also provides another method of typing a sample of nucleic acids which comprises determining the presence or absence of one or more particular nucleotide sequences, the presence or absence of each such nucleotide sequence being determined using one of the methods for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids as outlined above.

25 The subject invention also provides an additional method of typing a sample containing nucleic acids. First, the presence or absence of one or more particular nucleotide sequences is determined; the presence or absence of each such nucleotide sequence is determined using one of the methods for determining the presence or absence of a particular nucleotide sequence in a sample of nucleic acids as outlined above. Second, the nucleotide base or bases present at each of one or more specific positions is identified; each such base is identified using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

30 35 40 The subject invention further provides a method for identifying different alleles in a sample containing nucleic acids which comprises identifying the base or bases present at each of one or more specific positions. The identity of each nucleotide base is determined by the method for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above.

The subject invention also provides a method for determining the genotype of an organism at one or more particular genetic loci which comprises obtaining from the organism a sample containing genomic DNA and identifying the nucleotide base or bases present at each of one or more specific positions in nucleic acids of interest. The identity of each such base is determined by using one of the methods for determining the identity of a nucleotide base at a specific position in a nucleic acid of interest as outlined above. The identity of the nucleotide bases determine the different alleles and, thereby, determine the genotype of the organism at one or more particular genetic loci.

45 50 55 60 65 The chain termination reagent in combination with an appropriate oligonucleotide primer, and a DNA polymerase with or without an associated 3' to 5' exonuclease function, and an appropriate salt and cofactor mixture, can be used under appropriate hybridization conditions as a kit for diagnosing or typing nucleic acids, if appropriate primer separation techniques are used. To simplify the primer separation and the terminal nucleotide addition analysis this

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invention makes use of oligonucleotides that are modified in such ways that permit affinity separation as well as polymerase extension. The 5' termini and internal nucleotides of synthetic oligonucleotides can be modified in a number of different ways to permit different affinity separation approaches, e.g., biotinylation. These affinity reagents can be used with the terminator mixture to facilitate the analysis of extended oligonucleotide(s) in two ways:

(1) If a single affinity group is used on the oligonucleotide(s), the oligonucleotide(s) can be separated from the unincorporated terminator reagent. This eliminates the need of physical or size separation.

(2) More than one oligonucleotide can be separated from the terminator reagent and analyzed simultaneously if more than one affinity group is used. This permits the analysis of several nucleic acid species or more nucleic acid sequence information per extension reaction.

The affinity group(s) need not be on the priming oligonucleotide but could, alternatively, be present on the template. As long as the primer remains hydrogen bonded to the template during the affinity separation step, this will allow efficient separation of the primer from unincorporated terminator reagent. This also has the additional benefit of leaving sites free on the primer for the convenient attachment of additional moieties. For example, the 5'-terminus of the primer could be modified by coupling it to a suitable fluorescent group such as rhodamine, allowing the amount of primer in the primer:template complex to be easily quantified after the affinity separation step. The amounts of 3'-terminating terminators could then be normalized to the total amount of annealed primer.

The oligonucleotide primers and template can be any length or sequence, can be DNA or RNA, or any modification thereof. It is necessary, however, that conditions are chosen to optimize stringent hybridization of the primers to the target sequences of interest.

The conditions for the occurrence of the template-dependent, primer extension reaction can be created, in part, by the presence of a suitable template-dependent enzyme. Some of the suitable template-dependent enzymes are DNA polymerases. The DNA polymerase can be of several types. The DNA polymerase must, however, be primer and template dependent. For example, *E. coli* DNA polymerase I or the "Klenow fragment" thereof, T4 DNA polymerase, T7 DNA polymerase ("Sequenase"), *T. aquaticus* DNA polymerase, or a retroviral reverse transcriptase can be used. RNA polymerases such as T3 or T7 RNA polymerase could also be used in some protocols. Depending upon the polymerase, different conditions must be used, and different temperatures ranges may be required for the hybridization and extension reactions.

The reagents of the subject invention permit the typing of nucleic acids of interest by facilitating the analysis of the 3' terminal addition of terminators to a specific primer or primers under specific hybridization and polymerase chain extension conditions. Using only the terminator mixture as the nucleoside triphosphate substrate ensures addition of only one nucleotide residue to the 3' terminus of the primer in the polymerase reaction. Using all four terminators simultaneously ensures fidelity, i.e., suppression of misreading.

By specifically labeling one or more of the terminators, the sequence of the extended primer can be deduced. In principle, more than one reaction product can be analyzed per reaction if more than one terminator is specifically labeled.

By specifically tagging the oligonucleotide primer(s), or template(s) with a moiety that does not affect the 3' exten-

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sion reaction yet permits affinity separation, the extension product(s) can be separated post-reaction from the unincorporated terminators, other components of the reagents, and/or the template strand. Several oligonucleotides can be analyzed per extension reaction if more than one affinity agent is used.

In principle, the combination of four differently labeled terminators and many primers or templates tagged with different groups permits the typing of many different nucleic acid sequences simultaneously. Specificity in this diagnostic reaction is determined by

- (1) the stringency of oligonucleotide hybridization and
- (2) the sequence information gained by the single residue extension.

#### A. GENERAL METHODS

##### 1. Biotinylation of Oligodeoxynucleotides

Oligodeoxynucleotides, terminated at their 5'-ends with a primary amino group, were ordered from Midland Certified Reagents, Midland, Texas. These were biotinylated using biotin-XX-NHS ester (Clontech Laboratories, Inc., Palo Alto, Calif.), a derivative of biotin-N-hydroxysuccinimide. Reagents used were from the Clontech biotinylation kit. Typically, the oligonucleotide (9 nanomoles) was dissolved in 100  $\mu$ l of 0.1M NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> (pH 9), and 25  $\mu$ l of N,N-dimethylformamide containing 2.5 mg biotin-XX-NHS-ester was added. The mixture was incubated overnight at room temperature. It was then passed over a 6 ml Sephadex G-25 column ("DNA grade"—Pharmacia) equilibrated with H<sub>2</sub>O. Eluate fractions containing DNA were identified by mixing 4  $\mu$ l aliquots with an equal volume of ethidium bromide (2  $\mu$ g/ml) and the DNA-induced fluorescence was monitored with a UV transilluminator. Unreacted ester was detected by UV absorption at 220 nm. The tubes containing DNA were pooled, concentrated in a Centricon-3 microconcentrator (Amicon), and passed over Sephadex again.

Inhibition of the binding of [<sup>3</sup>H]-biotin to magnetic M-280 streptavidin Dynabeads (Dynal) was used to assay quantitatively the extent of biotinylation of the oligonucleotides. Eppendorf tubes and pipet tips were siliconized. A known amount (5–10 pmoles) of biotin-labeled oligonucleotide in 10  $\mu$ l 0.1 M NaCl was added to tubes containing 25  $\mu$ l of 1:4 suspension of beads in 0.1 M NaCl. The tubes were rotated for one hour on a Labquake shaker (Labindustries, Inc.). Increasing amounts of [<sup>3</sup>H]-biotin (5–35 pmoles) in 20  $\mu$ l of 0.1 M NaCl were added to the tubes and these were rotated again for one hour. Tubes were put on a Dynal MPC-E magnet to remove the beads from suspension, 10  $\mu$ l aliquots of the supernatant were withdrawn, and the amount of radioactivity in these was measured using a Beckman LS 5000 TD liquid scintillation counter. Counts were compared to those from tubes to which no oligonucleotide had been added. Alternatively, for some primers, biotinylation was monitored by size fractionation of the reaction products using analytical polyacrylamide gel electrophoresis in the presence of 8 M urea.

##### 2. Template-dependent Primer Extension/termination Reactions

Approximately five pmoles of 5'-biotinylated oligodeoxynucleotide template (see above) were mixed with approximately three pmoles of primer in 1- sequencing buffer (from Sequenase Version 2.0 kit, US Biochemical Corp.) (10  $\mu$ l final volume), the mixture was incubated at 65° C. for 2 min,

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then allowed to cool to room temperature in order to anneal the primer and template. The solution containing the annealed template-primer was separated into two 5  $\mu$ l portions, A and B, to which were added the following: Reactions A (for normalizing template concentrations)—0.5  $\mu$ l of 100 mM dithiothreitol, 1  $\mu$ l each of 10  $\mu$ M dATP, dGTP, ddCTP, 0.5  $\mu$ l of "Mn buffer" (from Sequenase Version 2.0 kit, US Biochemical Corp.), 0.5  $\mu$ l of [ $^3$ S]- $\alpha$ -thio-dTTP (10 mCi/ml, 1180 Ci/mmol) (Dupont-NEN), 1  $\mu$ l of Sequenase (1:8 dilution, US Biochemical Corp.); Reactions B (for template-specific labeling of primer 3'-ends)—same additions as in Reactions A except the nucleotides used were ddCTP, ddGTP, ddTTP, and [ $^{35}$ S]- $\alpha$ -thio-ddATP.

Reactions were for 5 min at 37° C. Control reactions omitting the primer or the Sequenase were also performed. Aliquots were removed and analyzed by electrophoresis on a 15% polyacrylamide, 8 M urea, DNA sequencing gel (see Maniatis, T., et al., *Molecular Cloning, a Laboratory Manual*, Cold Spring Harbor Laboratory (1982)). The gel was fixed in 10% methanol, 10% acetic acid, dried down onto Whatman's 3MM paper, and exposed to Kodak X-Omat AR film. Alternatively, for purposes of analyzing the products by liquid scintillation counting, the biotinylated template or template-primer was bound to an excess of M-280 streptavidin Dynabeads (Dynal) before or after the Sequenase reaction (see above, "1. Biotinylation of oligodeoxynucleotides", for binding conditions). Beads were washed three times with 0.1 M NaCl to remove unincorporated label, then scintillation fluid was added and the radioactivity measured by liquid scintillation counting.

### 3. Generation of Templates from Polymerase Chain Reaction Products

Polymerase chain reaction (PCR) reactions were carried out where one or the other of the amplification primers flanking the target stretch of DNA were biotinylated as described above. These primers (2  $\mu$ mol final concentration) and the target DNA (up to 1  $\mu$ g) were incubated with 2.5 units of Taq polymerase (Perkin Elmer/Cetus), 200  $\mu$ M each of DATP, dCTP, dGTP, and dTTP, 10 mM Tris-HCl (pH 8.3), 50 mM KC1, 1.5 mM MgCl<sub>2</sub>, and 0.01% gelatin (Sigma). Reaction mixtures were overlayed with paraffin oil and incubated for 30 cycles in Perkin Elmer/Cetus thermocycler. Each cycle consisted of 1 min at 94° C., 2 min at 60° C., and 3 min at 72° C. Reaction products were purified by phenol/chloroform extraction and ethanol precipitation, then analyzed by ethidium bromide staining after electrophoresis on a polyacrylamide gel. The yield of duplex PCR product was typically about 10  $\mu$ g. Approximately 5  $\mu$ g of this PCR product was incubated with gentle agitation for 60 min with 50  $\mu$ L of a suspension of prewashed M-280 Dynabeads in 0.1 M NaCl. The beads with the bound DNA (approximately 15 pmoles) were then incubated for 5 min at 25° C. with 0.15 M NaOH. Beads were washed once with 0.15 M NaOH to remove the unbiotinylated DNA strand, then washed three times with H<sub>2</sub>O. The beads were resuspended in H<sub>2</sub>O and the strand bound to the beads via the biotin-streptavidin link was used as template for further primer extension reactions.

## B. EXAMPLES

### Example 1

Primer oligo 182: 5' GCCTTGGCGTTGTAGAA3' [SEQ ID NO: 1] Template oligos 180(C)/181(T): 3' TCGGGTCGGAACCGCAACAT CTTC/TATAGACTA5' [SEQ ID NO: 2].

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Oligonucleotides 180 and 181 were synthesized with primary amino groups attached to their 5' termini. These were coupled with biotin as described above. Oligonucleotide 182 was annealed as a primer and extension reactions "A" and "B" (see above) were carried out. The expected template-dependent 3'-terminal extensions to oligonucleotide 182 were as follows ("\*" preceding a nucleotide signifies a radioactive label):

| Template | Reaction A         | Reaction B |
|----------|--------------------|------------|
| 180      | -dG-*dT-dA-*dT-ddC | -ddG       |
| 181      | -dA-*dT-dA-*dT-ddC | -*ddA      |

Thus, in the "A" reactions, both template oligonucleotides will direct a radioactively-labelled five nucleotide extension of the primer; the amount of labeling should be proportional to the amount of productively primed template present in the reactions. In the "B" reactions, both templates will direct a one nucleotide extension of the primer, but only for template 181 should this result in labeling of the primer. The "B" reaction, therefore, is an example of template-directed, sequence-specific labeling of an oligonucleotide via DNA polymerase-catalyzed extension of a productive primer-template complex.

The reaction products were fractionated by size on a 15% polyacrylamide/8M urea sequencing gel and visualized by autoradiography. The results (FIG. 1) show that, as expected, the "A" reactions yield labeling and extension of both primers whereas the "B" reaction results in labeling that is strongly biased in favor of template 181. Panel C in FIG. 1 shows a gel analysis of the same reaction products as in Panel B, except the reaction products were first purified as described above using M-280 streptavidin Dynabeads.

### Example 2

The experiment described in Example 1 shows template-directed labeling of oligonucleotide primer 182 [SEQ ID NO: 1] in which the labeling is specific with respect to oligonucleotides or other species that migrate similarly on a polyacrylamide gel. In order to assess more generally the template-directed specific labeling of oligonucleotide 182 with respect to all other labeled species, regardless of gel mobility, a direct measurement of incorporated radioactivity was performed. In this experiment, both reactions "A" and "B" were performed, reaction products were purified using Dynabeads, and total radioactivity in the aliquots was measured by liquid scintillation counting. This procedure assesses both misincorporation of label into other species and, in addition, the efficiency of the Dynabead washing procedure with respect to unincorporated nucleotides. As a practical matter, it would be of interest to minimize both sources of non-specific label in order to have a simple, non-gel-based, procedure for assessing specific, template directed labeling of the primer. The results of directly counting the reaction products after washing on the magnetic beads are as follows (all results expressed as cpm of [ $^{35}$ S]):

| Reaction         | Template 180 | Template 181 |
|------------------|--------------|--------------|
| A, complete      | 325,782      | 441,823      |
| A, no polymerase | 5,187        | 5,416        |

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## Example 4

-continued

| Reaction         | Template 180 | Template 181 |
|------------------|--------------|--------------|
| A, no primer     | 4,351        | 12,386       |
| B, complete      | 5,674        | 176,291      |
| B, no polymerase | 2,988        | 1,419        |
| B, no primer     | 1,889        | 1,266        |

As can be seen from these results, specific template-directed labeling of primer 182 can also be determined by measuring the total radioactivity of the reaction products after washing with magnetic beads to remove unreacted nucleotides. The background in this experiment due to nonspecific label from all other sources was approximately 3-4% (compare templates 180 and 181 in the "B, complete" reaction). Control experiments ("no polymerase" and "no primer") showed that the bulk of the background label was probably contributed by unincorporated nucleotides that were not completely removed by the washing step. The "A, complete" reactions showed that, for both templates, productive template:primer complexes were present.

## Example 3

Two amplification primers, TGL 105 [SEQ ID NO: 15] and TGL 106 [SEQ ID NO: 16] (FIG. 2), were used to amplify a cloned stretch of bovine DNA containing two DNA sequence polymorphisms: a C or T at position 114 and an A and G at position 190 [SEQ ID NO: 19] (FIG. 2). DNAs containing those polymorphisms were molecularly cloned and available on plasmids, as follows: plasmid p183, C114 and A190; plasmid p624, T114 and A190; plasmid p814, C114 and CG190. Four PCR reactions with biotinylated primers were performed to amplify and purify specific strands of these plasmids for use as templates:

| Primers                | Plasmids      | Detection Primers       |
|------------------------|---------------|-------------------------|
| 105 biotinylated       | p183 and p624 | TGL 182 [SEQ ID NO: 17] |
| 106 unbiotinylated     |               |                         |
| 105 unbiotinylated, 40 | p183 and p814 | TGL 166 [SEQ ID NO: 18] |
| 106 biotinylated       |               |                         |

The duplex PCR products were bound to magnetic microspheres, denatured with NaOH, and biotinylated strand purified as described above. Templates prepared with biotinylated TGL 105 were subjected to analysis by DNA sequencing with unbiotinylated primer TGL 106 in order to measure the amount of template present. Similarly, template prepared using biotinylated TGL 106 was analyzed by sequencing with unbiotinylated TGL 105.

Approximately equal amounts of template (2 pmoles) were annealed for 5 min at 65° C. to the polymorphism detection primers, TGL 182 [SEQ ID NO: 17] and TGL 166 [SEQ ID NO: 18] (see above and FIG. 2). These primers hydrogen-bond to the templates in a sequence-specific fashion such that their 3'-termini are adjacent to nucleotide positions 114 and 190, respectively (FIG. 2) [SEQ ID NO: 19]. Template-directed primer extension reactions (reaction "B" conditions) were carried out on these primer:template complexes in the presence of the four ddNTPs, one of which (ddATP) was labeled. The products of these extension reactions were analyzed by electrophoresis on a 15% polyacrylamide/8M urea gel followed by autoradiography (FIG. 3).

Primer oligo TGL391: 5'TGTTTGACAAAAGCA3'  
[SEQ ID NO 3]

Primer oligo TGL346: 5'GTTTTGCACAAAAGCAT3'  
[SEQ ID NO 4]

Template oligo TGL382: 3'ACAAAAACGTGTTTCGT  
AGGAS'-biotin: (streptavidin-bead) [SEQ ID NO: 5].

Oligonucleotide TGL382 was purchased from the Midland Certified Reagent Company, Midland, Tex. It was biotinylated using Midland Certified Reagent Company's "Biotin dX" reagent (a biotin derivative phosphoramidite) which is suitable for use in automated DNA synthesis in the 5' terminal nucleotide position. The biotinylated oligonucleotide was then purified by anion exchange HPLC. Streptavidin-conjugated M-280 Dynabeads were washed in TNET buffer (10 mM Tris-HCl, pH 7.5, 100 mM NaCl, 1 mM EDTA, 0.1% Triton X-100) and resuspended in the same buffer at a concentration of 7×10<sup>8</sup> beads/ml. 10-100 pmoles of biotinylated oligonucleotide TGL382 was incubated with 100 µl of the Dynabead suspension in TNET for 30 minutes at 20° C. in order to allow the biotin moiety to bind to the streptavidin. The beads were then washed (using a magnet to immobilize them) three times with 200 µl of TNET and resuspended in 100 µl of TNET. For annealing, 25 µl of this suspension of the Dynabeads with the attached template oligonucleotide was immobilized with the magnet, the TNET withdrawn, and 25 µl of 40 mM Tris-HCl, pH 7.5, 20 mM MgCl<sub>2</sub>, 50 mM NaCl, containing 2 µM of oligonucleotide primers 346 or 391, was added. The template and each primer were annealed by incubating them for 5 minutes at 65° C., followed by slow cooling over a period of 20 minutes to room temperature. Beads containing the bound template-primer complexes were washed twice with 200 µl TNET, followed by resuspension in 25 µl of 40 mM Tris-HCl, pH 7.5, 20 mM MgCl<sub>2</sub>, 50 mM NaCl.

The Following ddNTP Mixes Were Used:

<sup>35</sup>S-labelled dideoxynucleoside triphosphate mixes (labelled nucleotide indicated in the form ddN\*TP):

|          |             |             |             |
|----------|-------------|-------------|-------------|
| ddG Mix: | 5 µM ddG*TP | 10 µM ddATP | 10 µM ddTTP |
| ddA Mix: | 10 µM ddGTP | 5 µM ddA*TP | 10 µM ddTTP |
| ddT Mix: | 10 µM ddGTP | 10 µM ddATP | 5 µM ddT*TP |
| ddC Mix: | 10 µM ddGTP | 10 µM ddATP | 10 µM ddTTP |
|          | 5 µM ddCTP  |             |             |

The ddN\*TPs were the four respective [ $\alpha$ -thio-<sup>35</sup>S] dideoxynucleoside triphosphates (purchased from New England Nuclear).

For each bead-bound, template-primer complex, four extension reactions were carried out, one reaction for each of the four ddNTP mixes. Extension reactions contained the following components: 5.0 µl bead suspension containing the annealed template-primer complex, 0.5 µl of 100 mM dithiothreitol, 0.5 µl of "Mn<sup>++</sup> solution" (100 mM MnCl<sub>2</sub>, 150 mM DL-isocitrate, pH 7.0; purchased from U.S. Biochemicals, Cleveland, Ohio), 1.0 µl of ddG, ddA, ddT, or ddC mix, 2.0 µl of H<sub>2</sub>O, and 1.0 µl of T7 DNA polymerase ("Sequenase", version 2.0, US Biochemicals, 1625 units/ml in 50 mM Tris-HCl, pH 7.5, 10 mM 2-mercaptoethanol, 1 mg/ml bovine serum albumin).

Reactions were allowed to proceed for 15 minutes at 20° C., then stopped by washing the magnetically immobilized

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beads three times with 500  $\mu$ l TNET. Beads were resuspended in final volume of 25  $\mu$ l TNET prior to the detection assays.

Incorporation of labelled dideoxynucleotides by the primer extension reaction was assayed two different ways: gel electrophoresis followed by autoradiography, and direct autoradiographic analysis of labelled DNA.

1. Gel electrophoresis followed by autoradiography ( $^{35}$ S-labelled material only). Samples of washed, bead-bound DNA were heated at 94° C. for 5 minutes in 10  $\mu$ l of formamide loading buffer (80% formamide, 10 mM Tris-HCl, pH 8, 1 mM EDTA, 0.02% bromophenol blue) to denature the DNA and release the labelled primer from the primer:template complex. Samples were analyzed by electrophoresis on 8 or 12.5% polyacrylamide/8 M urea sequencing gels (19:1 acrylamide:bis-acrylamide ratio; 100 mM Tris-HCl, 100 mM borate, 2 mM EDTA, pH 8.3, running buffer; 60 watts constant power). After electrophoresis, gels were either dried down onto filter paper or frozen at -80° C. to prevent diffusion, covered with plastic wrap, and exposed to X-ray film to visualize the labelled DNA by autoradiography (FIG. 4).

2. Direct autoradiographic analysis of labelled DNA. For the analysis of total radioactivity bound to the beads, 10  $\mu$ l aliquots of the bead suspensions in TNET were spotted directly onto filter paper or nylon membranes. Filters or membranes were dried under an incandescent lamp, covered with plastic wrap, and exposed to X-ray film (FIG. 5).

#### Example 5

TGL240: 5'AGATGATGCTTTGTGCAAAACAC3'  
[SEQ ID NO: 6]  
TGL239: 5'TCAATACCTGAGTCCCGACACCCCTG3'  
[SEQ ID NO: 7]  
TGL308: 5'AGCCTCAGACCGCGTGGTGCCTGGT3'  
[SEQ ID NO: 8]

Oligonucleotide TGL240 [SEQ ID NO: 6] was synthesized with a primary amino group attached to its 5' terminus and coupled with biotin as described above. TGL240 (biotinylated) and TGL239 (unbiotinylated) were used to amplify, via the polymerase chain reaction procedure (see "A. General Methods"), a region of DNA comprising a particular genetic locus in samples of mammalian genomic DNA. DNAs from two different individuals, each homozygous for a particular set of linked sequence polymorphisms (the "A" allele and the "i" allele—see FIG. 6), were examined. After the PCR reaction, 2-20 pmoles of duplex PCR DNA was incubated with 100  $\mu$ l of streptavidin-conjugated M-280 Dynabeads (7 $\times$ 10<sup>8</sup> beads/ml) in TNET buffer in order to bind the biotinylated strand to the beads. After binding, the beads were magnetically immobilized and washed three times with 200  $\mu$ l of TNET, then resuspended in 100  $\mu$ l of TNET. To remove the non-biotinylated strand, 500  $\mu$ l of 0.15 N NaOH was added and the suspension incubated for 30 minutes at 20° C. The beads were then magnetically immobilized and washed once with 250  $\mu$ l of 0.15 N NaOH, three times with 500  $\mu$ l TNET, and resuspended in 100  $\mu$ l of TNET.

The detection primer, oligonucleotide TGL308 [SEQ ID NO: 8] FIG. 6), was annealed to the bead-bound PCR-generated template as described above in Example 4. Further washes, extension reactions, and detection assays were also carried out as described in Example 4. A gel autoradiographic analysis of the labelled primer extension products for the two homozygous individuals, ESB164 ("AA" genotype) [SEQ ID NO: 20] and EA2014 ("BB" genotype)

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[SEQ ID NO: 21], is shown in FIG. 7. Autoradiographic analyses of total bead-bound radioactivity, or primer-associated radioactivity after NaOH elution, are shown for these same individuals using the filter spotting assay FIG. 8). For the analysis of primer only, 10  $\mu$ l of the bead suspension. After incubation for 10 minutes at room temperature, the beads were immobilized magnetically and the supernatant withdrawn and spotted onto nylon blotting membrane.

#### Example 6

##### Genetic Bit Analysis

DNA Samples. Genomic DNA was isolated using the SDS/Proteinase K procedure (Maniatis, T. Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, 1989) from peripheral blood nucleated cells of humans or horses enriched from red blood cells by selective lysis accomplished by diluting blood with a three fold volume excess of ACK lysis buffer (0.15 M ammonium chloride, 1 mM potassium bicarbonate, 0.1 mM EDTA). Oligonucleotides were prepared by solid-phase phosphoramidite chemistry using an Applied Biosystems, Inc. (Foster City, Calif.) Model 391 automated DNA synthesizer. In the case of primers used in Genetic Bit Analysis (GBA) reactions, detritylation was not performed following the final cycle of synthesis and the full-length oligonucleotide was purified using the Applied Biosystems oligonucleotide purification cartridge (OPC) as recommended by the manufacturer. For most PCR reactions, primers were used directly by drying down the de-protection reaction. oligonucleotides derivatized with 5'-amino groups were prepared using Aminolink 2 purchased from Applied Biosystems and used according the manufacturer's recommendations.

Primers for first round amplification of equine locus JH85 were #91: [ SEQ ID NO: 9] 5' CGTCTGCAGAATC-CAXTGGCTTCTTCTAG 3' and #92: [ SEQ ID NO:10] 5' GCAGGATCCTGGAACTACTCATTGCT 3'. Second round amplification of equine locus was achieved using nested primers #239: [ SEQ ID NO: 11] 5' TCAATACCT-GAGTCCCGACACCCCTG 3' and #240: [ SEQ ID NO: 12] 5' AGGATGATGCCTTGTGCAAAACAC 3'. Amplification of human HLA DPA1 sequences (Marsh, S. G. E., Bodmer, J. G. HLA Class II Nucleotide Sequences, 1991. Human Immunol. 31:207-227) was accomplished with primers #467: [ SEQ ID NO: 13] 5' GCGGACCATGTGT-CAACTTAT 3' and #445: [ SEQ ID NO: 14] 5' GCCT-GAGTGTGGTTGGAACCTG 3'.

Template Preparation. Amplification of genomic sequences was performed using the polymerase chain reaction (PCR) (Saiki, R. K., Gelfand, D. H., Stoffel, S., Scharf, S. J., Higuchi, R., Horn, G. T., Mullis, K. B., Erlich, H. A., Primer Directed Enzymatic Amplification of DNA with a Thermostable DNA Polymerase. Science 239:487-491). In a first step, one hundred nanograms of genomic DNA was used in a reaction mixture containing each first round primer at a concentration of 2  $\mu$ M/10 mM Tris pH 8.3/50 mM KC1/1.5 mM MgCl<sub>2</sub>/0.1% gelatin/0.05 units per  $\mu$ l Taq DNA Polymerase (AmpliTaq, Perkin Elmer Cetus, Norwalk, Conn.). Reactions were assembled and incubated at 94° C. for 1.5 minutes, followed by 30 cycles of 94° C./1 minute, 60° C./2 minutes, 72° C./3 minutes. Single stranded DNA was prepared in a second "asymmetric" PCR in which the products of the first reaction were diluted 1/1000. One of the primers was used at the standard concentration of 2  $\mu$ M while the other was used at 0.08  $\mu$ M. Under these conditions, both single stranded and double stranded molecules were synthesized during the reaction.

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Solid Phase Immobilization of Nucleic Acids. GBA reactions were performed in 96-well plates (Nunc Nunclon plates, Roskilde, Denmark). The GBA primer was covalently coupled to the plate by incubating 10 pmoles of primer having a 5' amino group per well in 50  $\mu$ l of 3 mM sodium phosphate buffer, pH 6, 20 mM 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide (EDC) overnight at room temperature. After coupling, the plate was washed three times with 10 mM Tris pH 7.5/150 mM NaCl/0.05% Tween-20 (TNTw).

Biotinylated ddNTPs. Biotinylated ddNTPs were synthesized according to U.S. Pat. No. 5,047,519.

GBA in Microwell Plates. Hybridization of single-stranded DNA to primers covalently coupled to 96-well plates was accomplished by adding an equal volume of 3M NaCl/50 mM EDTA to the second round asymmetric PCR and incubating each well with 20  $\mu$ l of this mixture at 55° C. for 30 minutes. The plate was subsequently washed three times with TNTw. Twenty (20)  $\mu$ l of polymerase extension mix containing ddNTPs (3  $\mu$ M each, one of which was biotinylated/5 mM DTT/7.5 mM sodium isocitrate/5 mM MnCl<sub>2</sub>/0.04 units per  $\mu$ l of modified T7 DNA polymerase and incubated for 5 minutes at room temperature. Following the extension reaction, the plate was washed once with TNTw. Template strands were removed by incubating wells with 50  $\mu$ l 0.2N NaOH for 5 minutes at room temperature, then washing the wells with another 50  $\mu$ l 0.2N NaOH. The plate was then washed three times with TNTw. Incorporation of biotinylated ddNTPs was measured by an enzyme-linked assay. Each well was incubated with 20  $\mu$ l of streptavidin-conjugated horseradish peroxidase (1/1000 dilution in TNTw of product purchased from BRL, Gaithersburg, Md.) with agitation for 30 minutes at room temperature. After washing 5 times with TNTw, 100  $\mu$ l of o-phenylenediamine (OPD, 1 mg/ml in 0.1 M Citric acid, pH 4.5) (BRL) containing 0.012% H<sub>2</sub>O<sub>2</sub> was added to each well. The amount of bound enzyme was determined by photographing the plate after stopping the reaction or quantitatively using a Molecular Devices model "Vmax" 96-well spectrophotometer.

In order to demonstrate the generality of the procedure, the ability to type three different sites located on two different template molecules is shown. In the middle of FIGS. 9 through 11 is shown the polymorphic region of these loci together with the sequence of the GBA primers used to genotype the DNA samples. The genotype of the test DNA samples was previously determined by restriction analysis and gel electrophoresis (equine samples) or by allele specific hybridization (human samples).

At the top and bottom of FIGS. 9 through 11 are photographs of the non-radioactive GBA analysis of these sites. Analysis of the "plus" strand (which corresponds to the mRNA for the HLA DPA1 but is arbitrarily chosen for the equine locus JH85) is shown at the top of the figure, analysis of the "minus" strand is shown in the lower photograph. Using horseradish peroxidase activity genotyping data was observed visually. Because both strands were suitable templates for GBA, it was possible to get genotypic confirmation by using two different primers. For the HLA DPA1 locus, two sites of variation were typed (FIGS. 9 and 10). Identical results were achieved. Spectrophotometric quantitation of a separate experiment involving the equine locus JH85 is shown in FIG. 12. The average ratio of signals obtained with expected vs. inappropriate base incorporation was 62.2.

### C. EMBODIMENTS

An example of one method to practice the present invention involves obtaining from a convenient source, such as

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blood, epithelium, hair, or other tissue, samples of DNA or RNA, then amplifying in vitro specific regions of the nucleic acid using the polymerase chain reaction, transcription-based amplification (see Kwok, et al., Proc. Natl. Acad. sci. 80:1173 (1989)), etc. Amplification is accomplished using specific primers flanking the region of interest, with one or more of the primers being modified by having an attached affinity group (although in any given reaction only one such primer is modified at a time). A preferred modification is attachment of biotin moieties to the 5'-termini of the primers. A sample (typically, 0.5-5 pmoles) of the amplified DNA is then bound to streptavidin-conjugated magnetic microspheres (e.g., Dynal M-280 "Dynabeads") via the attached biotin moiety on the amplification primer. The DNA is denatured by adjusting the aqueous suspension containing the microspheres to a sufficiently alkaline pH, and the strand bound to the microspheres via the biotin-streptavidin link is separated from the complementary strand by washing under similar alkaline conditions. To accomplish this, the microspheres are centrifuged or immobilized by the application of a magnetic field. The microsphere-bound strand is then used as a template in the remaining manipulations.

To the template strand, generated as described above, a specific primer oligonucleotide is bound under high stringency annealing conditions, the sequence of the primer being consistent with unique binding to a site on the template strand immediately adjacent to a known DNA sequence polymorphism. A preferred sequence and mode of binding for the primer ensures that the primer forms a duplex with the template such that the 3'-terminal nucleotide of the primer forms a Watson-Crick basepair with the template nucleotide immediately adjacent to the site of the first nucleotide in the sequence polymorphism, without the duplex overlapping any of the polymorphic sequence to be analyzed. This arrangement causes the nucleotides added via template-directed, DNA polymerase-catalyzed, extension of the primer to be determined unambiguously by the polymorphic nucleotide sequence in the template.

The above-described primer:template complex is contacted, under conditions of salt, pH, and temperature compatible with template-directed DNA synthesis, with a suitable DNA polymerase and four different chain-terminating nucleotide analogues known to form specific base pairs with the bases in the template. Most likely, but not necessarily, the bases in the template as well as the chain-terminating analogues are based on the common nucleosides: adenosine, cytosine, guanine or inosine, thymidine or uridine. A preferred set of chain-terminating analogues are the four dideoxynucleoside triphosphates, ddATP, ddCTP, ddGTP, and ddTTP, where each of the four ddNTPs has been modified by attachment of a different fluorescent reporter group. These fluorescent tags would have the property of having spectroscopically distinguishable emission spectra, and in no case would the dideoxynucleoside triphosphate modification render the chain-terminating analogue unsuitable for DNA polymerase-catalyzed incorporation onto primer 3'-termini. The result of DNA polymerase-catalyzed chain extension in such a mixture with such a primer:template complex is the quantitative, specific and unambiguous incorporation of a fluorescent chain-terminating analogue onto the 3'-terminus of the primer, the particular fluorescent nucleotide added being solely dictated by the sequence of the polymorphic nucleotides in the template.

The fluorescently-tagged primer:template complex, still attached to the magnetic microspheres, is then separated from the reaction mix containing the unincorporated nucleotides by, for example, washing the magnetically immobi-

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lized beads in a suitable buffer. Additionally, it is desirable in some circumstances to then elute the primer from the immobilized template strand with NaOH, transfer the eluted primer to a separate medium or container, and subsequently determine the identity of the incorporated terminator. The identity of the attached fluorescent group is then assessed by illuminating the modified DNA strand with light, preferably provided by a laser, of a suitable wavelength and intensity and spectrophotometrically analyzing the emission spectrum produced. In general, for a two allele (diploid) system at any given site in the DNA sequence, there are ten possible canonical emission spectra produced, corresponding to the sixteen possible homozygotic and heterozygotic pairings. By suitable matching of the measured spectra to this library of canonical spectra it is possible to identify which chain-terminating nucleotide(s) have been added to the 3'-terminus of the primer and thereby identify the nature of the sequence polymorphism in the template. Spectra produced by multiple allele systems or by alleles present in a ratio other than 1:1 can also be deconvolved by suitable mathematical treatments to identify and estimate the relative ratios of each contributing nucleotide.

All of the above steps involve chemistries, manipulations, and protocols that have been, or are amenable to being, automated. Thereby, incorporation of the preferred mode of practice of this invention into the operation of a suitably programmed robotic workstation should result in significant cost savings and increases in productivity for virtually any diagnostic procedure that depends on the detection of specific nucleotide sequences or sequence differences in nucleic acids derived from biological samples.

Several features of the above-described method have been improved and constitute a preferred embodiment of subject invention. Specifically, the preferred embodiment, Genetic Bit Analysis (GBA), presents a more convenient solid phase. Magnetic microspheres must be manipulated with care in order to effectively wash and resuspend them. It is therefore difficult to envisage high volume, automated assays using these beads. Furthermore, they are deeply colored and are not adapted to calorimetric or fluorescent assays.

The GBA methodology has been adapted to allow the utilization of standard, polystyrene, 96-well microplates. These have the advantage of being widely used in clinical and research laboratories. There are a large number of liquid

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handling systems, including automated systems, adapted to this format. They are suited to optical signal detection methods and automated plate readers for different types of light detection are available.

The template for GBA will always come from the nucleic acid sample of interest. These nucleic acids may be from a sample suspected of containing an infectious agent, one from an individual whose genotype is being determined, a sample from a patient suspected of having cancer, etc. If the immobilized partner of the hybrid complex to be extended is the template, each nucleic acid sample would have to be treated in such a way as to make immobilization possible. On the other hand, the primer for a given nucleic acid position to be interrogated will always be the same. Therefore, methods have been devised which allow the binding of the primer to the microplates and hybridization of single stranded template molecules to the plate-bound primer. This provides the additional feature of being able to make use of single-stranded templates produced in many different ways, including direct analysis of RNA.

Radioactive methods are inconvenient and produce waste-which is difficult to dispose of. For this reason, most commercial biochemistry detection systems have been converted to non-radioactive methods. By using ddNTPs which are labeled with biotin, GBA can be performed non-radioactively using a variety of detection systems including enzyme linked calorimetric assays.

Quality control is an important issue for tests designed to be used in clinical settings. Because GBA interrogates the nucleic acid sequence itself, on double stranded molecules, there is an opportunity to derive complementary genetic information by independently interrogating both strands. Applicants have shown that this approach is feasible using both equine and human genetic variants.

In the previously described method, the template was prepared by PCR using derivatized primers to permit immobilization of the template on the solid phase. Derivatization of the template is no longer necessary when the primer is immobilized. Rather, using unequal concentrations of PCR primers in an otherwise standard PCR, it is possible to generate an excess of one single-stranded molecule or the other, depending on which primer is in excess. These serve as convenient templates for hybridization to plate-bound GBA primer molecules.

## SEQUENCE LISTING

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17

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28

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29

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21

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aacgccaagg ctgggctctg ccagaccttt gtatatggt gctgcagagc taagagaaac 180
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| cttggcattc tcccctgaca ctcgccccaaag gcccctaacc tgcaccggg caccaggcac | 240 |
| cacgcggctcg gaggcttcag cagggaaggc ctgtctccg ttcacactgc tttcaggccc  | 300 |
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| cttggcattc tcccctgaca ctcgccccaaag gcccctaacc tgcaccggg caccaggcac     | 240 |
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&lt;213&gt; ORGANISM: Artificial Sequence

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&lt;223&gt; OTHER INFORMATION: Synthetic primer

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&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Synthetic primer

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agcctcagac cgcgtggc ctggc

25

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What is claimed is:

1. A reagent composition which comprises an aqueous carrier, a oligonucleotide primer molecule, a DNA polymerase, and an admixture of at least two different nucleotide terminators, or terminator nucleotide analogs, of a nucleic acid template dependent, primer extension reaction, each of the terminators being capable of specifically terminating the extension reaction and at least one of the terminators being labeled with a detectable marker, and wherein the reagent composition lacks dATP, dCTP, dGTP and dTTP.

2. A reagent of claim 1, wherein the reagent comprises four different terminators.

3. A reagent of claim 2, wherein two of the terminators are labeled, each with a different detectable marker.

4. A reagent of claim 2, wherein three of the terminators are labeled, each with a different detectable marker.

5. A reagent of claim 2, wherein the four terminators are labeled, each with a different detectable marker.

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6. A reagent of any of claim 1-5, wherein the terminators comprise dideoxynucleotides.

7. A reagent of claim 6, wherein the terminators comprise one or more of ddATP, ddCTP, ddGTP, ddTTP or ddUTP.

8. A reagent of any of claims 1-5, wherein the terminators comprise arabinoside triphosphates.

9. A reagent of any of claims 1-5, wherein each of the detectable markers is an isotopically labeled moiety, a chromophore, a fluorophore, a protein moiety, or a moiety to which an isotopically labeled moiety, a chromophore, a fluorophore, or a protein moiety can be attached.

10. A reagent of any of claims 3-5, wherein each of the different detectable markers is a different fluorophore.

11. A reagent of any of claims 1-5, wherein the reagent further comprises pyrophosphatase.

\* \* \* \* \*

## CIVIL COVER SHEET

The JS 44 civil cover sheet and the information contained herein neither replace nor supplement the filing and service of pleadings or other papers as required by law, except as provided by local rules of court. This form, approved by the Judicial Conference of the United States in September 1974, is required for the use of the Clerk of Court for the purpose of initiating the civil docket sheet. (SEE INSTRUCTIONS ON THE REVERSE OF THE FORM.)

## I. (a) PLAINTIFFS

Beckman Coulter Inc. and Orchid Cellmark, Inc.

## DEFENDANTS

Sequenom, Inc.

ORIGINAL

(b) County of Residence of First Listed Plaintiff Orange  
(EXCEPT IN U.S. PLAINTIFF CASES)County of Residence of First Listed Defendant San Diego  
(IN U.S. PLAINTIFF CASES ONLY)

PM 4:03

NOTE: IN LAND CONDEMNATION CASES, USE THE LOCATION OF THE LAND INVOLVED.

Attorneys (If known)

DEPUTY

BY:

'08 CV 1013 W POR

## II. BASIS OF JURISDICTION (Place an "X" in One Box Only)

|  |  |
|--|--|
| <input type="checkbox"/> 1 U.S. Government Plaintiff | <input checked="" type="checkbox"/> 3 Federal Question (U.S. Government Not a Party) |
| <input type="checkbox"/> 2 U.S. Government Defendant | <input type="checkbox"/> 4 Diversity (Indicate Citizenship of Parties in Item III)   |

## III. CITIZENSHIP OF PRINCIPAL PARTIES (Place an "X" in One Box for Plaintiff and One Box for Defendant)

| PTF  | DEF  | PTF                        | DEF                        |
|--|--|----------------------------|----------------------------|
| <input type="checkbox"/> 1 Citizen of This State                   | <input type="checkbox"/> 1 Incorporated or Principal Place of Business In This State     | <input type="checkbox"/> 4 | <input type="checkbox"/> 4 |
| <input type="checkbox"/> 2 Citizen of Another State                | <input type="checkbox"/> 2 Incorporated and Principal Place of Business In Another State | <input type="checkbox"/> 5 | <input type="checkbox"/> 5 |
| <input type="checkbox"/> 3 Citizen or Subject of a Foreign Country | <input type="checkbox"/> 3 Foreign Nation  | <input type="checkbox"/> 6 | <input type="checkbox"/> 6 |

## IV. NATURE OF SUIT (Place an "X" in One Box Only)

| CONTRACT   | TORTS  | FORFEITURE/PENALTY   | BANKRUPTCY   | OTHER STATUTES   |
|--|--|--|--|--|
| <input type="checkbox"/> 110 Insurance<br><input type="checkbox"/> 120 Marine<br><input type="checkbox"/> 130 Miller Act<br><input type="checkbox"/> 140 Negotiable Instrument<br><input type="checkbox"/> 150 Recovery of Overpayment & Enforcement of Judgment<br><input type="checkbox"/> 151 Medicare Act<br><input type="checkbox"/> 152 Recovery of Defaulted Student Loans (Excl. Veterans)<br><input type="checkbox"/> 153 Recovery of Overpayment of Veteran's Benefits<br><input type="checkbox"/> 160 Stockholders' Suits<br><input type="checkbox"/> 190 Other Contract<br><input type="checkbox"/> 195 Contract Product Liability<br><input type="checkbox"/> 196 Franchise | <b>PERSONAL INJURY</b><br><input type="checkbox"/> 310 Airplane<br><input type="checkbox"/> 315 Airplane Product Liability<br><input type="checkbox"/> 320 Assault, Libel & Slander<br><input type="checkbox"/> 330 Federal Employers' Liability<br><input type="checkbox"/> 340 Marine<br><input type="checkbox"/> 345 Marine Product Liability<br><input type="checkbox"/> 350 Motor Vehicle<br><input type="checkbox"/> 355 Motor Vehicle Product Liability<br><input type="checkbox"/> 360 Other Personal Injury | <b>PERSONAL INJURY</b><br><input type="checkbox"/> 362 Personal Injury—Med. Malpractice<br><input type="checkbox"/> 365 Personal Injury—Product Liability<br><input type="checkbox"/> 368 Asbestos Personal Injury Product Liability<br><input type="checkbox"/> 370 Other Fraud<br><input type="checkbox"/> 371 Truth in Lending<br><input type="checkbox"/> 380 Other Personal Property Damage<br><input type="checkbox"/> 385 Property Damage Product Liability | <input type="checkbox"/> 610 Agriculture<br><input type="checkbox"/> 620 Other Food & Drug<br><input type="checkbox"/> 625 Drug Related Seizure of Property 21 USC 881<br><input type="checkbox"/> 630 Liquor Laws<br><input type="checkbox"/> 640 R.R. & Truck<br><input type="checkbox"/> 650 Airline Regs.<br><input type="checkbox"/> 660 Occupational Safety/Health<br><input type="checkbox"/> 690 Other | <input type="checkbox"/> 422 Appeal 28 USC 158<br><input type="checkbox"/> 423 Withdrawal 28 USC 157<br><b>PROPERTY RIGHTS</b><br><input type="checkbox"/> 820 Copyrights<br><input checked="" type="checkbox"/> 830 Patent<br><input type="checkbox"/> 840 Trademark  |
| <input type="checkbox"/> 210 Land Condemnation<br><input type="checkbox"/> 220 Foreclosure<br><input type="checkbox"/> 230 Rent Lease & Ejectment<br><input type="checkbox"/> 240 Torts to Land<br><input type="checkbox"/> 245 Tort Product Liability<br><input type="checkbox"/> 290 All Other Real Property   | <b>REAL PROPERTY</b><br><input type="checkbox"/> 441 Voting<br><input type="checkbox"/> 442 Employment<br><input type="checkbox"/> 443 Housing/ Accommodations<br><input type="checkbox"/> 444 Welfare<br><input type="checkbox"/> 445 Amer. w/Disabilities - Employment<br><input type="checkbox"/> 446 Amer. w/Disabilities Other<br><input type="checkbox"/> 440 Other Civil Rights   | <b>CIVIL RIGHTS</b><br><input type="checkbox"/> 510 Motions to Vacate Sentence<br><b>Habeas Corpus:</b><br><input type="checkbox"/> 530 General<br><input type="checkbox"/> 535 Death Penalty<br><input type="checkbox"/> 540 Mandamus & Other<br><input type="checkbox"/> 550 Civil Rights<br><input type="checkbox"/> 555 Prison Condition   | <b>PRISONER PETITIONS</b><br><input type="checkbox"/> 710 Fair Labor Standards Act<br><input type="checkbox"/> 720 Labor/Mgmt. Relations<br><input type="checkbox"/> 730 Labor/Mgmt. Reporting & Disclosure Act<br><input type="checkbox"/> 740 Railway Labor Act<br><input type="checkbox"/> 790 Other Labor Litigation<br><input type="checkbox"/> 791 Empl. Ret. Inc. Security Act                          | <b>SOCIAL SECURITY</b><br><input type="checkbox"/> 861 HIA (1395ff)<br><input type="checkbox"/> 862 Black Lung (923)<br><input type="checkbox"/> 863 DIWC/DIWW (405(g))<br><input type="checkbox"/> 864 SSID Title XVI<br><input type="checkbox"/> 865 RSI (405(g))  |
|  |  |  | <b>IMMIGRATION</b><br><input type="checkbox"/> 462 Naturalization Application<br><input type="checkbox"/> 463 Habeas Corpus – Alien Detainee<br><input type="checkbox"/> 465 Other Immigration Actions   | <b>FEDERAL TAX SUITS</b><br><input type="checkbox"/> 870 Taxes (U.S. Plaintiff or Defendant)<br><input type="checkbox"/> 871 IRS—Third Party 26 USC 7609<br><b>OTHER STATUTES</b><br><input type="checkbox"/> 875 Customer Challenge 12 USC 3410<br><input type="checkbox"/> 890 Other Statutory Actions<br><input type="checkbox"/> 891 Agricultural Acts<br><input type="checkbox"/> 892 Economic Stabilization Act<br><input type="checkbox"/> 893 Environmental Matters<br><input type="checkbox"/> 894 Energy Allocation Act<br><input type="checkbox"/> 895 Freedom of Information Act<br><input type="checkbox"/> 900 Appeal of Fee Determination Under Equal Access to Justice<br><input type="checkbox"/> 950 Constitutionality of State Statutes |

## V. ORIGIN (Place an "X" in One Box Only)

|   |   |  |
|---|---|--|
| <input checked="" type="checkbox"/> 1 Original Proceeding | <input type="checkbox"/> 2 Removed from State Court | <input type="checkbox"/> 3 Remanded from Appellate Court |
|---|---|--|

Transferred from

|   |   |
|---|---|
| <input type="checkbox"/> 4 Reinstated or Reopened | <input type="checkbox"/> 5 another district (specify) |
|---|---|

|   |   |
|---|---|
| <input type="checkbox"/> 6 Multidistrict Litigation | <input type="checkbox"/> 7 Judge from Magistrate Judgment |
|---|---|

Appeal to District

Cite the U.S. Civil Statute under which you are filing (Do not cite jurisdictional statutes unless diversity):

35 U.S.C. § 101, et seq.

Brief description of cause:

Patent Infringement

## VI. CAUSE OF ACTION

(See instructions):

JUDGE \_\_\_\_\_

DEMAND \$

CHECK YES only if demanded in complaint:

JURY DEMAND:  Yes  No

|   |   |
|---|---|
| DATE<br>June 5, 2008                          | SIGNATURE OF ATTORNEY OF RECORD<br><i>J. S. mtakes</i>  |
| FOR OFFICE USE ONLY<br>RECEIPT # <u>15169</u> | AMOUNT <u>\$ 350</u><br>APPLYING IFF <u>      </u><br>JUDGE <u>      </u><br>MAG. JUDGE <u>      </u> |

## INSTRUCTIONS FOR ATTORNEYS COMPLETING CIVIL COVER SHEET FORM JS 44

## Authority For Civil Cover Sheet

The JS 44 civil cover sheet and the information contained herein neither replaces nor supplements the filings and service of pleading or other papers as required by law, except as provided by local rules of court. This form, approved by the Judicial Conference of the United States in September 1974, is required for the use of the Clerk of Court for the purpose of initiating the civil docket sheet. Consequently, a civil cover sheet is submitted to the Clerk of Court for each civil complaint filed. The attorney filing a case should complete the form as follows:

**I. (a) Plaintiffs-Defendants.** Enter names (last, first, middle initial) of plaintiff and defendant. If the plaintiff or defendant is a government agency, use only the full name or standard abbreviations. If the plaintiff or defendant is an official within a government agency, identify first the agency and then the official, giving both name and title.

(b) County of Residence. For each civil case filed, except U.S. plaintiff cases, enter the name of the county where the first listed plaintiff resides at the time of filing. In U.S. plaintiff cases, enter the name of the county in which the first listed defendant resides at the time of filing. (NOTE: In land condemnation cases, the county of residence of the "defendant" is the location of the tract of land involved.)

(c) Attorneys. Enter the firm name, address, telephone number, and attorney of record. If there are several attorneys, list them on an attachment, noting in this section "(see attachment)".

**II. Jurisdiction.** The basis of jurisdiction is set forth under Rule 8(a), F.R.C.P., which requires that jurisdictions be shown in pleadings. Place an "X" in one of the boxes. If there is more than one basis of jurisdiction, precedence is given in the order shown below.

United States plaintiff. (1) Jurisdiction based on 28 U.S.C. 1345 and 1348. Suits by agencies and officers of the United States are included here.

United States defendant. (2) When the plaintiff is suing the United States, its officers or agencies, place an "X" in this box.

Federal question. (3) This refers to suits under 28 U.S.C. 1331, where jurisdiction arises under the Constitution of the United States, an amendment to the Constitution, an act of Congress or a treaty of the United States. In cases where the U.S. is a party, the U.S. plaintiff or defendant code takes precedence, and box 1 or 2 should be marked.

Diversity of citizenship. (4) This refers to suits under 28 U.S.C. 1332, where parties are citizens of different states. When Box 4 is checked, the citizenship of the different parties must be checked. (See Section III below; federal question actions take precedence over diversity cases.)

**III. Residence (citizenship) of Principal Parties.** This section of the JS 44 is to be completed if diversity of citizenship was indicated above. Mark this section for each principal party.

**IV. Nature of Suit.** Place an "X" in the appropriate box. If the nature of suit cannot be determined, be sure the cause of action, in Section VI below, is sufficient to enable the deputy clerk or the statistical clerks in the Administrative Office to determine the nature of suit. If the cause fits more than one nature of suit, select the most definitive.

**V. Origin.** Place an "X" in one of the seven boxes.

Original Proceedings. (1) Cases which originate in the United States district courts.

Removed from State Court. (2) Proceedings initiated in state courts may be removed to the district courts under Title 28 U.S.C., Section 1441. When the petition for removal is granted, check this box.

Remanded from Appellate Court. (3) Check this box for cases remanded to the district court for further action. Use the date of remand as the filing date.

Reinstated or Reopened. (4) Check this box for cases reinstated or reopened in the district court. Use the reopening date as the filing date.

Transferred from Another District. (5) For cases transferred under Title 28 U.S.C. Section 1404(a). Do not use this for within district transfers or multidistrict litigation transfers.

Multidistrict Litigation. (6) Check this box when a multidistrict case is transferred into the district under authority of Title 28 U.S.C. Section 1407. When this box is checked, do not check (5) above.

Appeal to District Judge from Magistrate Judgment. (7) Check this box for an appeal from a magistrate judge's decision.

**VI. Cause of Action.** Report the civil statute directly related to the cause of action and give a brief description of the cause. **Do not cite jurisdictional statutes unless diversity.** Example: U.S. Civil Statute: 47 USC 553  
Brief Description: Unauthorized reception of cable service

**VII. Requested in Complaint.** Class Action. Place an "X" in this box if you are filing a class action under Rule 23, F.R.Cv.P.

Demand. In this space enter the dollar amount (in thousands of dollars) being demanded or indicate other demand such as a preliminary injunction.

Jury Demand. Check the appropriate box to indicate whether or not a jury is being demanded.

**VIII. Related Cases.** This section of the JS 44 is used to reference related pending cases if any. If there are related pending cases, insert the docket numbers and the corresponding judge names for such cases.

**Date and Attorney Signature.** Date and sign the civil cover sheet.

UNITED STATES  
DISTRICT COURT  
SOUTHERN DISTRICT OF CALIFORNIA  
SAN DIEGO DIVISION

# 151669 - TC  
\* \* C O P Y \* \*  
June 05, 2008  
16:08:16

**Civ Fil Non-Pris**

USAO #: 08CV1013  
Judge.: THOMAS J WHELAN  
Amount.: \$350.00 CK  
Check#: BC16235

**Total→ \$350.00**

FROM: BECKMAN COULTER INC. AND  
ORCHID CELLMARK, INC.  
VS.  
SEQUENOM